Selective Medium for Isolation of *Pythium* spp. from Soil

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ABSTRACT

Conway, K. E. 1985. Selective medium for isolation of *Pythium* spp. from soil. Plant Disease 69:393-395.

A selective medium for isolation of Pythium spp. from soil and plant tissue was developed by incorporating the broad-spectrum fungicide etaconazole (1.125 EC) into 2% potato-dextrose agar amended with 300 μ g/ml of sodium ampicillin. Etaconazole at 17 μ g a.i./ml was as effective as a pimaricin-vancomycin-pentachloronitrobenzene medium in recovery of Pythium arrhenomanes from artificially infested soil and Pythium spp. from wheat, turf, forest nursery, and peanut field soil. Higher concentrations of etaconazole in the medium reduced the recovery of P. arrhenomanes and Pythium spp. from soil but allowed for mycelial growth in culture.

Pythium spp. cause damping-off of many plants and have a worldwide distribution (15). Pythium spp. have been associated with economic losses of peanuts (7), wheat (3), and tree seedlings in forest nurseries (18). The incidence and severity of Pythium spp. on these crops in Oklahoma have resulted in renewed interest in determining population dynamics of *Pythium* spp. in soil used for their production. Numerous media have been developed for selective isolation of Pythium spp. and Phytophthora spp. from soil (6, 13, 14, 17). Principles involved in the development of these media include either selective inhibition or enhancement of fungi (16). Selective inhibition is often based on the addition of several antibiotics, fungicides, and other inhibitory chemicals (4,5,9,12). Broad-spectrum fungicides have potential use in developing selective media. One fungicide, etaconazole (Vangard 1.125EC), has a broad spectrum of systemic activity against fungi in the classes Ascomycetes, Basidiomycetes, and Deuteromycetes but has little or no activity against Phycomycetes (1). This research was undertaken to determine if this activity could be incorporated into a selective medium for isolation of species of Pythium and/or Phytophthora from plant tissue and soil. A preliminary report has been published (2).

Journal Series No. 4496, Oklahoma Agricultural Experiment Station, Oklahoma State University, Stillwater.

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Accepted for publication 30 November 1984 (submitted for electronic processing).

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MATERIALS AND METHODS

Screening for fungicidal activity. A basal medium of 2% Difco potatodextrose agar (PDA) amended with 300 $\mu g/ml$ of sodium ampicillin was used throughout the experiment. Etaconazole (EANA) (Vangard 1.125EC) (1, 10, 100, and 1,000 μ g a.i./ml) was added to the basal medium after autoclaving. Radial growth of Pythium sp. from tomato roots, Rhizoctonia solani mating groups AG-1, AG-2, and AG-4, Sclerotium rolfsii from peanut, Fusarium moniliforme from sorghum, Phytophthora sp. from Taxus sp., Macrophomina phaseolina from arborvitae (Thuja sp.), Rhizopus sp. from soil, and Trichoderma harzianum from soil were compared at each concentration. Growth rates were determined at room temperature (25 C) by placing a 1-cm plug from the leading edge of a colony grown on PDA onto the selective medium and measuring the greatest colony diameter every 2 days until most of the cultures reached the edges of the petri dishes. Each treatment was replicated four times, and experiments were conducted at least twice with each organism.

Comparison of fungal growth. Excluding S. rolfsii, the same fungi used in the screening test were used for this comparison. Growth rates of these fungi on media amended with 100 and 1,000 μ g/ml of etaconazole (EANA [100] and EANA [1,000]) were compared with those grown on the modified pimaricinvancomycin-pentachloronitrobenzene (PVP) medium (17) developed for isolation of Phytophthora spp. and Pythium spp. from soil. PVP medium consisted of cornmeal agar (17 g/L), pimaricin (10 µg/ml), vancomycin (200 μ g/ml), and pentachloronitrobenzene (100 μ g/ml). Growth rates were determined as stated for the screening test, except a 0.7-cm inoculum plug was used and measurements were taken at 3, 5, 7, and 10 days. Each treatment was replicated four times.

