# Greenhouse and Field Evaluation of *Fusarium solani* Pathogenicity to Soybean Seedlings

J. F. KILLEBREW, Extension Plant Pathologist, K. W. ROY, Professor of Plant Pathology, G. W. LAWRENCE, Assistant Professor of Plant Pathology, and K. S. McLEAN, Graduate Research Assistant, Department of Plant Pathology and Weed Science, and H. H. HODGES, Professor of Agronomy, Department of Agronomy, Mississippi State University, Mississippi State 39762

### **ABSTRACT**

Killebrew, J. F., Roy, K. W., Lawrence, G. W., McLean, K. S., and Hodges, H. H. 1988. Greenhouse and field evaluation of *Fusarium solani* pathogenicity to soybean seedlings. Plant Disease 72:1067-1070.

Koch's postulates were completed with Fusarium solani under both greenhouse and field conditions. Root rot was the most prevalent and damaging symptom caused by the fungus. Five F. solani isolates tested were pathogenic when inoculated on soybean, and each was reisolated from inoculated, symptomatic plants. Isolates differed in virulence. Disease severity was not substantially changed when F. solani and soybean cyst nematode (Heterodera glycines) were inoculated on soybean in combination, as compared with F. solani or soybean cyst nematode alone. The fungus reduced yield under field conditions. Disease severity was greater with poorquality seeds inoculated with F. solani than with inoculated high-quality seeds.

Additional keywords: Glycine max, seed quality

Seedling disease complex of soybean (Glycine max (L.) Merr.) causes annual economic losses in Mississippi (11) and other soybean-producing areas in the United States (1,19). Fusarium spp. are major fungal components of the complex (1,6,14,19,22), although their pathogenicity has generally been difficult to demonstrate and some species are considered secondary invaders (3,9,15, 21,23,24). The presence of nematodes in soil was reported to increase seedling disease severity and/or colonization of seedlings by soilborne fungi (5,14).

Fusarium solani (Mart.) Appel & Wollenw. emend. Snyd. & Hans. is

Published with the approval of the director, Mississippi Agricultural and Forestry Experiment Station as Journal Series Paper 6754.

This research was supported in part by funds from the Mississippi Soybean Promotion Board.

Accepted for publication 28 June 1988.

frequently isolated from soybean seedlings, but there are conflicting reports regarding its pathogenicity (7,9,15,16,20,21,23,24). Results of Schlub et al (15,16) indicated that predisposition of soybean seeds by drought stress for 4–6 days was required for infection of seedlings. Only one study (24), conducted in the greenhouse, demonstrated that *F. solani* was capable of reducing soybean yield. There have been no reports of infection or reduced yield in the field.

This paper reports the completion of Koch's postulates with different isolates of *F. solani* in both greenhouse and field, the effect of *F. solani* isolate and inoculum concentration on disease severity, predisposing effects of nematodes and poor seed quality on infection of seedlings by *F. solani*, and effect of the fungus on yield.

# MATERIALS AND METHODS

Effect of F. solani isolate and inoculum level on disease severity in the greenhouse. Five single-spore isolates of

F. solani, obtained from symptomatic roots of cv. Coker 156 soybean seedlings and designated K15, K37, B312, H216, and C39, were tested. Their identification was based on descriptions reported by Booth (2) and Nelson et al (12). Isolates were maintained on slants of modified Bilay's medium (2) at 5 C.

