The Effect of Temperature and Genetic Background on Host Gene Expression and Interaction to Puccinia graminis tritici

N. H. Luig and S. Rajaram

Senior Research Fellow and Postgraduate Student, respectively, Department of Agricultural Botany, University of Sydney, New South Wales, Australia 2006. Present address of second author: CIMMYT, Londres 40, Mexico 6, D.F., Mexico.

The authors acknowledge the interest and help of I. A. Watson, who originally suggested this problem. Financial assistance from the Namoi FAO Research Foundation and the Rotary Club of Narrabri, New South Wales, and from the Wheat Industry Research Council of Australia is also gratefully acknowledged.

Accepted for publication 7 March 1972.

ABSTRACT

Infection types produced by *Puccinia graminis* f. sp. tritici on seedlings of near-isogenic lines of Marquis wheat, and on homozygous and heterozygous combinations between them, were studied in growth cabinets at six temperature levels. At the high temperatures, 27 and 30 C, seedlings heterozygous or homozygous for the genes Sr5, Sr8, and Sr9b became susceptible or semisusceptible.

When Sr5 and Sr9b were transferred by successive backcrosses to the highly susceptible background of

W2691 and W3498, the infection types became much higher, and heterozygous seedlings were semisusceptible at the intermediate temperature range.

In most instances, combinations of host genes resulted in higher levels of resistance. Higher temperatures and susceptible genetic backgrounds interacted in decreasing the resistance provided by all genes.

Phytopathology 62:1171-1174.

Additional key words: gene combinations.

While it is well known that high temperatures can alter infection types caused by wheat stem rust, *Puccinia graminis* Pers. f. sp. *tritici* Eriks. & E. Henn., towards susceptibility in instances where temperature-sensitive genes for resistance are involved (3, 4, 5, 8, 10), low temperatures can have the same effect (14). Low light intensity and chemicals can also render certain genes for resistance ineffective (2, 3, 5, 7, 8).

Knott & Green (9) reported that the gene action of Sr6, Sr8, Sr9a, and Sr9b was not modified to any extent by other genes present in the cultivars which they used. However, when Sr10 and Sr7 were backcrossed to Marquis more than 3 and 6 times, respectively, a distinct loss of resistance occurred.

Watson & Luig (21) demonstrated that the genes Sr6 and Sr11 each were capable of interactions resulting in low infection types at three different levels of expression. Thus, in a given environment, avirulence of rust cultures could be further characterized according to the infection types induced by these two genes. Furthermore, it was found that the expression and the dominance relationship of Sr6 were greatly altered by changes in temperature (10). The present study, utilizing Sr6 and five genes previously considered temperature-insensitive, reports on (i) interactions between these genes when present in pairs either in a heterozygous or homozygous condition; (ii) influence of constant temperatures on infection type; and (iii) effects of the genetic background on gene expression.

MATERIALS AND METHODS.—Six near-isogenic lines in a Marquis background, originally provided by D. R. Knott, Saskatoon, Sask., Canada, namely, Kenya 58 X Mq⁶ W2400 (Sr6), Mq⁶ X Red Egyptian W2401 (Sr8), Mq⁶ X Kenya 117A W2402 (Sr9b),

Thatcher \times Mq⁶ W2928 (Sr₅), Khapstein \times Mq⁶ W2931 (Sr₁3), and Lee \times Mq¹ W3015 (Sr₁11), were used to synthesize lines having different paired combinations for low reaction to stem rust. A diallele set of crosses between the six lines was made. Approximately 25 to 35 seedlings raised from 300 F₃ lines of each cross were inoculated with appropriate strains of P. graminis tritici virulent on one of the parents of each cross. Residual seed of homozygous-resistant lines were sown, and at least 20 seedlings were inoculated with strains virulent on the second parent. Again, lines homozygous-resistant were chosen, and these were further tested with strain 34-1,2,3,6,7 which is virulent on plants with Sr5, Sr6, Sr8, Sr9b and Sr11. Except for lines possessing the Sr13 allele, the resulting susceptible infection types were additional evidence for the presence of the genes derived from the near-isogenic parents.

