Effect of Mineral Fertilizers on the Incidence of Blossom-end Rot of Watermelon

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ABSTRACT

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Potassium and phosphorus fertilizers had no effect on the percentage of blossom-end rot of watermelon in either medium-textured sandy loam soil or in fine-textured loam soil. In sandy loam soil, in 1974, the percentage of blossom-end rot in the Charleston Gray watermelon was significantly reduced by 8.06 and 8.24% in calcium (20 kg/100 m² gypsum)- and nitrogen (5 kg/100 m² urea [44-45% N])-treated plots, respectively. In loam soil, in 1975, treatment with calcium reduced the percentage of blossom-end

rot in cultivars Crimson Sweet and Charleston Gray. In 1976, in loam soil the incidence of blossom-end rot in Crimson Sweet also was significantly reduced and marketable yield was increased with increasing levels of gypsum. In this experiment the increasing levels of nitrogen had no effect on disease incidence, but significantly increased the yield of marketable melons.

Additional key words: Citrullus lanatus, calcium, fruit disorder.

"Blossom-end rot" is a nonparasitic disease of watermelon (Citrullus lanatus [Thunb.] Matsum & Nakai) recently described in southern Italy (5). In commercial watermelon plantings in the climatic and soil conditions of Puglia and Lucania regions, the disease (Fig. 1) first appears around the middle of July, and the incidence increases until harvest time. Commonly 60–80% of fruits are affected by blossom-end rot in the period from the end of July through August, a period characterized by high temperatures, low humidity, and warm, dry winds. In field trials, cultivars with cylindrical or sub-cylinderical fruit generally are susceptible, and spherical-fruited cultivars usually possess a complete or high resistance to the disease (6).

In physiological disorders of fruits of vegetable crops, such as blossom-end rot of the tomato (*Lycopersicon esculentum Mill.*) water stress and the availability of mineral elements play an important role in influencing disease incidence. In field experiments conducted in southern Italy the frequency of furrow irrigation did not significantly affect the percentage of blossom-end rotted watermelon fruits (6).

In this paper we report on the influence of mineral fertilizer elements on the incidence of blossom-end rot of watermelon.

MATERIALS AND METHODS

Field trials were carried out on medium-textured sandy loam soil near Lecce and on fine-textured loam soil near Policoro. The chemical and textural characteristic of these two soils prior to fertilization are listed in Table I.

Lecce, 1974. The trial included 16 treatments obtained by combining factorially two levels of the following fertilizers: nitrogen (0 and 2.5 kg/100 m² urea [44-45% N]); phosphorus (0 and 5 kg/100 m² triple-superphosphate [45-48% P₂O₅]; potassium (0 and 5 kg/100 m² potassium sulfate [48-52% K₂O]) and, gypsum (0 and 20 kg/100 m²). The design of this experiment was a randomized block in which the treatments were replicated four times. The area of each plot-treatment was 21 m² and included 25 plants. Cultivar Charleston Gray, which is highly susceptible to the disease, was used.

Policoro, 1975. Two watermelon cultivars, the highly susceptile Charleston Gray and the moderately resistant Crimson Sweet, were used. Plots of each cultivar were treated with two levels of four

fertilizers, a total of 16 treatments. A split-plot design was used arranging phosphorus levels (0 and 7 kg/100 m² superphosphate [18–20% P_2O_5]) in whole plots, potassium levels (0 and 5 kg/100 m² potassium sulfate [48–52% $K_2O_{\rm J}$) in split plots, nitrogen levels (0 and 5 kg/100 m² of urea [44–45% N]) in split-split plots, and calcium levels (0 and 20 kg/100 m² gypsum) in split-split-plots. The area of each split-plot unit was 24 m² and each had 30 plants.

Policoro, 1976. In this trial the Crimson Sweet watermelon was used with nine treatments obtained by combining three levels of two fertilizers. A split-plot design was used in which three nitrogen levels (0 kg/100 m², 5 kg/100 m² and 10 kg/100 m² of urea [44-45% N]) in whole plots and three calcium levels (0 kg/100 m², 20 kg/100 m², and 40 kg/100 m² gypsum) in split plots. The area of each of the split-plot units was 56 m² and included 60 plants. In all the experiments, plots were separated by a strip of fallow soil at least 5 m wide. Treatments of phosphorus, potassium, and calcium were incorporated into the soil about 1 wk before sowing. Nitrogenous fertilizer was applied twice, half before sowing and half 2 mo after the plants had emerged. Furrow irrigation was applied at 6-day intervals. At harvest the total yield was counted and weighed. The number of fruits with blossom-end rot was expressed as a percentage of the total fruits harvested.

