

For the Record

# Structure-Based Multiple Alignment of Extracellular Pectate Lyase Sequences

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Received 26 August 1994. Accepted 8 December 1994.

**Pectate lyases are secreted virulence factors which degrade the pectate component of plant cell walls. The evolutionary-based multiple alignment of extracellular pectate lyases has been corrected using three-dimensional structural information derived from *Erwinia chrysanthemi* pectate lyases C and E. The new multiple alignment reveals invariant amino acids likely to be involved in two different enzymatic functions.**

*Additional keywords:* pectate lyase sequence alignment.

Pectate lyases (EC 4.2.2.2) are depolymerizing enzymes which degrade the pectate component of the plant cell wall, leading to tissue maceration and death. The most-studied enzymes are secreted from the phytopathogenic *Erwinia* species and are the primary virulence factors in 'soft-rot' diseases (Collmer and Keen 1986; Kotoujansky 1987). Sequence studies have identified four distinct families: the extracellular *pelADE* family; the extracellular *pelBC* family; the periplasmic pectate lyases from *Yersinia pseudotuberculosis* and *Erwinia carotovora* strains; and some fungal pectate lyases with a small molecular weight of 24 kDa (Gonzalez-Candelas and Kolattukudy 1992). Within each family, the genes share more than 50% sequence similarity, but only the *pelBC* and *pelADE* subfamilies appear to be evolutionarily related. In the latter subfamilies, three conserved sequence patterns have been identified which have been used to characterize a new DNA sequence as belonging to the pectate lyase family. The patterns include AxDIKGxxxxVTxS, VxxRxPxxRxGxxHxxxN, and vWiDH (Tamaki et al. 1988; Hinton et al. 1989; Hugouvioux-Cotte-Patat et al. 1992; Barras et al. 1994). The extracellular pectate lyases also share the patterns with pectin lyases (Gysler et al. 1990; Kuster-van Someren et al. 1992), plant pollen proteins (Wing et al. 1989; Rafnar et al. 1991), and plant style proteins (Budelier et al. 1990), but none of the latter proteins has yet been found to possess in vitro pectate lyase activity.

The three-dimensional structures of three pectate lyases have recently been determined: *E. chrysanthemi* pectate lyase C (PelC) (Yoder et al. 1993); *E. chrysanthemi* pectate lyase E

(PelE) (Lietzke et al. 1994); and *Bacillus subtilis* pectate lyase (Pickersgill et al. 1994). All reveal that the enzymes have an unusual structure, that of parallel  $\beta$  strands folded into a large helix. Because the protein fold is novel, various studies have been undertaken to understand the unique features. The present study involves a superposition of the atomic coordinates for PelC and PelE using a least-square fit between  $\alpha$ Cs in the parallel  $\beta$  helix domain. The PILEUP multiple sequence alignment program of the Genetics Computer Group package has been used to align the sequences of other extracellular pectate lyases to the structurally aligned sequences of PelC and PelE. The multiple alignment of 14 extracellular pectate lyases is shown in Table 1. Of the 353 amino acids in PelC, 49 are invariant, using only evolutionary relationships (Hinton et al. 1989), but a different set of only 27 is invariant, using the structural information. An analysis of the discrepancies indicates that, in addition to new sequence information, those alignments which relied, in part (Barras et al. 1994) or in total (Hinton et al. 1989), on evolutionary relationships contain misplaced gaps in sequences in loop regions of PelC and PelE, but correctly align most sequences within the parallel  $\beta$  helix. As a consequence, the evolutionary-based multiple sequence alignments of pectate lyases and their homologues are, at best, 70% correct.