Effect of EANA on Pythium spp. Growth of P. debaryanum, P. splendens, P. ultimum, P. pulchrum, P. vexans, P. mamillatum, P. aphanidermatum, P. irregulare (obtained from J. McRitchie. Florida Department of Plant Industries, Gainesville), and P. arrhenomanes were measured on agar amended with 34 $\mu g/ml$ of EANA (EANA [34]) to determine if EANA differentially affected species of Pythium.

Selective isolation from soil. Wheat cultivar TAM W 101 was grown in an unsterilized field soil artificially inoculated with mycelium of P. arrhenomanes. After 2 mo, the soil was air-dried, sieved (2-mm screen), and stored in a loosely covered bucket at room temperature.

Soil samples were collected from fields in different locations in Oklahoma that had been planted to turf (Stillwater), peanut (Perkins), forest nursery (Washington), and wheat (Stillwater). Each soil was thoroughly mixed, airdried, and sieved before analysis.

Media used included PVP and PDA amended with 300 μ g/ml of sodium ampicillin and two concentrations of EANA (17 and 34 μ g/ml). Two grams of soil from each location were added to 38 ml of a dilution suspension containing water agar (0.3%), CaCl₂ · 2H₂O (0.368 g/100 ml) and adjusted to pH 5.5 with 1% phosphoric acid. This suspension was mixed and 1 ml was spread on the selective medium in each of 10 petri dishes. After 24 hr of incubation at 25 C in the dark, the surface of agar in each petri dish was gently washed in a stream of water to remove soil and bacterial growth. Colonies of Pythium spp. were counted after additional incubation of 24. 48, 72, and 96 hr. The experiment was performed twice with infested soil and three times with each field soil. Representative colonies were transferred to PDA for verification and speciation of Pythium. Average number of Pythium colonies from 10 replicated petri dishes for each medium were analyzed using a one-tailed t test with the null hypothesis stated to ascertain if $\bar{X}_{PVP} \leq \bar{X}_{EANA}$.

RESULTS AND DISCUSSION

Screening for fungicidal activity. Greatest growth inhibition on the media was obtained when both EANA and sodium ampicillin were added to PDA after autoclaving. First evaluations of inhibition were made 2 days after inoculation. Concentrations of 1 and 10 μg/ml of EANA were not sufficient to

differentiate *Rhizopus* sp. and *R. solani* AG-4 from *Pythium* sp. Colony diameters of fungi grown on media amended with 100 and 1,000 µg/ml of EANA are shown in Table 1. After 2 days of incubation, EANA (100) inhibited growth of all fungi except *Pythium* sp., which covered the entire surface of the medium in the petri dishes. After 4 days of incubation,

Rhizopus sp. had also overgrown the medium. EANA (100) inhibited growth of R. solani AG-1, S. rolfsii, F. moniliforme, and T. harzianum for up to 10 days of incubation.

EANA (1,000) gave excellent inhibition of all fungi except *Pythium* sp. for up to 10 days. *R. solani* AG-1, *S. rolfsii*, *F. moniliforme*, *Rhizopus* sp., and *T.*

Table 1. Growth (cm) of selected fungi on a medium amended with etaconazole at two concentrations

Fungus		NA (10 s of gro	•	EANA (1,000) ^a (days of growth)				
	2	4	10	2	4	10	30	
Pythium sp.	8.2 ^b	8.5	8.5	3.8	7.9	8.5	8.5	
Phytophthora sp.	2.1	3.5	7.6	1.3	1.9	4.3	7.8	
Rhizoctonia solani								
AG-1	1.6	2.3	2.3	1.2	1.3	1.3	1.3	
AG-2	1.6	2.8	6.2	1.1	1.8	2.8	8.0	
AG-3	2.0	3.2	7.8	1.4	2.5	4.5	8.5	
Rhizopus sp.	3.8	8.5	8.5	1.0	1.0	1.0	1.0	
Trichoderma harzianum	1.0°	1.5	2.0	1.0	1.0	1.0	1.0	
Macrophomina phaseolina	1.7	4.0	8.5	1.2	2.4	3.9	8.2	
Fusarium moniliforme	1.0	1.2	2.0	1.0	1.0	1.4	4.5	
Sclerotium rolfsii	1.5	1.8	2.8	1.0	1.1	1.2	1.7	