To produce inocula, isolates were grown at 24 C for 10 days in sterile sandcornmeal cultures (250 cm<sup>3</sup> of dry sand, 14 cm<sup>3</sup> of cornmeal, and 100 ml of distilled water). A mechanical soil mixer was used to mix portions of the inocula with sterilized soil to obtain inoculum levels of 0.050%, 0.025%, and 0.0125% (w/w). The potting medium used in all tests was a 1:1 (v/v) mixture of sand and soil fumigated with methyl bromide. Infested and noninfested (control) soil each were distributed to 12.5-cmdiameter pots. Ten soybean cv. Bragg seeds were surface-disinfested in 1% sodium hypochlorite, rinsed in sterile water, and aseptically plated on Difco potato-dextrose agar (PDA). After 36 hr, germinated seeds free from microorganisms were placed on the soil surface in each pot and covered with 2.5 cm of the appropriate soil. Experimental units, each replicated four times, were arranged in a completely random design and plants were watered daily. Disease severity, assessed 3 wk after planting, was based on a 1-6 scale: 1 = no symptoms, 2-4 =increasingly severe root-hypocotyl necrosis, 5 = postemergence dampingoff, and 6 = preemergence damping-off. To assure uniformity in experimental units, plants were randomly culled to five per pot before disease assessment. During the course of experiments, temperatures in the greenhouse ranged

<sup>© 1988</sup> The American Phytopathological Society

from 16 to 25 C during the night and from 25 to 34 C during the day.

Effect of F. solani alone and in combination with H. glycines on disease severity in the greenhouse. Isolates U32, H216, FS, and FSC of F. solani, either alone or in combination with soybean cyst nematode (Heterodera glycines Ichinohe), were tested for pathogenicity. An inoculum level of 0.050%, obtained as previously described, was used to inoculate Bragg seedlings. Nematodes were obtained by increasing a population of race 3 on Coker 156 soybean for 60 days in the greenhouse. Light brown to tan cysts were dislodged from roots with a stream of water and collected on nested 20- and 60-mesh sieves. Cysts were suspended in water, then immediately poured through a 850-µm-pore sieve nested on a 250- $\mu$ m-pore sieve. They were then washed from the 250- $\mu$ m-pore sieve into 20-ml glass test tubes with 10 ml of tap water and crushed with a modified Seinhorst cyst crusher (18) for 1 min (G. W. Lawrence, unpublished). The resultant suspension was passed through a 75-μm-pore sieve nested on a 28-μmpore sieve to remove broken cysts and debris. Eggs and second-stage juveniles were standardized to a population of 686 eggs and 43 juveniles per milliliter. Five

germinated and apparently healthy seeds, obtained as previously described, were placed on the surface of methyl bromide-fumigated soil in each pot and covered with 2.5 cm of the appropriate soil. Seven milliliters of the H. glycines suspension were pipetted into three  $1 \times 4$ cm depressions in soil, and a thin layer of fumigated soil was added to prevent egg desiccation. Experimental units, each replicated seven times, were arranged in a completely random design, and plants were watered daily. Disease severity was assessed 3 wk after planting, using the previously described method. Sections of root tissue from inoculated, symptomatic plants and from noninoculated plants were surface-disinfested in 95% ethanol for 5 sec and 1% NaOCl for 1 min, rinsed in sterile water, then aseptically plated on PDA. Colonies of F. solani growing from tissue were determined during a 1-wk incubation period. This experiment was conducted twice, during which time greenhouse temperatures ranged from 15 to 21 C during the night and from 21 to 29 C during the day.

For each experiment, data were subjected to analysis of variance and means were separated using Duncan's multiple range test. Percentage stand data were transformed to arcsine before

Table 1. Pathogenicity of Fusarium solani isolates to 3-wk-old Bragg soybean seedlings in the greenhouse

Isolate	Disease severity index <sup>y</sup>	Root volume (cm³)	Height (cm)	Stand (% of control)
K15	2.7 b <sup>z</sup>	8.0 ab	15.4 a	92 b
K37	2.8 b	7.6 ab	15.6 a	91 b
B312	2.9 b	8.6 a	15.1 a	92 b
H216	3.7 a	6.0 b	14.6 a	92 b
C39	2.6 b	9.0 a	15.0 a	95 b
Control	2.1 c	10.1 a	15.9 a	100 a

<sup>&</sup>lt;sup>y</sup>Disease severity was based on a 1-6 scale: 1 = no symptoms, 2-4 = increasingly severe roothypocotyl necrosis, 5 = postemergence damping-off, and 6 = preemergence damping-off.