RESULTS

Potassium and phosphorus fertilizers had no effect on the incidence of blossom-end rot in both types of soils. Nitrogen affected the incidence of blossom-end rot only in sandy loam soil that was low in total N. Calcium applications reduced the number of fruits with blossom-end rot in all experiments.

Medium-textured sandy loam (Lecce, 1974). The percentage of Charleston Gray watermelon fruits with blossom-end rot was reduced (P=0.05) by 8.06 and 8.24% (total means) in gypsum-and urea-treated plots, respectively (Fig. 2).

Fine-textured, loam soil (Policoro 1975). With Crimson Sweet, but not with the Charleston Gray watermelons, there was a significant (P=0.05) interaction between superphosphate and gypsum applications that resulted in lower blossom-end rot percentages with increasing levels of P. In the 20 kg/100 m² gypsum plots, blossom-end rot was 15.92 and 10.33% for P₀ (0 kg/100 m² and P₁ (7 kg/100 m² superphosphate) levels of phosphorus, respectively. Applications of gypsum resulted in a statistically significant (P=0.01) reduction of the disease (Table 2) in Crimson Sweet and

TABLE 1. Chemical and textural characteristics of the soils used in studies of blossom-end rot of watermelon at Lecce and Policoro, Italy

Location and soil type	Sand (%)	Silt (%)	Clay (%)	Soluble carbonates ^a (%)	Organic matter ^b (%)	Total N° (%)	Available K ^d (ppm)	Available P° (ppm)	рН
Lecce Sandy loam	81.4	5.1	13.5	0.75	1.1	0.67	108	16.00	7.5
Policoro	01.4	5.1	15.5	0.75		0.07	100	10.00	7.5
loam	42.5	33.6	23.9	6.10	2.20	1.38	363	1.25	7.6

Extraction procedures:

a = ammonium oxalate

b = Walkley-Black

c= Kjeldahl

d= ammonium acetate

c = Bray

Charleston Gray watermelons. Applications of urea had no effect on disease incidence.

Policoro, 1976. In view of the 1974 and 1975 results, an experiment was carried out in 1976 on the loam soil at Policoro to test more throughly the influence of increasing levels of calcium and nitrogen on incidence of the disease in the Crimson Sweet watermelon. The incidence of blossom-end rot was again reduced statistically with increasing levels of gypsum; the percentages of diseased fruits were 34.02, 22.23, and 19.53% for 0 kg/100 m², 20 kg/100 m², and 40 kg/100 m² gypsum, respectively. The average marketable yield was 419.2 kg/100 m², 491.7 kg/100 m² and 514 kg/100 m² for plots that had received 0 kg/100 m², 20 kg/100 m², and 40 kg/100 m² gypsum, respectively. In this experiment the different levels of nitrogen had no significant effect upon disease incidence although there was a statistically significant increase in marketable yield in plots receiving higher rate of nitrogen fertilization (Fig. 3).

DISCUSSION

The results of these experiments indicate that some mineral fertilizer elements affect the incidence of blossom-end rot of watermelon both in the highly susceptible cultivar Charleston Gray and the moderately resistant cultivar Crimson Sweet.

Additions of potassium produced no reduction in the amounts of blossom-end rot, either in medium textured sandy loam at Leece or in the fine textured loam at Policoro. The interaction between superphosphate and gypsum applications in the 1975 Policoro trials seems to be due to the additive effect of the 30% CaSO₄

Fig. 1. Blossom-end rot of watermelon.

content of the superphosphate rather than to the phosphorus supplied by this fertilizer. It would seem, therefore, that applications of these elements in soils with potassium and phosphorus content near that of our soils (Table 1) will not reduce the incidence of blossom-end rot.

