Effect of Plant Nutrition on Susceptibility of Chrysanthemum morifolium to Erwinia chrysanthemi

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ABSTRACT

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Susceptibility of Chrysanthemum morifolium cv. Bonnie Jean to Erwinia chrysanthemi was affected by rate of complete fertilizer (NPK, 20-10-20) and by form of nitrogen. The rate of complete fertilizer was varied from 0 to a concentration containing about 473, 105, and 413 ppm of nitrogen, phosphorus, and potassium, respectively. In separate experiments, three forms of nitrogen were varied from 0 to 400 ppm. Susceptibility was based on the amount of pith maceration in cuttings 4 days after dip inoculation in a bacterial suspension containing about 10⁸ colony-forming units per milliliter. Susceptibility increased with increasing rates of complete fertilizer and nitrogen in the form of (NH₄)₂SO₄. However, susceptibility was maximal at moderate levels of Ca(NO₃)₂ and NH₄NO₃ (100–200 ppm of nitrogen) and decreased when nitrogen was increased to 400 ppm in both forms.

Host nutrition has been widely implicated in the predisposition of plants to disease (7). Extremely high levels of nitrogen significantly reduced the severity of bacterial leaf blight in Philodendron selloum by Erwinia chrysanthemi Burkholder, McFadden, & Dimock (6). Conversely, high levels of nitrogen increased susceptibility of apples to E. amylovora (12,17,18), corn to E. stewartii (14) and E. chrysanthemi (19), and cabbage, tomato, and sugar beets to E. carotovora (4,20,22). Low levels of nitrogen predisposed cucumbers to E. tracheiphila (23). High rates of fertilizer were also found to increase the susceptibility of Chrysanthemum morifolium Ramat. to bacterial leaf spot incited by Pseudomonas cichorii (9).

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The form in which a nutrient is administered is often the key factor in its influence on disease. This concept, as applied to nitrogen, has been well documented by Huber and Watson (8). High levels of NH₄NO₃, for example, applied in a complete fertilizer increased Fusarium wilt in chrysanthemum (11). When applied independently, however, nitrate nitrogen (NO₃-N) decreased and ammonium nitrogen (NH₄-N) increased the severity of disease (24).

The purpose of this research was to examine the effects of different rates of a complete fertilizer on the susceptibility of *C. morifolium* cuttings to *E. chrysanthemi*. We were also interested in determining the effects of different forms of nitrogen on susceptibility. The selection of rates for experimentation was based on the general recommendation that 200 ppm of both nitrogen and potassium be applied daily to chrysanthemums (5,16).

MATERIALS AND METHODS

In a preliminary experiment during fall

1981, cuttings of C. morifolium cv. Bonnie Jean that received daily applications of 1.2 kg/L of a complete fertilizer (Peters 20-10-20, Peter's Fertilizer Products, W. R. Grace Co., Fogelsville, PA) were more susceptible to E. chrysanthemi than cuttings from unfertilized plants. In a subsequent experiment during winter 1981, complete fertilizer at 0.6, 1.2, or 2.4 kg/L was applied daily for 5 wk. Control plants received daily applications of tap water alone. In separate experiments during summer 1982, five rates of nitrogen (25, 50, 100, 200, and 400 ppm) in the form of Ca(NO₃)₂, NH₄NO₃, or (NH₄)₂SO₄ were applied in conjunction with 200 ppm of potassium in the form of K2(SO4) for 5 wk. Control plants received only 200 ppm of potassium.

Plants were grown in 17-cm-diameter pots using Cornell Peat-lite medium A (16) and were maintained in a greenhouse at about 29 C with a photoperiod of at least 16 hr. The Peat-lite medium was amended with dolomitic limestone at a rate of 3.4 kg/m³ in the Ca(NO₃)₂ treatment and 5.2 kg/m³ in the NH₄NO₃ and (NH₄)₂SO₄ treatments. Five plants were used per treatment, and their shoot tips were removed 6 days after potting to encourage the growth of side shoots.

