Degrees of Sensitivity to Metalaxyl Within the *Pythium* spp. Pathogenic to Wheat in the Pacific Northwest

R. J. COOK, Research Plant Pathologist, U.S. Department of Agriculture, Agricultural Research Service, Washington State University, Pullman 99164, and BING-XING ZHANG, Visiting Plant Pathologist to Washington State University from Zhejiang Agricultural University, Hangzhou, Zhejiang, Peoples Republic of China

ABSTRACT

Cook, R. J., and Zhang, B.-X. 1985. Degrees of sensitivity to metalaxyl within the *Pythium* spp. pathogenic to wheat in the Pacific Northwest. Plant Disease 69:686-688.

A minimum of 20–25%, and in some cases more than 80%, of *Pythium* colonies selected at random from soil-dilution plates of a *Pythium*-selective medium grew when transferred to the same medium amended with metalaxyl at 1 ppm. The other colonies made no visible growth within 72 hr. Isolates of *P. ultimum* var. *ultimum*, *P. ultimum* var. *sporangiiferum*, *P. aristosporum*, *P. heterothallicum* (both male and female isolates), an unidentified heterothallic species (both male and female isolates), and an unidentified homothallic *Pythium* sp., all from wheat roots, failed to grow within 72 hr on the selective medium amended with metalaxyl at 1 ppm. In contrast, isolates of *P. torulosum*, *P. irregulare*, and a second unidentified homothallic *Pythium* sp., also from wheat roots, grew on the metalaxyl-amended *Pythium*-selective medium at 15–50% of their respective normal growth rates on the same medium without metalaxyl. On cornmeal agar with metalaxyl at 1 ppm, the less sensitive isolates grew at 30–80% of normal growth on the same medium without metalaxyl. With 0.1 ppm of metalaxyl added to cornmeal agar, most isolates of the more sensitive species grew at 70–80% and those of the less sensitive species grew at 95–100% of normal growth. The failure of metalaxyl to inhibit growth of some species of *Pythium* involved in root rot of Pacific Northwest wheat may account for observed failures of this fungicide to control this disease in the field

Additional key words: Triticum aestivum

Pythium spp. have become more important as pathogens of winter wheat (Triticum aestivum) in the Pacific Northwest states of the United States as growers have changed their management from common use of summer fallow, rotation, and conventional tillage to more recropping (wheat after wheat) and less tillage (6). Several species or subspecies may be involved, including P. torulosum Coker & Patterson, P. aristosporum Vanderpool, P. volutum Vanderpool & Truscott, P. ultimum Trow var. ultimum Drechs., P. ultimum

College of Agriculture Research Center Scientific Paper SP 6938

Cooperative investigations of the USDA-ARS and the Washington State University College of Agriculture and Home Economics Research and Extension Center.

Mention of a trademark or proprietary product does not constitute a guarantee or warranty of the product by the USDA and does not imply its approval to the exclusion of other products that also may be suitable.

Accepted for publication 8 February 1985 (submitted for electronic processing).

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. § 1734 solely to indicate this fact.

This article is in the public domain and not copyrightable. It may be freely reprinted with customary crediting of the source. The American Phytopathological Society, 1985.

var. sporangiiferum Drechs., P. irregulare Buisman, and P. heterothallicum Campbell & Hendrix (3,4). Damage may occur either as seedling blight or as a decay of the roots, rootlets, and root hairs, resulting in poor stands or stunted, poorly tillered plants. Yields in some fields have been 20-25% greater in plots with soil treated to eliminate or inhibit Pythium species (6). Trends toward consecutive cereal crops and minimum tillage are likely to continue in the region, and thus a method for the control of Pythium is needed.