^aBasal medium consisted of 2% potato-dextrose agar and 300 μ g/ml of sodium ampicillin with the addition of either 100 or 1,000 μ g/ml of etaconazole (EANA).

Table 2. Growth (cm) of selected fungi on three selective media 3 and 10 days after inoculation

Fungus	Days after inoculation									
		3	10							
	EANA (100) ^a	EANA (1,000) ^a	PVPb	EANA (100)	EANA (1,000)	PVP				
Pythium sp.	8.5°	4.0	8.5	8.5	8.5	8.5				
Phytophthora sp.	2.3	0.8	4.3	6.7	1.5	8.5				
Rhizoctonia solani										
AG-1	2.2	1.0	1.1	3.3	1.3	5.0				
AG-2	1.8	0.8	1.0	2.9	1.2	4.0				
AG-4	3.6	1.6	1.6	8.5	3.8	8.3				
Rhizopus sp.	8.3	0.9	0.7^{d}	8.5	1.0	0.7				
Trichoderma harzianum	1.9	0.7	2.5	4.5	0.7	8.5				
Macrophomina phaseolina	2.6	1.9	1.2	8.5	4.9	4.1				
Fusarium moniliforme	0.9	0.7	1.3	2.0	0.8	4.8				

^aBasal medium consisted of 2% potato-dextrose agar and 300 μ g/ml of sodium ampicillin with the addition of either 100 or 1,000 μ g/ml of etaconazole (EANA).

harzianum were inhibited for up to 30 days.

Comparison of fungal growth. After 3 days of incubation, *Pythium* sp. grew significantly more on EANA (1,000) and PVP than the other fungi (Table 2). Growth of *Pythium* sp. and *Rhizopus* sp. was similar on EANA (100) and was greater than that of other fungi. *Pythium* sp. and *Phytophthora* sp. were inhibited more on EANA (1,000) than on EANA (100) or PVP.

After 10 days of incubation on EANA (100), Pythium sp., R. solani AG-4, Rhizopus sp., and M. phaseolina had all completely covered the agar surface. On PVP, Pythium sp., R. solani AG-4, and T. harzianum also covered the agar surface. However, only Pythium sp. completely covered the agar surface of EANA (1,000) because the other fungi were still being inhibited.

Effect of EANA on Pythium spp. Seven of the nine Pythium isolates completely covered the amended agar (EANA [34]) surface (85 mm) by the third day. P. pulchrum and P. vexans did not reach full diameter until after the fifth day on both EANA (34) and PDA. There were no indications that EANA (34) inhibited mycelial growth of Pythium spp. tested. Pythium spp. in this experiment grew as rapidly as Pythium sp. from tomato grown on agar amended with EANA (100).

Selective isolation from soil. Although many selective media have been developed to isolate Pythium spp. from soil, PVP medium has been used preferentially. EANA (100) has been used routinely in our Plant Disease Diagnostic Laboratory (2) as an easy-to-prepare and inexpensive medium for isolation of *Pythium* spp. and Phytophthora spp. from diseased plant tissue. However, initial comparisons of isolation efficacy from soil indicated that 100 μ g/ml of EANA was too inhibitory to Pythium spp. to allow for comparison with PVP. Concentrations of 17 and 34 μ g/ml of EANA were incorporated into a basal medium and used for comparison with PVP (Table 3). Comparison of propagule counts from the selective media indicated that Pythium spp. were recovered on PVP