The corrected multiple sequence alignment reveals several new features in extracellular pectate lyases. Of the 27 invariant amino acids, 14 possess chemical properties that are compatible with catalysis (Zvelebil and Sternberg 1988). In PelC, these include T10, T92, D131, D144, H145, D170, T179, S181, K190, T206, R218, R223, H28, and K342. On the three-dimensional structures of PelC and PelE, the 'potentially catalytic' amino acids cluster into two widely separated regions, suggesting that there are two active sites. The first group, D131, D170, K190, R218, and R223, are found near the  $\text{Ca}^{2+}$  binding site on the protein, with the two invariant aspartic acids coordinating directly to  $\text{Ca}^{2+}$  (Pickersgill et al. 1994). Because  $\text{Ca}^{2+}$  is essential for in vitro catalytic activity, the region around the  $\text{Ca}^{2+}$  site is believed to be the pectinolytic active site (Yoder et al. 1993; Pickersgill et al. 1994). The remaining 'potentially catalytic' invariant amino acids are widely separated in the primary sequence but cluster spatially in a region surrounding the vWiDH sequence. Interestingly, the cluster includes, not only the

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**Table 1.** Multiple alignment of pectate lyase sequence<sup>a</sup>

Seq. Name											Res #			
	β <sub>N-Terminal</sub>													
				Branch										
				β										
				Helix										
Pelc_Erwch	ATDT			GGYAA	..	TAGGNV	VTG	AV	..	SKT	ATSMQ	DIVNI	32	
Pelb_Erwch	.ADT			GGYTK	..	TDGGDV	SVG	AV	..	KKT	ASSMQ	DIVNI	31	
Pel2_Erwca	.ANT			GGYAT	..	TDGGEV	SVG	AV	..	KKT	ARSMKE	IVDI	31	
Pel3_Erwca	.ANT			GGYAT	..	TDGGDV	AVG	AV	..	KKT	ARSMQDI	IIDI	31	
Pela_Erwca	.ANT			GGYAT	..	TDGGDV	SVG	AV	..	KKT	ARSLQE	IINL	31	
Pelb_Erwca	.ANT			GGYAT	..	TDGGDV	SVG	AV	..	KKT	ARSLQE	IVDI	31	
Pelc_Erwca	.ANT			GGYAT	..	TDGGDV	AVG	AV	..	KKT	ARSMQDI	IIDI	31	
Pela_Erwch	..	AELVSD	KALESAPT	..	..	VGWASQN	GFTTGGAA	..	..	ATSDNIYI	VTNISEFT	SA	47	
Peld_Erwch	..	TLQTT	KATEAA.S	..	..	TGWATQ	GTTGGAK	..	..	AASAKIYA	VKNISEFK	AA	44	
Pele_Erwch	..	..	A VETDAA.T	..	..	TGWATQN	GTTGGAK	..	..	AAKAVE	VKNISDFK	KA	39	
Pelf_Erwch	..	ASLQTT	KATEAA.S	..	..	TGWATQS	GTTGGAK	..	..	ASSSKIYA	VKSISEFK	AA	46	
Pel_Bacsub	..	..	AD LGHQTLGSN	..	..	DGWAYS	TGTTGGSK	..	..	ASSSNVYT	VSNRNQLV	SA	44	
Pel_Psemar	..	..	..	..	..	TGWATQN	GGTKGSR	..	..	AAANDIYT	VKNAAELK	KA	42	
Pela_Emnid	..	..	..	..	..	AR HELTRRQASE	SCPIGYCTQN	GGTTGGAA	..	..	GDTVTT	VTNLADL	TEA	45
Pelc_Erwch	..	IDAARLDANG	KVKVGGA	..	..	YPLVITYT	GNEDSLINAA	AANICGQWSK	DPRG	..	..	..	81	
Pelb_Erwch	..	IEAAKVDANG	KVKVGGA	..	..	YPLVITYT	GNEDSLINAA	AANICGQWSK	DARG	..	..	..	80	
