# Septoria Leaf Spot on Canavalia kauensis, a Native Hawaiian Bean

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#### ABSTRACT

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A Septoria sp. caused extensive infections and a leaf spot on Canavalia kauensis, a rare, recently discovered legume native to the island of Hawaii. The pathogen is referred to S. molleriana. Voucher specimens were deposited in the USDA National Fungus Collections Herbarium, Beltsville, MD.

Additional key words: Septoria canavaliae

Canavalia kauensis St. John, a beanlike legume, was newly discovered on the island of Hawaii in the early 1970s (4). Before its discovery, this species had apparently existed for years only as dormant seeds in the soil of the Kau Desert region of Hawaii Volcanoes National Park. Elimination of extreme browsing pressure from feral goats by construction of an experimental exclosure of about 1,000 m<sup>2</sup> permitted a dramatic recovery of the vegetation within. The resulting alterations, especially in soil moisture characteristics, presumably were favorable for germination of the C.

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kauensis seeds.

C. kauensis growth at first was vigorous, and a dense stand developed. Subsequently, these plants became uniformly infected with a leaf-spotting, fungal pathogen (Fig. 1). The spots were dark brown, circular to somewhat irregular, apparent on both leaf surfaces, and measured 0.5–1.5 mm in diameter (mean: 0.9 mm). Coalescence of spots sometimes produced larger necrotic areas. Defoliation apparently resulted from such uniform, extensive infections. Preliminary observations indicated that a species of Septoria was the pathogen.

#### MATERIALS AND METHODS

Semipermanent slides prepared of conidia mounted in lactophenol amended with cotton (aniline) blue rendered the nonstaining septa clearly discernible. Permanent slides prepared of leaf-spot tissue were made by following standard techniques for killing and fixing, dehydration in a tertiary butyl alcohol

series, and embedding in paraffin. Sections 15  $\mu$ m thick were cut, mounted, stained in aqueous safranin O, and counterstained with aniline blue (5). Observations and photographs were made with a Leitz Ortholux II microscope equipped with an Orthomat automatic camera using Kodak Plus-X pan 35-mm film.

The Septoria sp. was isolated from fresh tissue taken from leaf spots and cultured on potato-dextrose agar (PDA). Young, healthy host plants grown from seed in the greenhouse were inoculated by applying an aqueous conidial suspension (approximately  $5 \times 10^4$  conidia per milliliter) from the PDA cultures to the leaves. Newly inoculated plants were covered with plastic bags for 48 hr.

Voucher specimens of the Septoria sp. from naturally infected C. kauensis leaves were deposited in the USDA National Fungus Collections Herbarium, Beltsville, MD.

#### RESULTS AND DISCUSSION

Observations of leaf-spot tissue revealed pycnidia and associated pycnidiospores (conidia) typical of Septoria sp. (Fig. 2). The pycnidia were approximately 80  $\mu$ m in diameter, and the conidia were (15)20-40(46)  $\mu$ m long × 1.5-4.5  $\mu$ m wide, with (1)2-5(6) septa.

Leaf spots on *C. kauensis* resulting from the greenhouse inoculations were somewhat larger (0.5–2.0 mm diam; mean: 1.43 mm) than were those on



Fig. 1. Typical Septoria leaf-spot symptoms observed on Canavalia kauensis in Hawaii.



Fig. 2. Pycnidium of Septoria cf. molleriana from a leaf spot on Canavalia kauensis. (Scale bar represents 20  $\mu$ m.)

naturally infected leaves in the field (0.5-1.5; mean: 0.9 mm), but were otherwise comparable. Conidia similar to those with which the leaves had been inoculated were readily obtained from

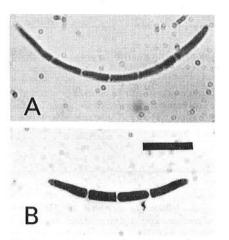


Fig. 3. Septoria spp. conidia: (A) S. canavaliae from a leaf spot on Canavalia ensiformis; (B) S. cf. molleriana from a leaf spot on C. kauensis. (Scale bar represents  $10 \mu m$ .)

these leaf spots, and the fungus was recultured on PDA.

S. canavaliae Lyon ex Sydow, which had previously been collected as a new species on the Hawaiian island of Oahu from leaf spots on jack bean (C. ensiformis (L.) DC.) (1,6,7), was initially considered to be the most likely species to which the new Septoria sp. should be referred. Comparison of conidia from the C. kauensis leaf spots with those of

herbarium specimens from infected *C. ensiformis* leaves (7), and also with the original description of *S. canavaliae* (3), however, indicated easily recognizable differences in conidial morphology (Fig. 3).

Furthermore, inoculation of *C. kauensis* and *C. ensiformis* with the new *Septoria* sp. pathogen consistently resulted in moderate to abundant infections on *C. kauensis* following about 15 days of incubation, whereas the inoculated plants of *C. ensiformis* remained entirely free of infections. *C. kauensis* was not inoculated with *S. canavaliae* because no live, infected *C. ensiformis* plants were available as a source of inoculum.

The Septoria sp. from C. kauensis is referred to S. molleriana Bres. & Roum. (2), which was also described from a Canavalia sp. (C. obtusifolia St. Thome) and whose description (pycnidia 70-80 μm in diam; conidia three-septate, 25-30  $\mu$ m  $\times$  3–4  $\mu$ m) closely approximates that of the Septoria sp. discussed here. Attempts to locate herbarium specimens of this Septoria sp. were not successful. It is commonly recognized that numerous new Septoria spp. have in the past been established primarily on the basis of the host with which they were associated, with little support from cross inoculation or morphological stability tests. Although the Septoria sp. attacking C. kauensis may have been described as a new species, perpetuation of this somewhat arbitrary classification method was not considered advantageous.

Although the natural, long-term population dynamics of *C. kauensis* are not known, definite periodic cycles of vigorous development followed by dying back of the stands have been observed for several years. The extent to which these cycles are correlated with Septoria leaf spot severity, or with other possible pathogenic or ecologic influences, is not known and must await further investigation.

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