Table 3. Enumeration of Pythium spp. (propagules/g of soil) from various fields on three selective media

Medium	Source of soil for analysis													
	Artificially infested wheat ^a			Wheat		Peanut		Forest		Turf				
	1 ^b	2	1	2	3	1	2	3	1	2	3	1	2	3
PVP ^c	70	117 120	174 176	202 156°	182 116°	74 46	78 80	70 48	6 18	16 8	16	6 4	10 12	4 8
EANA (17) ^d EANA (34) ^d	126 82	102	170 	4 ^e	4 ^e	6°	18 ^e	10 ^e	6	6				

Soil was collected from a wheat field and artificially inoculated with Pythium arrhenomanes.

^bEach value is the average of the greatest diameter of each colony from two trials with four observations per trial.

^cValues of 1.0 indicate no growth from the plug used to inoculate the media.

^bPVP medium consisted of cornmeal agar (17 g/L), pimaricin (10 μ g/ml), vancomycin (200 μ g/ml), and pentachloronitrobenzene (100 μ g/ml).

^cEach value is an average of the greatest diameter of each colony from four observations.

^dValues of 0.7 indicate no growth from the plug used to inoculate the media.

^bEach column represents a trial, and each value is a mean of colonies per dish (multiplied by dilution factor of 20), 10 petri dishes per trial.

ePVP medium consisted of cornmeal agar (17 g/L), pimaracin (10 μ g/ml), vancomycin (200 μ g/ml), and pentachloronitrobenzene (100 μ g/ml).

^dBasal medium consisted of 2% potato-dextrose agar and 300 μ g/ml of sodium ampicillin with the addition of either 17 or 34 μ g/ml of etaconazole (EANA).

^eValues are significantly less (P = 0.05) than with PVP medium as determined by a one-tailed t test.

No colonies of Pythium spp. recovered.

medium in significantly greater numbers in only two of 14 trials compared with EANA (17). The concentration of etaconazole in EANA (34) was too inhibitory for comparable recovery of Pythium spp. from soil. Most cultures isolated from each medium belonged to the P. debaryanum-P. irregulare complex (8). In addition, P. ultimum was recovered on PVP from forest soil. Other Pythium spp. recovered on EANA (17) included P. ultimum from peanut soil and P. papillatum and P. torulosum from turf. It is interesting to note that most isolates from each medium possessed sphaerical sporangia. Neither P. myriotylum from peanut nor P. arrhenomanes from wheat were recovered on selective media. P. myriotylum was isolated from peanut tissue from plants in the same field (H. A. Melouk, personal communication); however, difficulty in recovery of P. myriotylum from soil has been reported previously (10). P. arrhenomanes infects wheat seedlings during late fall and winter and may not have been active when soil samples were collected from the fallow wheat field in September when soil temperatures were 30-40 C. Contamination by Mucoraceous fungi was not a problem with EANA or PVP media. Bacterial contamination was not as great on EANA media as on PVP, which permitted easier isolation of clean cultures of *Pythium* spp. from EANA.

The greatest benefit of using EANA (17) or EANA (34) was the length of time that *Pythium* spp. were restricted and countable. PVP medium was rapidly overgrown by *Pythium* spp. and had to be counted within 48 hr of incubation. EANA (17) and EANA (34) produced discrete colonies and were best counted

between 60 and 72 hr and 72 and 96 hr, respectively.

Mircetich and Kraft (11) noted a differential sensitivity between mycelia and spores of Pythium spp. to pimaricin. Greater recovery of Pythium spp. was accomplished using lower concentrations of pimaricin. A differential sensitivity for EANA was also noted in recovery of Pythium spp. from soil. Concentrations of EANA that were effective in recovery of Pythium spp. from plant tissue were too great for adequate recovery from soil. EANA (17) was as good as PVP for recovery of Pythium spp. from soil in 12 of 14 trials, allowed a longer period for counting of colonies, recovered a similar spectrum of Pythium spp. as PVP medium, and is less expensive to prepare.