Pel2_Erwca	..	IEAAQVDSKG	KVKVGGA	..	..	YPLIITYS	GNEDSLIKAA	EKNICGQWSK	DARG	..	..	..	80	
Pel3_Erwca	..	IEAAKLDNSG	KVKVGGA	..	..	YPLVITYN	GNEDALIKAA	EANICGQWSK	DARG	..	..	..	80	
Pela_Erwca	..	IEEAQLDSKG	KKLKGA	..	..	YPLVITYN	GNEDALIKAA	EANICGQWSK	DPRG	..	..	..	80	
Pelb_Erwca	..	IEAAKVDNSG	KVKVGGA	..	..	YPLIITYN	GNEDSLIKAA	EKNICGQWSK	DARG	..	..	..	80	
Pelc_Erwca	..	IEAAKLDNSG	KVKVGGA	..	..	YPLVITYN	GNEDALIKAA	ENDICGQWKK	DARG	..	..	..	80	
Pela_Erwch	..	LS	..	..	..	A GAEAKIIQIK	GT	..	..	IDISGG	TPYTFDFADQK	ARSQ	82	
Peld_Erwch	..	LN	..	..	..	GT DTDPKIIQVT	GA	..	..	IDISGG	KAYTSFDDQK	ARSQ	80	
Pele_Erwch	..	LN	..	..	..	GT DSSAKIIKVT	GP	..	..	IDISGG	KAYTSFDDQK	ARSQ	75	
Pelf_Erwch	..	LN	..	..	..	GT DSSPKIIQVT	GA	..	..	IDISGG	KAYTSFDDQK	ARSQ	82	
Pel_Bacsub	..	LG	..	..	..	KET NTTPKIIYIK	GT	..	..	..	IDMNV	DNLKPLGLND	77	
Pel_Psemar	..	LS	..	..	..	ASA GSNRIIKIT	GI	..	..	..	IDVSEG	KVYTKTADMK	VRGR	79
Pela_Emnid	..	AE	..	..	..	SDGPLTIIVS	GS	..	..	..	..	..	59	
Pelc_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	113	
Pelb_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pel2_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pel3_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pela_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pelb_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pelc_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	112	
Pela_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	116	
Peld_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	111	
Pele_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	106	
Pelf_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	113	
Pel_Bacsub	..	..	..	..	..	..	..	..	..	..	..	..	152	
Pel_Psemar	..	..	..	..	..	..	..	..	..	..	..	..	109	
Pela_Emnid	..	..	..	..	..	..	..	..	..	..	..	..	95	
Pelc_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	163	
Pelb_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pel2_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pel3_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pela_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pelb_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pelc_Erwca	..	..	..	..	..	..	..	..	..	..	..	..	162	
Pela_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	181	
Peld_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	175	
Pele_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	170	
Pelf_Erwch	..	..	..	..	..	..	..	..	..	..	..	..	177	
Pel_Bacsub	..	..	..	..	..	..	..	..	..	..	..	..	219	
Pel_Psemar	..	..	..	..	..	..	..	..	..	..	..	..	170	
Pela_Emnid	..	..	..	..	..	..	..	..	..	..	..	..	130	