In an earlier experiment (7), metalaxyl at about 2 kg/ha in the seed furrow as a 5G (5% a.i. granular) formulation was equivalent to fumigation with methyl bromide for control of Pythium damage to direct-drilled (no-till) wheat. Metalaxyl was then tested in various formulations (wettable powder, emulsifiable concentrate, and granular) with various rates and application methods (to seed, in the seed furrow, and postplant) in an effort to find the best method and rate of application for wheat. Yields were up to 20% greater in some trials, but there was no response in other trials despite high populations of *Pythium* in the fields (6). Moreover, Pythium oospores were found in abundance in roots and rootlets of metalaxyl-treated and untreated plants

Use of metalaxyl to control members of the Peronosporales has been shown to

select for resistant strains in an otherwise sensitive species, including Phytophthora infestans (10), Pseudoperonospora cubensis (11), and Pythium aphanidermatum (13). In addition, resistance to metalaxyl has been induced to Phytophthora megasperma f. sp. medicaginis (9) and Pythium ultimum (2). However, the wheat fields where metalaxyl was ineffective had no prior history of metalaxyl use, ruling out any possibility that a resistant strain had been favored. In preliminary work (8), a portion of the Pythium population in Pacific Northwest wheat fields was relatively insensitive to metalaxyl. Of the four Pythium spp. tested. P. torulosum showed the least sensitivity. At least 10 species or subspecies are involved in the Pythium root rot complex of Pacific Northwest wheat (4); this has prompted studies of sensitivity of the six species not included in our preliminary work (8). New isolates of the original four species were also included. Coffey et al (5) and Shew (14) have reported that strains as well as species of Phytophthora vary in ED50 values when exposed to metalaxyl.

MATERIALS AND METHODS

Tests of Pythium spp. for sensitivity to metalaxyl. Cultures of P. torulosum, P. aristosporum, P. ultimum vars. ultimum and sporangiiferum, P. irregulare, P. heterothallicum (both male and female), and three unidentified Pythium spp. (one heterothallic, both male and female) were grown on the selective medium of Mircetich (12) and also on cornmeal agar (Difco) (plus 10 g of additional agar), each amended with metalaxyl at 1 ppm. Each culture was isolated originally from roots of wheat taken directly from fields. Tests were also conducted with metalaxyl at 0.01, 0.1, 1, 2, and 5 ppm added to cornmeal agar (plus 10 g of agar). The metalaxyl was added to the molten medium as a water dilution of the 2EC formulation. The same media without metalaxyl were used as controls. Colony growth rates at 20 C were measured after 48 or 72 hr.

Tests of randomly selected *Pythium* colonies for sensitivity to metalaxyl. The proportion of the population of *Pythium* spp. sensitive to metalaxyl was assessed by first preparing soil dilution plates using the *Pythium*-selective medium of Mircetich (12) without metalaxyl, then

transferring randomly selected colonies to the same medium containing metalaxyl at 1 ppm. Soil dilutions of about 1:50 and 1:100 were used. Transfers to the medium with metalaxyl were made 48 hr after the dilution plates were seeded, by transferring blocks of agar containing mycelium from each colony selected. Five blocks representing five colonies were placed equidistantly around the margins of each petri dish of metalaxyl-amended medium. At least 50 colonies were transferred for each soil dilution plated. The presence or absence of growth from the respective disks was recorded after incubation for 48 hr at 20 C.

RESULTS

Growth of Pythium spp. on metalaxylamended media. All isolates of P. aristosporum, P. ultimum vars. ultimum and sporangiiferum, P. heterothallicum (both male and female), the unidentified heterothallic species (both male and female), and one of the two unidentified homothallic species failed to grow within 72 hr on either the selective medium (12) or cornmeal agar amended with metalaxyl at 1 ppm (Tables 1 and 2). In contrast, growth of the isolates of P. torulosum, P. irregulare, and the other unidentified (homothallic) Pythium spp. on media amended with metalaxyl at 1 ppm were 15-50% of the controls on the selective medium (12) (Table 1) and 30-80% of the controls on cornmeal agar (Table 2). Growth rates of P. irregulare within 72 hr were 10-20\% of the control at 2 ppm and were completely prevented or limited to a few short hyphae at 5 ppm. The more sensitive species grew at 70-80% and the less sensitive isolates grew at 95-100% of their normal growth rates, respectively, on cornmeal agar with metalaxyl at 0.1 ppm (Table 3). None of the isolates tested had been previously exposed to metalaxyl.

Ability of isolates of *Pythium* from wheat field soils to grow on metalaxylamended media. With each wheat field soil tested, as few as 22%, but as many as 86%, of the colonies selected at random from dilution plates of the selective medium grew when transferred to the same medium amended with metalaxyl at 1 ppm (Table 4).