By the above procedures, the homozygous combinations (i) Sr5, Sr9b (21-1,2,3,7, 34-2,4,5); (ii) Sr5, Sr13 (21-1,2,3,7, 126-1,6,7); (iii) Sr6, Sr8 (126-6,7, 21-1,2,3,7); and (iv) Sr8, Sr9b (126-1,6,7 21-1,2,3,7), were obtained. The strains used for inoculation which have been described earlier (20) are indicated in parentheses.

In addition to the four homozygous combinations, four double heterozygous combinations (the F_1 from crossing two isogenic lines), five single heterozygotes (the F_1 from crossing an isogenic line with Marquis), and the six isogenic parents were tested. Marquis (as the susceptible check), Reliance (Sr5), and Kanred (Sr5) were used for purposes of comparison.

Two very susceptible lines, W2691 and W3498, which have been described (12) were used as the recurrent parents in a program designed to obtain nearisogenic lines possessing major genes for resistance in

a susceptible background. Three lines (A-Sr5, B-Sr9b, C-Sr11) resulting from this program, as well as W2691 and W3498, were included in the study.

Strain 21-4,5, which resembles strain 21-2 in interactions with Sr6 (21) and which is also avirulent on the other isogenic lines (20), was chosen. After inoculation, seedlings (10-20 in each line) were transferred to "Sherer" plant growth rooms where temperatures of 15, 18, 21, 24, 27, and 30 C with an average fluctuation of \pm 1 C were maintained. Light intensity was ca. 3,000 ft-c. Infection types were recorded after 12 days. The system of Stakman et al. (18) was followed, excepting a "3+" denoting full susceptibility. Backcross lines involving W2691 and W3498 were tested mainly in the greenhouse at 15 to 24 C.

RESULTS.—Infection types are listed in Table 1. Seedlings of W2928 (Sr5 Sr5) exhibited infection type "O-O;" at 15, 18, 21, and 24 C, "O; 1=" at 27 C, and "3cn" at 30 C, whereas F₁ plants of the cross W2928 X Marquis (Sr5 sr5) produced "O-O;" at the 4 lower temperatures, ";1=" at 27 C, and "3" at 30 C. These results indicated that Sr5 in a Marquis background was completely dominant at the lower temperatures, partially dominant at 27 C, and recessive at 30 C. However, at the last temperature, Sr5 when present in W2928 was largely ineffective, whereas seedlings of Kanred (Sr5) and Reliance (Sr5), remained fully resistant ("O" and "O;" infection type).

The temperature sensitivity of Sr5 at 30 C was confirmed in the tests involving homozygous and heterozygous combinations with the genes Sr9b and Sr13 (Table 1). Since infection types on seedlings having the gene combinations were lower than those on seedlings possessing Sr5 alone, the data suggest additive gene interaction. This was surprising in the case of the heterozygous combination Sr5 Sr9b Sr9b, as the heterozygote Sr9b Sr9b appeared to be fully susceptible at 30 C.

Lines of Sr5 in a W2691-W3498 background were tested against strain 21-4,5 in the glasshouse at average temperatures of 21 C. Those homozygous and heterozygous for Sr5, and backcrossed to W2691 and W3498 five times, exhibited ";" and "X=" infection types, respectively. The same genotypes after seven backcrosses showed "X" and "3c"; and the infection types recorded after nine backcrosses were "X+3" and "3". Thus, by changing the genetic background, it was possible to achieve a partial breakdown of the Sr5 resistance similar to that occurring at constant temperatures of 30 C. The infection types recorded in the growth chambers on a homozygous line synthesized after nine backcrosses are shown in Table 1.