(continued on next page)

<sup>a</sup> Sequences shown and residue numbers listed correspond to mature proteins. The difference between residue number for precursor and residue number for mature protein for each of the sequences shown is: Pelc\_Erwch, 22; Pelb\_Erwch, 22; Pel2\_Erwca, 22; Pel3\_Erwca, 22; Pela\_Erwca, 22; Pelb\_Erwca, 22; Pelc\_Erwca, 22; Pela\_Erwch, 32; Peld\_Erwch, 31; Pele\_Erwch, 30; Pelf\_Erwch, 41; Pel\_Bacsub, 21; Pel\_Psemar, 29; Pela\_Emnid, 22. Invariant residues in the alignment are shown in bold.

<sup>b</sup> The nomenclature describes the structural elements that are similar between PelC and PelE. The solid lines represent the residue range of a particular structural element found in PelC that is also structurally conserved in PelE. T1 refers to the polypeptide segment, either a turn or a loop, connecting parallel β strands 3 and 1. The portions of the T1 and T3 turns which are not structurally conserved in both PelC and PelE are represented by a dotted line. The \* designates the position of the asparagine ladder found as the second residue of a 2-residue polypeptide connection between parallel β strands 2 and 3.

Table 1. (Continued)

Seq. Name											Res #		
Pelc_Erwch	TFESA	VDIKGASNTV	TVSYNYIHGV	KKVGLDGSST	SDTG	...	...	...	...	...	229		
Pelb_Erwch	TFESA	IDIKKGATYV	TISYNYIHGV	KKVGLSGFSS	SDTAE	...	...	...	...	...	229		
Pel2_Erwca	TFESA	VDIKKGSTNV	TVSYNYIHGI	KKVGLSGASN	TDTG	...	...	...	...	...	228		
Pel3_Erwca	TFESA	IDIKKASTNV	TISYNYIHGI	KKVGLSGFSS	SDTG	...	...	...	...	...	228		
Pela_Erwca	TFESA	VDIKKGSTNV	TVSYNYIHGV	KKVGLSGSSN	TDTG	...	...	...	...	...	228		
Pelb_Erwca	TFESA	VDIKKGSTNV	TVSYNYIHGI	KKVGLSGASN	TDTG	...	...	...	...	...	228		
Pelc_Erwca	TFESA	IDIKKASTNV	TISYNYIHGI	KKVGLSGFSS	SDTG	...	...	...	...	...	228		
Pela_Erwch	QHDGA	LDIKRGSYDV	TISNSLIDQH	DKTMLIGHND	T...	NSAQD	KGKLVTLFN	NVFNRTERA	PRVRYGSIHS		252		
Peld_Erwch	QHDGA	LDIKRGSYDV	TISNSRFLH	DKTILIGHSD	S...	NGSQD	SGKLRVTFHN	NVFDRTERT	PRVRFSGIHA		246		
Pele_Erwch	QHDGA	LDIKRGSYDV	TISYSRFELH	DKTILIGHSD	S...	NGSQD	SGKLRVTFHN	NVFDRTERT	PRVRFSGIHA		241		
Pelf_Erwch	QHDGS	LDIKRGSYDV	TVSNSRFELH	DKTILIGHSD	N...	NGSQD	AGKLRVTFHN	NLFDRTVGER	PRVRFSGVHA		248		
Pel_Bacsub	QHHDGQ	TDASNGANYI	TMSYNYIHDH	DKSSIFGSSD	S...	KTSDD	.GKLIKTLHH	NRYKNIVORA	PRVRFQVHV		290		
Pel_Psemar	EHERPK	LDVKNGANFV	TISYSVFKSH	EKNNLIGSSD	S...	RTDD	.GKLVITLHN	TLFENISARA	PRVRYQVHL		247		
Pela_Emnid	DLSG	GKDD	LDGL	VDISHGAEWI	TVSNTYFHDH	WKGSLIGHSD	N...	NEDED	LGHHLVTYAN	NYWVNVYSRT	PLIRFATVHI	208	
Pelc_Erwch	YNNLYTNI	TGSG	LNVRQNGQAL	IENNWFEKAI	NP	VTSRYDGKNF	G				274		
Pelb_Erwch	YNNLYTGI	TSSG	LNVRQNGKAL	IENNWFENAV	SP	VTSRYDGSNF	G				274		