**Table 2.** Pathogenicity of *Fusarium solani* isolates alone or in combination with soybean cyst nematode, *Heterodera glycines* race 3, to 3-wk-old Bragg soybean seedlings in the greenhouse

Isolate and/or nematode treatment	Disease severity index <sup>y</sup>	Stand (% of control)	Root volume (cm³)	Height (cm)
U32	2.7 c²	96 b	11.3 b	13.5 ab
U32 + H. glycines	2.8 bc	95 b	10.6 bc	12.5 ab
H216	3.5 ab	93 с	8.6 cd	12.4 b
H216 + H. glycines	3.0 bc	95 b	9.7 bc	13.5 ab
FS	2.5 cd	96 b	10.0 bc	13.0 ab
FS + H. glycines	2.4 cd	96 b	10.4 bc	12.7 ab
FSC	4.2 a	82 e	5.7 e	10.9 с
FSC + H. glycines	4.2 a	86 d	7.2 de	10.8 c
H. glycines	1.9 de	99 a	14.0 a	12.2 b
Control	1.5 e	100 a	14.5 a	13.9 a

<sup>&</sup>lt;sup>y</sup> Disease severity was based on a 1-6 scale: 1 = no symptoms, 2-4 = increasingly severe roothypocotyl necrosis, 5 = postemergence damping-off, and 6 = preemergence damping-off.

analysis.

Pathogenicity tests in the field. Field plots of soybean cv. Asgrow 5474 were established at the Mississippi State University Plant Science Farm, Starkville, in June 1985 and 1986. Asgrow 5474 is as susceptible to seedling infection by F. solani as Coker 156 (authors, unpublished). Isolates of F. solani were grown on a modified Bilay's medium (2) for 10 days at 24 C under a fluorescent light bank (160 Wm<sup>-2</sup>) set at a 12-hr photoperiod. The isolates used were recovered from diseased root tissues of soybean seedlings. Preliminary tests indicated that inoculation of seeds with conidia produced high levels of seedling infection (authors, unpublished). Inoculum of each isolate was obtained by flooding plates with sterile distilled water, dislodging conidia with a camel'shair brush, and adjusting inoculum to 1.5 × 10<sup>6</sup> macroconidia per milliliter using a hemacytometer. Seeds were placed in inflated plastic bags, the appropriate inoculum was added, and seeds were agitated for 1 min to obtain uniform coverage. Seeds similarly treated with sterile distilled water served as controls. Seeds were stored at 10 C until planted. Immediately before planting, subsamples of inoculated seeds were plated on PDA to verify viability of inoculum.

To determine the effect of different isolates on disease severity, isolates H216, B312, and C39 were compared in 1985 and H216, B312, C39, and U32 were compared in 1986. Field plots were three rows each 6 m long on 1-m centers and were established in a randomized complete block design, with four replications in 1985 and seven in 1986.

To determine the effect of seed quality on disease severity, poor-, intermediate-, and high-quality seeds, testing 60, 70, and 85% germination on PDA, respectively, were inoculated with isolate H216, the most virulent isolate in previous pathogenicity tests. The levels of seed quality were obtained by examining seeds with a 7× magnifier and separating seeds having fissured seed coats or dull appearance (poor quality) from those appearing healthy (high quality); the two seed lots were intermixed to obtain intermediate quality. Three-row plots 6 m long on 1-m centers were established in a randomized complete block design with a split-plot arrangement of treatments, each replicated four times. Inoculation and seed quality were main and subplots, respectively.

In both years, seedling numbers (stand), plant height, and disease severity were determined 6 wk after planting in each field test. Plant height and disease severity were determined from 25 randomly selected plants per replicate. Postemergence damping-off was monitored during the 6-wk period, and yield was determined for the center row in each plot at maturity. Disease severity was

Means within a column followed by the same letter are not significantly different (P = 0.05) according to Duncan's multiple range test.

Means are averages of two experiments (14 total replications). Means within a column followed by the same letter are not significantly different (P = 0.05) according to Duncan's multiple range test.

based on a 1-5 scale: 1 = no symptoms, 2-4 = increasingly severe root-hypocotyl necrosis, and 5 = postemergence damping-off. In each experiment, sections of root tissue from 25 randomly sampled plants per replicate were surface-disinfested in 1% NaOCl and aseptically plated on PDA acidified with HCL (APDA), and colonies of F. solani growing from tissues were determined.