Differences in susceptibility to the bacterium were assessed by bacterial maceration assays. Three shoots were selected randomly from each test plant, trimmed to 40 mm, and inoculated with a standardized suspension of *E. chrysanthemi* strain 159, corresponding to about 10⁸ colony-forming units per milliliter. The inoculum level was determined to be optimal for our

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experimental conditions. To produce bacterial inoculum, 24-hr nutrient agar (Difco) cultures of strain 159 of E. chrysanthemi were streaked on slants of modified yeast-dextrose-calcium carbonate agar (2) containing 5 g/L of dextrose rather than 20 g/L. All cultures were grown at 27 C and used within 24 hr. Shoot bases were placed in tubes containing sterile water, enclosed in plastic bags, and incubated for 4 days at 29 C. After incubation, each cutting was dissected longitudinally and the extent of pith maceration was measured (mm). In addition, three recently enlarged, mature leaves were removed from each plant, pooled by treatment, and used for nitrogen analysis by the micro-Kjeldahl method (1). Phosphorus and potassium were measured by inductively coupled argon plasma atomic emission spectroscopy (ICP) developed by the Department of Pomology, Cornell University, Ithaca, NY. In this procedure, 0.2 or 0.4 g of dry, ground sample is weighed into either quartz crucibles or test tubes. The sample

is placed in a muffle furnace and dryashed overnight at 450 C. The ashed sample is cooled and 0.25 ml of concentrated nitric acid is added before additional digestion on a hot plate until dry. The acid is dissolved in a 2-ml solution of 10% hydrochloric acid and nitric acid, then 8 ml of distilled water is added. The sample is mixed, and when the sediment has settled to the bottom, it is assayed twice by ICP.

RESULTS AND DISCUSSION

Susceptibility as measured by pith maceration increased with increasing rates of all forms of fertilizer studied (Fig. 1). This increase in susceptibility was correlated with content of NPK (complete fertilizer) (Table 1) and with increased uptake of nitrogen in three forms (Table 2). Susceptibility was maximal at the highest rate of complete fertilizer and (NH₄)₂SO₄. Daily application of complete fertilizer in 2 yr of experimentation proved effective in maintaining high susceptibility to E. chrysanthemi with

В PITH MACERATION 2.4 400 20-10-20 (kg/l) N (ppm) С D 2 0 20 MACERATION 200

Α

Fig. 1. Regression of pith maceration on nutrients for cuttings of the chrysanthemum cultivar Bonnie Jean with daily applications of (A) 20-10-20 complete fertilizer (y = 6.64 + 4.89x, P = 0.02, r^2 = 0.97, SD = 1.2), (B) (NH₄)₂SO₄ (y = 4.50 + 0.048, P = 0.001, r² = 0.94, SD = 4.0), (C) Ca(NO₃)₂ (y $= 8.57 + 0.129x - 0.0003x^2$, P = 0.008, $r^2 = 0.93$, SD = 3.0), and **(D)** NH_4NO_3 (y = 11.86 + 0.108x - 0.008). $0.0002 x^2$, P = 0.04, $r^2 = 0.85$, SD = 5.8). Statistical analysis according to methods of SAS Institute, Cary, NC.

which to investigate viroid-E. chrysanthemi interactions (McGovern et al, unpublished).

Susceptibility was maximal at moderate levels of $Ca(NO_3)_2$ and NH_4NO_3 (100-200 ppm of nitrogen) and decreased sharply when nitrogen was increased to 400 ppm in both forms. Pith maceration increased linearly with increasing rates of complete fertilizer and (NH₄)₂SO₄ (Fig. 1A,B). For increasing concentration of Ca(NO₃)₂ and NH₄NO₃, pith maceration increased curvilinearly (Fig. 1C,D).

Plants can accumulate NO₃-N in excess of their needs, and some cases, this accumulation can reach phytotoxic levels (13). Transport of NO₃-N in a predominantly unreduced form occurs in the xylem of chrysanthemum, whereas reduction occurs in leaves (25). Perhaps at 400 ppm of nitrogen, unreduced Ca(NO₃)₂ accumulated in stem tissue at a level that directly inhibited the bacterium or adversely altered its environment by changing pH, osmotic pressure, or amino acid content or by induction of inhibitory plant-produced stress metabolites. Naidu et al (15) observed that increased susceptibility of rice to Xanthomonas oryzae resulted from treatment with ammonium sulfate and urea, whereas plants to which calcium or ammonium nitrate were applied were more resistant. This increased susceptibility was correlated with decreased phenolics and sugars and increased nitrogen content. Tejerina et al (21) reported that E. carotovora

Table 1. Effect of daily application of a 20-10-20 complete fertilizer on the uptake of NPK in Chrysanthemum morifolium cv. Bonnie Jean

	Mean % dry weight leaf tissue		
Treatment	N	P	K
Control ^a Complete fertilizer (g/L		0.238	3.82
0.6 ^b	2.86	0.351	3.95
1.2°	4.12	0.577	4.61
2.4 ^d	5.10	0.830	5.77

Plants were irrigated with tap water.