Pel2_Erwca	YNNLYDGI	TGSG	FNVKQKIAL	IESNWFEAL	NP	VSTARNDSSNF	G				273		
Pel3_Erwca	YNNLYTGI	TSSG	LNVRQKIAL	IERNWFENAK	NP	VTSRYDGSNF	G				273		
Pela_Erwca	YTNLYDGI	KSSG	FNVKQKIAL	IESNWFEAL	NP	VSTARNDSSNF	G				273		
Pelb_Erwca	YTNLYDGI	TGSG	FNVKQKIAL	IESNWFEAL	NP	VSTARNDSSNF	G				273		
Pelc_Erwca	YNNLYTGI	TSSG	LNVRQKIAL	IERNWFENAK	NP	VTSRYDGSNF	G				273		
Pela_Erwch	FNNVFKG.DA	KDPVYRYQYS	FGIGTSGSVL	SEGNSFTI	ANL	SASKACKV	VKKF	...	...	...	307		
Peld_Erwch	YNNVYLG.DV	KNSVYPYLYS	FGLGTSGTIL	SESNSFTL	SNLKSIDGKN	PE	...	CSI	VKQF	...	NSK	305	
Pele_Erwch	YNNVYLG.DV	KNSVYPYLYS	FGLGTSGSIL	SESNSFTL	SNLKSIDGKN	PE	...	CSI	VKQF	...	NSK	300	
Pelf_Erwch	YNNVYVG.DV	NHKAYRYQYS	FGIGTSGSLL	SESNAFTI	DNMKKISGRD	KE	...	CSV	VKAF	...	NGK	307	
Pel_Bacsub	YNNYYEG.ST	SSSSYPFYSYA	WGIGKSSKIY	AQNVIDV							PGLSAA	KTISVFSGGT	343
Pel_Psemar	YNNYHVG.ST	SHKVYPFYSYA	HGVGKNSKIF	SERNAFEI							AGISGC	DKIAGDYGGS	300
Pela_Emnid	INNYWDSL	...	IDTG	VNCRMDAQVL	IQSSAFHN								238
Pelc_Erwch			TWVLKGNIT	KPADFSTYSI	TWTADTKPYV	NADSWTS							311
Pelb_Erwch			TWVLKGNIT	KPADFATYNI	TWTPDTKEYR	NADTWTS							311
Pel2_Erwca			TWELRNNIT	SPSDFAKYKI	TWGPSSPHI	NADNWKS							310
Pel3_Erwca			TWELRNNVM	SPADFAKYNI	TWDKDSKPYV	NAEDWKS							310
Pela_Erwca			TWELRNNIT	SPSDFAKYKI	TWGPSTPHI	NADWKS							310
Pelb_Erwca			TWELRNNIT	KPADFSKYNI	TWGRPSTPHV	NADWKS							310
Pelc_Erwca			TWELRNNVM	SPADFAKYNI	TWDKDTKPYV	NAEDWKS							310
Pela_Erwch			IFSDNGSVL			NGS	A	VDLSGCGF					328
Peld_Erwch			VFSDNGSLV			NGS	STTKLDTCAV						327
Pele_Erwch			VFSDKGSVL			NGS	TTTKLDTCTL						322
Pelf_Erwch			IFSDKGSII			NGS	S	YNLNGCGF	GF				330
Pel_Bacsub			ALYDSGTL			NGT	Q		INASAANG		LSSS		368
Pel_Psemar			VYRDTGSTL			NGS	A				LSCSWSSS		321
Pela_Emnid	CPDRAIFFAD	SDYTYAVVD	DVDLGGSSN										369
Pelc_Erwch	TGTF	PT	VAYNY	SPVSAQC	V	KDKLPG	YAGVGKNLAT	LTSTACK					353
Pelb_Erwch	TGTY	PT	VPYSY	SPVSAQC	V	KDKLAN	YAGVGKNLAT	LASSACK					353
Pel2_Erwca	TGKF	PS	ISYKY	TPVSAQC	V	KDKLAN	YAGVGKNLAV	LTAANCK					352
Pel3_Erwca	TGTF	AS	VPYSY	SPVSAQC	V	KDKLAN	YAGVKNLAV	LTAANCN					352
Pela_Erwca	TGKF	PA	VPYSY	SPVSAQC	V	KDKLAN	YAGVGKQAV	LTAANCK					352
Pelb_Erwca	TGKF	PS	ISYKY	TPVSAQC	V	KDKLAN	YAGVGKNLAV	LTAANCK					352
Pelc_Erwca	TGTF	AS	VPYSY	SPVSAQC	V	KDKLAN	YAGVKNLAV	LTAANCN					352
Pela_Erwch	SAY	TSK	IPYIY	DVQPMT	TELAQSITD	NAGSGKL							361
Peld_Erwch	TAY	KPT	LPYKY	SAQTMT	SSLASSINS	NAGYGKL							360
Pele_Erwch	TAY	KPT	LPYKY	SAQTMT	SSLATSINN	NAGYGKL							355
Pelf_Erwch	SAY	SAK	IPYKY	SAQTIT	TSLAGSISS	NAGYGKL							363
Pel_Bacsub	VGW	TPS	L		KSNVIN	QAGAGKLN							399
Pel_Psemar	IGW	TP	PYSY	TPLAADK	V	AADVKA	KAGAGKL						351
Pela_Emnid	PEGTLT	PSS	LPY	AAITALGSGQ	V	ASVIPG	TAGQ.KL						304