As can be seen in the second part of Table 1, the gene Sr6, when homozygous in a Marquis background, was ineffective at 24 C and above, whereas the heterozygous F_1 plants were susceptible at all six temperatures. In the glasshouse, plants with Sr6 were much more resistant to strain 21-4,5, especially at temperatures of 15 to 18 C when this gene is dominant (10). Better light conditions in the glasshouse were probably responsible for this difference. Sr8 when heterozygous was ineffective at high temperatures. The homozygous and heterozygous combina-

tions between Sr6 and Sr8 produced lower infection types than those with the two components singly, again indicating additive gene interaction.

Gene interaction was also observed in the homozygous and heterozygous combinations between Sr8 and Sr9b. At 30 C, the two single heterozygotes were susceptible, whereas the double heterozygote exhibited "2+3-" infection types.

The expression of the gene *Sr11* when homozygous was less influenced by genetic background than that of *Sr9b*. *Sr11* was also less affected by high temperatures, and by the temperature-background interactions (Table 1).

DISCUSSION.—Host genes conditioning resistance to *P. graminis tritici* can be grouped according to the stability of their expression under different environmental conditions. Extreme cases of thermal sensitivity are genes *Sr6* (10) and *Sr15* (20), whereas five genes (*Sr5*, *Sr8*, *Sr9b*, *Sr11*, and *Sr13*) condition similar infection types at temperatures normally encountered in our glasshouses. However, at constant temperatures of 30 C, *Sr5* in W2928 (Thatcher X Marquis⁶) conditioned a semisusceptible reaction only. This is the first report of thermal sensitivity concerning this gene. Aamodt (1) found *Sr5* to be completely dominant.

At the same constant temperature, Kanred (Sr5) and Reliance (Sr5) exhibited their characteristic immunity ("O" and "O;" infection type). This shows that high temperatures are unable to influence the resistant reaction expressed by Sr5 in the genetic background of these two cultivars.

Furthermore, it was shown that when Sr5 was transferred by several backcrosses to the "susceptible" genetic background of W2691 and W3498, it became largely ineffective. This clearly demonstrates that susceptible wheat cultivars could influence different segregation patterns in crosses with resistant cultivars possessing one or more major genes for resistance. Even in a favorable environment, the expression of such genes would be determined by the genetic background.

W2691 and W3498 were selected for susceptibility to rye stem rust, Puccinia graminis f. sp. secalis, and Sanghi & Luig (15) have shown that the resistance to this pathogen in wheat cultivars susceptible to wheat stem rust is due to many previously unknown genes. Since W2691 and W3498 are not only susceptible to certain strains of P. graminis secalis but also provide a susceptible genetic background for resistance genes to P. graminis tritici, it is suggested that the genes for resistance to P. graminis secalis act as modifiers of genes for resistance to P. graminis tritici. The effect of the genes for resistance to rye stem rust on reducing the spread of wheat stem rust in the field on cultivars which possess no factor for resistance to this pathogen has not been determined; however, several independent observations suggest that these may constitute one kind of nonspecific resistance as defined by Watson (19) or van der Plank's horizontal resistance (13).

In many host-pathogen interactions that showed temperature sensitivity, mesothetic infection types

TABLE 1. Infection types on homozygous and heterozygous lines and three cultivars of wheat when tested with strain 21-4,5 of *Puccinia graminis tritici* at six constant temperatures under artificial light