Table 2. Effect of daily application of Ca(NO₃)₂, NH₄NO₃ and (NH₄)₂SO₄ on the uptake of N in Chrysanthemum morifolium cv. Bonnie Jean

N (ppm)	Mean % N of dry weight leaf tissue			
	Ca(NO ₃) ₂	NH ₄ NO ₃	(NH ₄) ₂ SO ₄	
0	1.30	1.98	1.26	
25	2.24	2.32	2.20	
50	2.62	3.02	2.80	
100	3.36	3.42	3.72	
200	4.04	4.08	4.70	
400	3.92	4.46	6.82	

^bAbout 118, 26, and 103 ppm of N, P, and K, respectively

About 236, 52, and 206 ppm of N, P, and K, respectively.

dAbout 473, 105, and 413 ppm of N, P, and K, respectively.

underwent morphological changes, which they related to losses in pathogenicity, in a growth medium containing high concentrations of KNO₃. Excessive levels of nitrogen in the form of Ca(NO₃)₂ were also observed to significantly depress bacterial wilt of tomato incited by *Pseudomonas solanacearum* (10).

Although pith maceration was lower at 400 than at 200 ppm of nitrogen with NH₄NO₃, it was higher than at a comparable level of Ca(NO₃)₂. Perhaps this phenomenon may be attributed to a lower ratio of NO₃-N to NH₄-N in the tissue, which created a more favorable environment for the bacterium. Kelman (10) observed an increase in susceptibility of tomato to P. solanacearum when the ratio of NO₃-N to NH₄-N was increased. Although cuttings of Bonnie Jean from the (NH₄)₂SO₄ treatment were actually more susceptible to E. chrysanthemi at 400 ppm of nitrogen than their counterparts from the Ca(NO₃)₂ and NH₄NO₃ treatments, low and moderate rates of (NH₄)₂SO₄ resulted in less maceration than in the other two treatments.

At low and moderate rates of (NH₄)₂SO₄, the plant may be able to compete more successfully with the bacterium for the utilization of nitrogen. Although NH4-N can more readily induce phytotoxicity than NO₃-N (2), it should theoretically provide a more easily assimilated form of nitrogen when provided at nonexcessive levels (3). In fact, chrysanthemums convert NH₄-N directly into amino acids in their roots (25). At the highest rate of (NH₄)₂SO₄, plants were stunted and their foliage was darkened. These symptoms are typically observed with ammonium toxicity (3). NH₄-N and amino acids accumulating in excess of plant needs could have thereby provided an enhanced nutrient environment for the bacterium.