'potentially catalytic' invariant amino acids, but many other invariant residues found in the amino (N)-terminal branch (GyatxxxxTxGG), the midregion of the parallel  $\beta$  helix (vWiDH; TxS; HaynN) and the carboxy (C)-terminal branch of the enzymes (AGxxK), all folded together in three dimensions. The vWiDH cluster is separated from the  $\text{Ca}^{2+}$  site by approximately 95 Å, too far to be involved in the pectinolytic function. Thus, the identification of a second unique cluster of 'potentially catalytic' residues suggests that the pectate lyases have a second, as yet unidentified, enzymatic function. Although site-directed mutational studies will aid the search for a second function, the relative orientation of the invariant DH side chains in the PelC and PelE structures is reminiscent of active sites found in many protease structures (Branden and Tooze 1991).

An analysis of the invariant amino acids lacking in catalytic properties is also revealing. Two, I54 and N232 in PelC, are involved in the internal side-chain stacking interactions which appear to confer stabilizing properties on the pectate lyases in solution as well as in a hostile extracellular environment. The invariant proline, P220 in PelC, has an unusual *cis* conformation in the three-dimensional structures, apparently to properly align the invariant arginine, R218 in PelC, for an important catalytic function. At 26%, the percentage of invariant glycines is unusually high, indicating crucial roles in controlling conformational changes or in avoiding steric clashes. Most notable is the GyatxxxxTxGG sequence in the N-terminal branch. A database search reveals that a similar pattern is found in many precursor sequences (personal observations). Andersson and von Heijne (1991) have predicted that exported proteins contain sequence signals in the first 30 amino acids following the N-terminal signal peptide which is removed during or after export of the protein. Indeed, they found that export was blocked by the insertion of a string of lysines within the first 30 amino acids. From an enzymatic and structural viewpoint, it is also logical to expect that some type of signal may exist. For example, during enzymatic cleavage of an internal bond, in this case, the cleavage of the peptide bond between the signal sequence and the mature protein, there are generally groups on both sides of the scissile bond which contribute to the recognition of the substrate by the enzyme. The structural comparison of PelC and PelE suggest that the GyatxxxxTxGG sequence may form part of the signal on the mature protein side of the scissile bond. Not only is the unusual conformation of the N-terminal branch structurally similar, but the conserved aromatic group, Y7 in PelC, is tightly bound in an apparent specificity pocket lined with conserved aromatic groups. Moreover, a small rotation about the  $\phi$  bond of the invariant glycine preceding the conserved aromatic group brings the N terminus to the invariant histidine in the vWiDH cluster. The invariant residues in the TxGG sequence are linked to the invariant tryptophan in the vWiDH sequence through a hydrogen bonding network. Thus, the invariant amino acids in the N-terminal branch appear to have some type of alignment function for a catalytic activity occurring at the vWiDH sequence.

In summary, a structure-based multiple alignment of extracellular pectate lyase sequences corrects errors in the evolu-

tionary-based alignments and reveals new sequence patterns which suggest a second catalytic function for the pectate lyases.

## ACKNOWLEDGMENTS

The research is supported by the United States Department of Agriculture (award #94-37303-0730) and the San Diego Supercomputer Center.

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