Genotypes	Temperature ± 1 C					
	15	18	21	24	27	30
Marquis	3+ca	3+c	3+c	3+c	3+	3+
Kanred (Sr5 Sr5)	0	0	0	0	0	0
Reliance (Sr5 Sr5)	0	0	0	0;	0;	0;
Sr5 Sr5 (W2928)	0-0;	0	0-0;	0-0;	0;1=	3cn
Sr5 sr5	0-0;	0	0	0;	;1=	3
Sr9b Sr9b (W2402)	23c	2-3c	2-3c	3-c	2+3-c	2+3c
Sr9b sr9b	2+3-	2+3+c	2+3+c	3+c	3+c	3+
Sr5 Sr5 Sr9b Sr9b	0	0	0	0	0	;2=
Sr5 sr5 Sr9b sr9b	0	0	0;	0;	;1=	X=
Sr13 Sr13 (W2931)	2	2-	2=2	2=2	2-	2
Sr5 Sr5 Sr13 Sr13	0	0	0	0	0;	;1-
Sr5 sr5 Sr13 sr13	0	0	0;	0		
Sr6 Sr6 (W2400)	;	1+3-cn	3cn	3+	3+	3+
Sr6 sr6	33+	33+	3+	3+	3+	3+
Sr8 Sr8 (W2401)	2-2	2-2	2-2	2-2	2-2	2
Sr8 sr8		22+	2+3+	2+3+	3+	3+
Sr6 Sr6 Sr8 Sr8	0;	;1-2-	2-	2=2-	2-	
Sr6 sr6 Sr8 sr8	2-2	2+3c	2+3c	2+3c	2+3c	3c3+
Sr8 Sr8 Sr9b Sr9b	2-	2-	2=2-	2=2-	2=2-	2-
Sr8 sr8 Sr9b sr9b	22+	22+	22+	2+3=	2+3-	2+3-
Sr11 Sr11 (W3015)	2≡2=	2≡2=	2=2=	2≡2=	2≡2=	2=2-
Sr11 sr11	2=2-	2≡2=	2≡2=	2==2=	2=2-	
W2691	3+	3+	3+	3+	3+	3+
W3498	3+	3+	3+	3+	3+	3+
Line A Sr5 Sr5	X+3-	X+3-	X+3	X+3	X+3	3
Line B Sr9b Sr9b	2+	2+	2+	2+3-	2+3-	3-
Line C Sr11 Sr11	2≡2=	2=	2=	2=	2=	2-

^aInfection types are according to Stakman et al. (18). 0 = immune; 0; = nearly immune; 1 = very resistant; 2 = moderately resistant; 3 = susceptible; X = heterogeneous; $\equiv \text{and} = \text{indicate lower limits of type}$; + and - indicate variation within a given infection type; + c = chlorosis; + b = browning.

have been exhibited (6, 10). In this regard, Sr5 resembles Sr6 and Sr15. Although, at the average normal temperature range experienced in our glasshouse (21-24 C), the infection types of the three genes when tested with strain 21-4,5 are quite different ("O", ";1++", and "X+3"), it now seems that these different phenotypes are due to different temperature requirements of the three genes. Thus, a labile mesothetic "X" infection type can be produced by all three genes; namely, by Sr5 at constant 27-30 C, by Sr6 at 18 C, and by Sr15 at 15 C. Slight variations in temperature greatly influence the amount of necrosis produced in the three host-pathogen combinations, and this in turn changes the infection types. On the other hand, our studies suggest that the chlorotic "2" infection type conditioned by Sr8, Sr9b, Sr11, and Sr13 is less influenced by high temperature. However, plants heterozygous for certain of these were rendered susceptible by the highest temperatures.

Additive gene interaction was found in a number of instances, especially when *Sr6* was involved. Such nonallelic interaction was described earlier (16). Since Marquis did not provide a fully susceptible back-

ground (Table 1), it was difficult to identify the effects of individual genes and to study interactions between them. To minimize the interference of the genetic background, near-isogenic lines in a susceptible background will have to be developed and employed in a diallel crossing program. At present, many genes have been transferred by backcrossing to W3498, and these lines should prove valuable for future studies on gene interaction.

The findings reported here might be applied in combining two resistance genes when the three appropriate cultures of the pathogen are unavailable. High temperatures could be used to inactivate one while selection is practized for the other. Simons (17) distinguished genetically different sources of oat crown rust resistance using environmental modifications such as high temperature and high nitrogen fertilization. Green & Johnson (4) separated foundation stock lines of Selkirk wheat employing the temperature sensitivity of Sr6. Under field conditions, cultivars having two or more genes conditioning moderate resistance normally exhibit a higher level of resistance to avirulent strains than is shown by those having the same genes present singly (11).