#### RESULTS

Effect of F. solani isolate and inoculum level on disease severity in the greenhouse. Isolates of F. solani caused root rot and preemergence damping-off and were reisolated from symptomatic root tissue plated on APDA (Table 1). Root rot was the most prevalent symptom. Lesions varied in color from light to dark brown, were often sunken. and seldom occurred above the soil line. Little postemergence damping-off was observed. All isolates were pathogenic as measured by disease severity index, root volume, and stand. Isolate H216 was the most virulent. Virulence of the other four isolates did not differ. None of the isolates had a significant effect on seedling height in these tests. Inoculum level had no effect on disease severity and did not interact with isolates (P = 0.05).

Effect of F. solani alone and in combination with H. glycines on disease severity in the greenhouse. H. glycines alone had no significant effect on disease severity index, root volume, or stand but reduced seedling height (Table 2). Each isolate of F. solani was pathogenic as measured by disease severity index, stand, and root volume. Isolates FSC and H216 caused significant height reductions. Combinations of H216 and H. glycines and of FSC and H. glycines caused less stand reduction than did H216 or FSC alone. Isolates H216 and FSC caused more severe symptoms and greater reductions in stand and root volume than did isolates U32 and FS. Isolate FSC caused significantly greater reductions in stand, root volume, and height than the other three isolates.

F. solani was recovered from more than 90% of the surface-disinfested, symptomatic plants but from less than 1% of the noninoculated plants.

Effect of different isolates on disease severity in the field. Each isolate of F. solani caused seedling disease in the field (Table 3). The isolates differed in virulence. In both years, isolate H216 was the most virulent and caused reduced stand, seedling height, and yield. In 1985, B312 was the only other isolate that reduced plant performance, causing a yield reduction. In 1986, isolates U32, B312, and C39 had no significant effect on stand, but each caused significant levels of disease on roots and hypocotyls. Isolate U32 reduced seedling height and yield, and isolate B312 reduced yield. In

general, isolate C39 was the least virulent and did not significantly reduce plant performance.

In both years, *F. solani* was recovered from surface-disinfested tissues of more than 75% of the inoculated plants but from less than 20% of noninoculated plants.

Effect of seed quality on disease severity in the field. The mean squares for both main effects and the interaction were significant. As measured by each parameter, high-quality seeds were superior to intermediate- and poorquality seeds (Table 4). Seedling disease was produced by F. solani on poor-, intermediate-, and high-quality seeds but reduced stand, seedling height, and yield only on poor-quality seeds. The interaction indicated that F. solani was more destructive on poor-quality seeds than on intermediate- and high-quality seeds, as measured by disease severity index and three other parameters.

Percentage reisolation of *F. solani* from surface-disinfested tissues of inoculated and noninoculated plants was similar to that in previous field tests.

## DISCUSSION

Koch's postulates were fulfilled with five F. solani isolates under both greenhouse and field conditions. This is the first demonstration of pathogenicity and subsequent yield reduction by this fungus on soybean in the field.

F. solani was pathogenic on soybeans in some studies (7.16.20.24) and nonpathogenic in others (9,15,21,23). Strain differences, which are suggested by the differential virulence of F. solani isolates tested herein, may at least partially explain these conflicting reports. This is supported by comparative results of our study and of Klag et al (9). Klag et al used similar inoculation methods for pathogenicity trials with F. solani in the greenhouse, yet none of their several isolates tested was pathogenic. Nonpathogenic strains of F. solani (2,10) and other Fusarium species (3) are known to occur on other hosts.