ACKNOWLEDGMENTS

Contributions of etaconazole (Vangard 1.125EC [CGA-64251]) from Aithel McMahon, Ciba Geigy, Ardmore, OK 73401, and sodium ampicillin from Richard L. Sgroi, Bristol Laboratories, Syracuse, NY 13201, is gratefully acknowledged. I wish to thank Christine Fisher and Bryan Brown for excellent technical assistance and P. L. Claypool for statistical advice. Portions of this research were funded by a grant from the Oklahoma Department of Agriculture, Division of Forestry, Oklahoma City 73105.

LITERATURE CITED

- Anonymous. 1978. Ciba-Geigy Technical Release. Agricultural Division, Greensboro, NC.
- Conway, K. E. 1981. A selective medium for Pythium spp. for routine diagnostic laboratory isolation. (Abstr.) Phytopathology 71:868.
- Cook, R. J., Sitton, J. W., and Waldher, J. T. 1980. Evidence for *Pythium* as a pathogen of direct-drilled wheat in the Pacific Northwest. Plant Dis. 64:102-103.
- Elad, Y., and Chet, I. 1983. Improved selective media for isolation of *Trichoderma* spp. or *Fusarium* spp. Phytoparasitica 11:55-58.
- 5. Elad, Y., Chet, I., and Henis, Y. 1981. A selective

- medium for improving quantitative isolation of *Trichoderma* spp. from soil. Phytoparasitica 9:59-67
- Flower, R. A., and Hendrix, J. W. 1969. Gallic acid in a procedure for isolation of *Phytophthora* parasitica var. nicotiana and *Pythium* spp. from soil. Phytopathology 59:729-731.
- Garcia, R., and Mitchell, D. J. 1975. Interactions
 of Pythium myriotylum with Fusarium solani,
 Rhizoctonia solani, and Meloidogyne arenaria
 in pre-emergence damping-off of peanut. Plant
 Dis. Rep. 59:665-669.
- Hendrix, F. F., Jr., and Campbell, W. A. 1973. Pythiums as plant pathogens. Annu. Rev. Phytopathol. 11:77-98.
- Ko, W., and Hora, F. K. 1971. A selective medium for the quantative determination of *Rhizoctonia solani* in soil. Phytopathology 61:707-710.
- Lumsden, R. D., Ayers, W. A., and Dow, R. L. 1975. Differential isolation of *Pythium* species from soil by means of selective media, temperature, and pH. Can. J. Microbiol. 21:606-612.
- Mircetich, S. M., and Kraft, J. M. 1973. Efficiency of various selective media in determining *Pythium* populations in soil. Mycopathol. Mycol. Appl. 50:151-161.
- Papavizas, G. C., Morris, B. B., and Marois, J. J. 1983. Selective isolation and enumeration of *Laetisaria arvalis* from soil. Phytopathology 73:220-223.
- Rao, B., Schmitthenner, A. F., Caldwell, R., and Ellett, C. W. 1978. Prevalence and virulence of Pythium species associated with root rot of corn in poorly drained soil. Phytopathology 68:1557-1563.
- Ribeiro, O. K. 1978. A Source Book of the Genus *Phytophthora*. Cramer, Vaduz, Liechtenstein. 417 pp.
- Stanghellini, M. E. 1974. Spore germination, growth and survival of *Pythium* in soil. Proc. Am. Phytopathol. Soc. 1:211-214.
- Tsao, P. H. 1970. Selective media for isolation of pathogenic fungi. Annu. Rev. Phytopathol. 8:157-186.
- Tsao, P. H., and Ocana, G. 1969. Selective isolation of species of *Phytophthora* from natural soils on an improved antibiotic medium. Nature (Lond.) 223:636-638.
- Vaartaja, O. 1968. Pythium and Mortierella in soils of Ontario forest nurseries. Can. J. Microbiol. 14:265-269.