DISCUSSION

As reported for *Phytophthora* spp. (5,14), considerable variability in sensitivity to metalaxyl exists among and even within species and strains of *Pythium* spp. Unfortunately, it appears that the less sensitive species make up 25–75% and possibly more of the total population of *Pythium* in wheat field soils of the Pacific Northwest. This could possibly explain, at least in part, why metalaxyl has failed to protect wheat roots and give a yield response in fields with a known problem of Pythium root rot.

Of the several *Pythium* taxa thus far isolated from roots of wheat in the Pacific

Northwest, *P. torulosum*, *P. irregulare*, and one of unidentified homothallic species were the least sensitive in vitro to metalaxyl. The ED₅₀ values for the less sensitive species are estimated at 0.6–0.7 or greater. In contrast, isolates of *P. aristosporum*, *P. ultimum* vars. *ultimum* and *sporangiiferum*, *P. heterothallicum*, and two unidentified species (one heterothallic) showed the degree of sensitivity to metalaxyl commonly reported for members of the Perono-

sporales (1,15), with ED₅₀ values estimated at 0.3 ppm or less. The 1-ppm concentration added to the medium serves mainly to separate the more sensitive from the less sensitive species.

Although the prospects for control of *Pythium* with metalaxyl are not encouraging, the potential for use of this fungicide should not be ruled out. Conceivably, different crop or soil management, practiced consistently in the different fields for many years, might

Table 1. Sensitivities of different Pythium spp. to metalaxyl added to a Pythium-selective medium^a

	Colony diameters (mm)				
Pythium sp.	Control		1 ppm metalaxyl		
(no. of isolates)	2 Days	3 Days	2 Days	3 Days	
P. aristosporum (2)	Tr ^b	Tr-15	0	0	
P. torulosum (1)	19.5	30	10.5	16	
P. ultimum					
var. sporangiiferum (2)	55.5-58.5	76-<85	0	0	
var. ultimum (1)	53.5	80	0	0	
P. ultimum (3)	55-63.5	<85	0	0	
P. irregulare (8)	48-58	<85	9-26	21-45	
Unknown A (homothallic)	51	<85	22	36	
Unknown B (homothallic)	18.5	29.5	0	0	
P. heterothallicum (female)	42	71.5	0	0	
P. heterothallicum (male)	39	60	0	0	
Unknown (male)	19.5	29.5	0	0	
Unknown (female)	21.5	35	0	0	

^a Mircetich's (12) selective medium unamended or amended with 1 ppm of metalaxyl.

Table 2. Sensitivities of different Pythium spp. to metalaxyl added to cornmeal agar

	Colony diameters (mm)			
<i>Pythium</i> sp.	Metalaxyl (ppm)		Percent of	
(no. of isolates)	0	1	control	
P. ultimum				
var. ultimum (3)	<85	0-Tr ^a	0	
var. sporangiiferum (4)	<85	0-Tr	0	
P. torulosum (4)	32-40	18-23	30-66	
P. aristosporum (3)	28-37	0-Tr	0	
P. irregulare (5)	67-<85	38-<85	45-<85	
Unknown (male)	72	0	0	
Unknown (female)	<85	0	0	
P. heterothallicum (male)	<85	Tr	0	
P. heterothallicum (female)	<85	Tr	0	

 $[^]a$ Tr=trace, i.e., a few hyphae were observed to have grown a very short distance from the agar block into the metalaxyl-amended medium.

Table 3. Sensitivities of different *Pythium* spp. to concentrations of metalaxyl added to cornmeal agar

Pythium spp.	Growth at percent of no-metalaxyl control ^a at indicated metalaxyl concentration (ppm)		
(no. of isolates)	0.01	0.1	1
P. ultimum			
var. sporangiiferum (2)	101	71	0
var. ultimum (1)	100	100	14
P. ultimum (3)	99-106	68-88	0
P. irregulare (8)	93-100	79-102	14-33
Unknown A (homothallic)	99	95	32
Unknown B (homothallic)	102	79	0
P. heterothallicum (male)	96	75	0
P. heterothallicum (female)	99	75	0
Unknown (heterothallic)	99	80	0

^a Measured after 48 hr at 20 C.