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LITERATURE CITED

- Association of Official Agricultural Chemists. 1950. Official Methods of Analysis. 8th ed. The Association: Washington, DC. 910 pp.
- Baigent, N. L., Devay, J. E., and Starr, M. P. 1963. Bacteriophages of *Pseudomonas syringae*. N.Z. J. Sci. 6:75-100.
- Barker, A. V., and Mills, H. A. 1980. Ammonium and nitrate nutrition of horticultural crops. Pages 395-423 in: Horticultural Reviews. Vol. 2.
 J. Janick, ed. AVI Publishing, Westport, CT.
- Bartz, J., Geraldson, C. M., and Crill, J. P. 1979. Nitrogen nutrition of tomato plants and susceptibility of the fruit to bacterial soft rot. Phytopathology 69:163-166.
- Crater, G. D. 1980. Pot mums. Pages 261-285 in: Introduction to Floriculture. R. A. Larson, ed. Academic Press, New York. 607 pp.
- Haygood, R. A., Strider, D. L., and Nelson, P. V. 1982. Influence of nitrogen and potassium on growth and bacterial leaf blight of *Philodendron* selloum. Plant Dis. 66:728-730.
- Huber, D. M. 1980. The role of mineral nutrition in defense. Pages 381-406 in: Plant Disease. An Advanced Treatise. Vol. 5. How Plants Defend Themselves. J. G. Horsfall and E. B. Cowling, eds. Academic Press, New York. 534 pp.
- 8. Huber, D. M., and Watson, R. D. 1974. Nitrogen form and plant disease. Annu. Rev. Phytopathol. 12:139-165.
- 9. Jones, J. B., Chase, A. R., Raju, B. C., and Harbaugh, B. K. 1984. The roles of fertilizer rate, light intensity, and humidity as pre-inoculation factors in susceptibility of chrysanthemums to *Pseudomonas cichorii* leaf spot. (Abstr.) Phytopathology 74:628.
- Kelman, A. 1950. Influence of nitrogen nutrition on the development of bacterial wilt in tomato and tobacco. (Abstr.) Phytopathology 40:14.
- Littrell, R. H. 1966. Effects of nitrogen on the susceptibility of chrysanthemums to an apparently new biotype of Fusarium oxysporum. Plant Dis. Rep. 50:882-884.
- 12. Link, G. K. K., and Wilcox, H. W. 1936. Relation of nitrogen-carbohydrate nutrition of Staymann

- apple trees to susceptibility to fire blight. Phytopathology 26:643-655.
- Maynard, D. N., and Barker, A. V. 1971. Critical nitrate levels for leaf lettuce, radish, and spinach plants. Commun. Soil Sci. Plant Anal. 2:461-470.
- McNew, G. L., and Spencer, E. L. 1939. Effect of nitrogen supply of sweet corn on the wilt bacterium. Phytopathology 29:1051-1067.
- Naidu, V. D., Rao, B. S., and Rao, C. S. 1979. Effects of nitrogen nutrition and bacterial leaf blight on rice leaves. Phytopathol. Z. 96:83-86.
- New York State College of Agriculture and Life Sciences. 1981. Cornell recommendations for commercial floricultural crops. Coop. Ext. N.Y. State Coll. Agric. Life Sci. Cornell Univ., Ithaca. 60 pp.
- Nightingale, A. A. 1936. Some chemical constituents of apple associated with susceptibility to fire blight. N.J. Agric. Exp. Stn. Bull. 613.
- Parker, K. G., Luepschen, N. S., and Fischer, E. G. 1961. Tree nutrition and fire blight development. Phytopathology 51:557-560.
- Saxena, S. C., and Lal, S. 1981. Effect of fertilizer application on the incidence of bacterial stalk rot of maize. Indian J. Mycol. Plant Pathol. 11:164-168.
- 20. Stuntz, Von E., Meyer, J., Meier, D., and Knosel, D. 1983. Versuche zur Verhutung der bakteriellen Strunkfaule des Lagerkohls durch Anbau-und Dungemabnachmen. [Investigation about the prevention of bacterial stalk rot of stored cabbage by measures of cultivation and fertilization.] Nachrichtenbl. Dtsch. Pflanzenschutzdienstes (Braunschweig) 35:49-50.
- Tejerina, G., Serra, M. T., and Castresana, M. C. 1978. Influence of fertilizer on *Erwinia* carotovora pathogenesis. Proc. 4th Int. Conf. Plant Path. Bact. Angers 2:607-615.
- Thomson, S. V., Hiels, F. J., Whitney, E. D., and Schroth, M. J. 1981. Sugar and root yield of sugar beets as affected by bacterial vascular necrosis and rot, nitrogen fertilization, and plant spacing. Phytopathology 71:605-608.
- Wei, C. T., Walker, J. C., and Scheffer, R. P. 1952. Plant nutrition in relation to disease development. VIII: Cucurbit wilts. Am. J. Bot. 39:245-249.
- Woltz, S. S., and Engelhard, A. W. 1973. Fusarium wilt of chrysanthemum: Effect of nitrogen source and lime on disease development. Phytopathology 63:155-157.
- Woodson, W. R. 1983. Accumulation, assimilation, and partitioning of nitrogen in Chrysanthemum × morifolium Ramat. Ph.D. thesis. Cornell University, Ithaca, NY. 140 pp.