Nematodes and fungi can act synergistically to increase the incidence and/or severity of certain soilborne diseases of soybean (5,13,14). Therefore, it was surprising that when F. solani was

**Table 3.** Pathogenicity of *Fusarium solani* isolates inoculated on Asgrow 5474 soybean seeds in the field in 1985 and 1986

	Disease parameter <sup>y</sup>			
Isolate	Seedling disease severity index	Stand (% of control)	Seedling height (cm)	Seed yield (kg/ha)
		1985		
C39	3.8 ab <sup>z</sup>	77 ab	21.4 a	2055 a
B312	3.7 ab	84 ab	21.0 a	1822 b
H216	4.2 a	65 b	16.4 b	1578 b
Control	3.3 b	100 a	23.0 a	2138 a
		1986		
H216	4.1 a	82 b	13.2 b	849 b
U32	3.9 b	96 a	13.0 b	916 b
B312	3.8 b	96 a	13.8 ab	936 b
C39	3.8 b	93 a	13.9 ab	1058 a
Control	3.5 c	44 a	14.5 a	1078 a

<sup>&</sup>lt;sup>y</sup> Disease severity index, stand, and plant height were obtained 6 wk after planting. Disease severity was based on a 1–5 scale: 1 = no symptoms, 2-4 = increasingly severe root-hypocotyl necrosis, and 5 = postemergence damping-off.

**Table 4.** Effect of different levels of seed quality on pathogenicity of *Fusarium solani* inoculated on Asgrow 5474 soybean seeds in 1986

Seed quality	Disease parameter <sup>y</sup>				
	Seedling disease severity index	Seedling numbers (stand)	Seedling height (cm)	Seed yield (kg/ha)	
Poor	4.7 a <sup>z</sup>	16 d	16.9 c	578 d	
Control	4.1 b	26 c	23.3 b	1097 c	
Intermediate	4.1 b	41 b	26.3 ab	1636 b	
Control	3.8 c	45 b	28.0 a	1878 b	
High	3.3 d	80 a	29.6 a	2350 a	
Control	3.1 e	84 a	28.8 a	2245 a	

y Disease severity index, stand (per 6 m of row), and plant height were obtained 6 wk after planting. Disease severity was based on a 1-5 scale: I = no symptoms, 2-4 = increasingly severe roothypocotyl necrosis, and 5 = postemergence damping-off.

<sup>&</sup>lt;sup>2</sup> Means within a column and year followed by the same letter are not significantly different (P = 0.05) according to Duncan's multiple range test.

Means within a column followed by the same letter are not significantly different (P = 0.05) according to Duncan's multiple range test.

combined with H. glycines, seedling stand was significantly greater than that which occurred when either isolate was used alone. Even though this effect on stand was inconsistent with the effect on disease severity, seedling height, and yield, it warrants further investigation.

Results of Schlub et al (15,16) indicated that predisposition of soybean seeds by drought stress for 4-6 days was required for infection of seedlings grown in soil infested with F. solani. We obtained high levels of infection without predisposing seeds, but disease severity was negatively associated with increased seed quality. Some evidence (8,17) indicates that exudates from seeds are at least in part responsible for increased infection of soybean seedlings by certain soilborne fungi. Seeds with fissured seed coats or other kinds of damage express greater amounts of exudates than healthy seeds (4,17). Exudates are known to contain amino acids, sugars, and other nutrients utilized by fungi for growth (4,8,17). Thus, a likely explanation for the association of higher levels of F. solani infection with poor-quality seeds is that the growth of F. solani was enhanced by exudates from such seeds.

Even though stand was reduced in some instances by some F. solani isolates, the slight reductions were of secondary importance compared with root rot. Data indicate that the yield reductions observed resulted from early, and presumably prolonged, colonization of roots by F. solani.