^bTr = trace, i.e., a few hyphae were observed to have grown a very short distance from the agar block into the *Pythium*-selective medium.

Table 4. Total population of *Pythium* in field soils and percentage of *Pythium* colonies from soil dilution plates that grew when transferred to a medium amended with metalaxyl at 1 ppm

Field	Total population (prop./g)	Growth on metalaxyl medium ^a (%)
1	346	70
2	380	79
3	515	22
4	758	86
5	628	86
6	430	51

^a Mircetich's (12) selective medium amended I ppm of metalaxyl. Growth based on presence of mycelium growing into the agar 48 hr after transfer of an agar block from a colony formed on the same medium without metalaxyl. At least 50 random colonies were tested from the dilution plates for each field sampled.

lead to different mixtures of the species in the fields. Different populations could account for response of wheat to metalaxyl in some fields but not in others. Metalaxyl might even be used as a tool to characterize the population of *Pythium* spp. in fields. The greater yields obtained in some cases provide encouragement that metalaxyl and related compounds could in some way contribute to the total management for control of Pythium.

LITERATURE CITED

- Bruin, G. C. A., and Edgington, L. V. 1981. Adaptive resistance in Peronosporales to metalaxyl. Can. J. Plant Pathol. 3:201-206.
- Bruin, G. C. A., and Edgington, L. V. 1982. Induction of fungal resistance to metalaxyl by ultraviolet irradiation. Phytopathology 72:476-480.
- Chamswarng, C., and Cook, R. J. 1983. Pythium spp. pathogenic to Pacific Northwest wheat and the influence of tillage practices on their populations. Phytopathology 73:811.
- Chamswarng, C., and Cook, R. J. 1985. Identification and comparative pathogenicity of Pythium species from wheat roots and wheatfield soils in the Pacific Northwest. Phytopathology 75:In press.
- Coffey, M. D., Klure, L. J., and Bower, L. A. 1984. Variability in sensitivity to metalaxyl of isolates of *Phytophthora cinnamomi* and *P. citricola*. Phytopathology 74:417-422.
- Cook, R. J., and Haglund W. A. 1982. Pythium root rot: A barrier to yield of Pacific Northwest wheat. Wash. State Coll. Agric. Res. Bull. XB0913. 20 pp.
- 7. Cook, R. J., Sitton, J. W., and Waldher, J. T. 1980. Evidence for *Pythium* as a pathogen of

- direct-drilled wheat in the Pacific Northwest. Plant Dis. 64:102-103.
- Cook, R. J., Zhang, B.-X., and Doerr, A. 1983.
 Failure of metalaxyl to control all of the *Pythium* population pathogenic on roots of Pacific Northwest wheat. (Abstr.) Phytopathology 73:957
- Davidse, L. C. 1980. Resistance to Ridomil in Phytophthora megasperma f. sp. medicaginis. Acta Bot. Neerl. 29:216.
- Davidse, L. C., Looijen, D., Turkensteen, L. J., and Van Der Wal, D. 1981. Occurrence of metalaxyl-resistant strains of *Phytophthora* infestans in Dutch potato fields. Neth. J. Plant Pathol. 87:65-68.
- Georgopoulos, S. G., and Grigoriu, A. C. 1981. Metalaxyl-resistant strains of Pseudoperonospora cubensis in cucumber greenhouses of southern Greece. Plant Dis. 65:729-731.
- Mircetich, S. M. 1971. The role of *Pythium* in feeder roots of diseased and symptomless peach trees and in orchard soils in peach tree decline. Phytopathology 61:357-360.
- Sanders, P. L. 1984. Failure of metalaxyl to control Pythium blight on turfgrass in Pennsylvania. Plant Dis. 68:776-777.
- Shew, H. D. 1984. In vitro growth response of *Phytophthora parasitica* var. nicotianae isolates to metalaxyl. Plant Dis. 68:764-766.
- Urech, P. A., Schwinn, F., and Staub, T. 1977. CGA 48988, a novel fungicide for the control of late blight, downy mildews and related soilborne diseases. Pages 623-631 in: Pests and Diseases. 1977 British Crop Protection Conference, Brighton, England.