LITERATURE CITED

- AAMODT, O. S. 1923. The inheritance of growth habit and resistance to stem rust in a cross between two varieties of common wheat. J. Agr. Res. 24:457-470.
- DALY, J. M., P. M. SEEVERS, & P. LUDDEN. 1970. Studies on wheat stem rust resistance controlled at the Sr6 locus. III. Ethylene and disease reaction. Phytopathology 60:1648-1652.
- 3. FORSYTH, F. R. 1956. Interaction of temperature and light on the seedling reaction of McMurachy wheat to race 15B of stem rust. Can. J. Bot. 34:745-749.
- GREEN, G. J., & T. JOHNSON. 1955. The reaction of foundation stock lines of Selkirk wheat to some physiologic races of wheat stem rust. Can. J. Agr. Sci. 35:323-328.
- HART, H., & K. ZALESKI. 1935. The effect of light intensity and temperature on infection of Hope wheat by Puccinia graminis tritici. Phytopathology 25:1041-1066.
- JOHNSON, L. B., & J. F. SCHAFER. 1965. Identification of wheat leaf rust resistance combinations by differential temperature effects. Plant Dis. Reptr. 49:222-224.
- JOHNSON, T. 1946. The effect of DDT on the stem rust reaction of Khapli wheat. Can. J. Res. Sect. C 24:23-25.
- 8. JOHNSON, T., & M. NEWTON. 1940. The influence of light and certain other environmental factors on the mature plant resistance of Hope wheat to stem rust. Can. J. Res. Sect. C 18:357-371.
- 9. KNOTT, D. R., & G. J. GREEN. 1965. A comparison of the reactions to stem rust of wheat lines backcrossed five and nine times to Marquis that carry the same resistance genes. Can. J. Plant Sci. 45:106-107.
- LUIG, N. H., & I. A. WATSON. 1965. Studies on the genetic nature of resistance to Puccinia graminis var. tritici in six varieites of common wheat. Linneaus Soc.

- New South Wales Proc. 90:299-327.
- LUIG, N. H., & I. A. WATSON. 1970. The effect of complex genetic resistance in wheat on the variability of Puccinia graminis f. sp. tritici. Linnaeus Soc. New South Wales Proc. 95:22-45.
- LUIG, N. H., & I. A. WATSON. 1972. The role of wild and cultivated grasses in the hybridisation of formae speciales of Puccinia graminis. Australian J. Biol. Sci. 25:335-342.
- PLANK, VAN DER, J. E. 1968. Disease resistance in plants. Academic Press, New York. 206 p.
- RAJARAM, S., N. H. LUIG, & I. A. WATSON. 1971.
 The inheritance of leaf rust resistance in four varities of common wheat. Euphytica 20:574-585.
- SANGHI, A. K., & N. H. LUIG. 1971. Resistance in wheat to formae speciales tritici and secalis of Puccinia graminis. Can. J. Genet. Cytol. 13:119-127.
- SCHAFER, J. F., R. M. CALDWELL, F. L. PATTER-SON, & L. E. COMPTON. 1963. Wheat leaf rust resistance combinations. Phytopathology 53:569-573.
- 17. SIMONS, M. D. 1955. The use of pathological techniques to distinguish genetically different sources of resistance to crown rust of oats. Phytopathology 45:410-413.
- STAKMAN, E. C., D. M. STEWART, & W. Q. LOEGER-ING. 1962. Identification of physiologic races of Puccinia graminis var. tritici. U.S. Dep. Agr., Agr. Res. Service E617.
- WATSON, I. A. 1970. Changes in virulence and population shifts in plant pathogens. Annu. Rev. Phytopathol. 8:209-230.
- WATSON, I. A., & N. H. LUIG. 1966. Sr15-a new gene for use in the classification of Puccinia graminis var. tritici. Euphytica 15:239-250.
- WATSON, I. A., & N. H. LUIG. 1968. Progressive increase in virulence in Puccinia graminis f. sp. tritici. Phytopathology 58:70-73.