#### LITERATURE CITED

- 1. Athow, K. L. 1973. Fungal diseases. Pages 459-489 in: Soybeans: Improvement, Production and Uses. B. E. Caldwell, ed. American Society of Agronomy, Madison, WI.
- 2. Booth, C. 1971. The Genus Fusarium. Commonwealth Mycological Institute, Kew, Surrey, England. 237 pp.
- Cromwell, R. O. 1917. Fusarium blight, or wilt disease, of the soybean. J. Agric. Res. 8:421-439.
- 4. Flentje, N. T. 1959. The physiology of penetration and infection. In: Plant Pathology Problems and Progress, 1908-1958. C. S. Holton et al, eds. University of Wisconsin Press, Madison.
- 5. Fortnum, B. A., and Lewis, S. A. 1977. Populations of certain fungi in soybean roots and rhizosphere related to infection by Meloidogyne incognita and Hoplolaimus columbus. (Abstr.) Phytopathology 67:222.
- 6. French, E. R., and Kennedy, B. W. 1963. The role of Fusarium in the root rot complex of soybean in Minnesota. Plant Dis. Rep. 47:672-676.
- 7. Grant, C. E., Phipps, P. M., and Roane, C. W. 1981. Etiology of damping-off disease of soybeans in Virginia. (Abstr.) Phytopathology 71:767.
- 8. Keeling, B. L. 1972. Studies on the nature of soybean resistance to seed rot caused by Pythium. (Abstr.) Phytopathology 62:768.
- Klag, N. G., Papavizas, G. C., Bean, G. A., and Kantzes, J. G. 1978. Root rot of soybeans in Maryland. Plant Dis. Rep. 62:235-239.
- 10. Kraft, J. M., Burke, D. W., and Haglund, W. A. 1981. Fusarium diseases of bean, peas, and lentils. Pages 142-156 in: Fusarium: Diseases, Biology, and Taxonomy. P. E. Nelson, T. A. Toussoun, and R. J. Cook, eds. Pennsylvania State University Press, University Park.
- 11. Mulrooney, R. P. 1986. Southern United States soybean disease loss estimate for 1985. Pages 9-13 in: Proc. South. Soybean Dis. Workers Conf., Baton Rouge, LA.
- Nelson, P. E., Toussoun, T. A., and Marasas, W. F. O. 1983. Fusarium Species: An Illustrated Manual for Identification. Pennsylvania State

- University Press, University Park. 193 pp.
- 13. Ross, J. P. 1965. Predisposition of soybeans to Fusarium wilt by Heterodera glycines and Meloidogyne incognita. Phytopathology 55:361-364.
- 14. Schenck, N. C., and Kinlock, R. A. 1974. Pathogenic fungi, parasitic nematodes, and endomycorrhizal fungi associated with soybean roots in Florida. Plant Dis. Rep. 58:169-173.
- 15. Schlub, R. L., Lockwood, J. L., and Komada, H. 1981. Colonization of soybean seeds and plant tissue by Fusarium species in soil. Phytopathology
- 16. Schlub, R. L., and Schmitthenner, A. F. 1978. Effects of soybean seed coat cracks on seed exudation and seedling quality in soil infested with Pythium ultimum. Phytopathology 68:1186-1191.
- 17. Schroth, M. N., and Cook, R. J. 1964. Seed exudation and its influence on preemergence damping-off of bean. Phytopathology 54:670-673.
- 18. Seinhorst, J. W., and Den Ouden, H. 1966. An improvement of Bijloo's method for determining the egg content of Heterodera cysts. Nematologica 12:170-171.
- 19. Sinclair, J. B., ed. 1982. Compendium of Soybean Diseases. American Phytopathological Society, St. Paul, MN. 104 pp.
- 20. Spaulding, D. W., and Crittenden, H. W. 1965. Fungi associated with diseased roots of soybeans in Delaware. (Abstr.) Phytopathology 55:506.
- 21. Tu, J. C. 1978. Prevention of soybean root nodulation by tetracycline and its effect on soybean root rot caused by an alfalfa strain of Fusarium oxysporum. Phytopathology 68:1303-1306.
- 22. Warren, H. L., and Kommedahl, T. 1973. Fusarium species in roots and soil associated with monoculture of soybeans in Minnesota. Plant Dis. Rep. 57:912-914.
- 23. Zakaria, G., and Lockwood, J. L. 1981. Fusarium spp. from soybean roots. Phytopathology 71:157-161.
- 24. Zambolin, L., and Schenck, N. C. 1981. Interactions between vesicular-arbuscular mycorrhiza and root-infecting fungi on soybean. (Abstr.) Phytopathology 71:207.