In our tests, nitrogen, suppplied as urea, produced no increase in blossom-end rot even when applied at the high level of $10 \, \text{kg}/100 \, \text{m}^2$. On the contrary, urea, with the sandy loam soil low in total N content, applied at the rate of 2.5 kg/100 m², resulted in a statistically significant reduction of blossom-end rot. The reduced incidence of the disease following the applications of calcium to the soil in these experiments with watermelon agrees with numerous observations of its effect on blossom-end rot of tomato and indicates that this disorder is related to the availability of calcium in

TABLE 2. Influence of field applications of calcium (as gypsum) on the blossom-end rot (B.E.R.) and marketable yield of cultivars Crimson Sweet and Charleston Gray watermelon grown in loam soil, Policoro, Italy, in 1975

	Crimso	n Sweet	Charleston Gray		
Gypsum (kg/100 m ²)	Yield (kg/100 m ²)	Fruits with B.E.R. (%)	Yield (kg/100 m ²)	Fruits with B.E.R. (%)	
0	457.1	19.6	305.3	57.7	
20	521.1	13.1	328.5	54.0	
LSD $(P = 0.05)$	49.9	2.6	30.1	3.6	
(P = 0.01)	67.5	3.6	40.7	4.9	

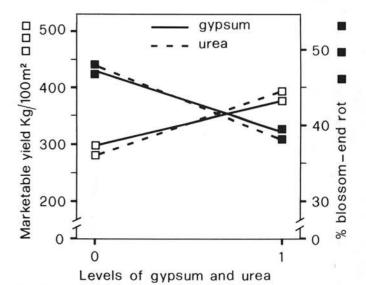


Fig. 2. Influence of field applications of calcium (as gypsum) and nitrogen (as urea) on the blossom-end rot and the marketable yield of watermelon cultivar Charleston Gray grown in sandy loam soil, Lecce, Italy, in 1974. Legend: gypsum $0=0~kg/100~m^2$, $1=20~kg/100~m^2$); and urea $(0=0~kg/100~m^2,~1=5~kg/100~m^2)$.

the soil (10,12) and the calcium content of the fruit (22,26), and that calcium has a fundamental role in the etiology of blossom-end rot in both tomato and watermelon.

Calcium, as calcium pectate, is a constituent of the middle lamella (7) and thus contributes to cell-wall stability. More recently, however, there is evidence that calcium in plants is more directly related to interactions with the cell membranes than with cell walls. Marinos (20) reports that calcium deficiency in the shoot apex of barley initially causes a break up of the nuclear envelope and the plasma and vacuolar membranes, the disorganization of mitochondria and Golgi apparatus and, later, the weakening of cell walls eventually followed by cell necrosis. The involvement of Ca in membrane maintenance and function has a great influence on a number of enzymatic activities (eg, ATPase [9] and phospholipase [8]) and on the selective ion-absorption mechanism (27). The effects of calcium on mitochondria (20) also explains its influence on the active uptake of other ions (eg, phosphate, nitrate, chloride, and bromide [15,16] and potassium [17,18]) since uptake is a respiration-dependent phenomenon.

In studies of tomatoes, it has been shown that factors other than mineral element deficiency interact to produce blossom-end rot. An important role is attributed to water stress in the plant. In normal tomato fruits, the tissues of the distal end would be more permeable to water because of poor orientation of cellulose molecules of the cell wall (13). Thus, it would seem that water stress due to high transpiration or low soil moisture availability may increase the sensitivity of this portion of the fruit and induce

collapse and necrosis of the cells. To fully appreciate the importance of the combined effects of Ca deficiency and water stress in the etiology of blossom-end rot, one has to consider that in normal tomato plants only one-half of the Ca²⁺ is water-insoluble whereas in calcium-deficient plants nearly all the Ca2+ is water-insoluble (21). Thus, since Ca is poorly translocated (1), it would be expected that the apex of plant organs, including actively growing fruits, would tend to be deficient in Ca and would be more severely affected since the Ca requirement for maintaining integrity of fruit cells is high. Furthermore, the dramatic influence of Ca on rapidly growing fruits is increased because the translocation of this element depends on the water flow into the fruit. Gerard and Hipp (14), report that the incidence of blossom-end rot in tomato is highly correlated with the number of hours above (90F) or high evaporative conditions which determine, together with a poor input of Ca, a deficit in the water balance of the fruit. Consequently, a further lowering of blossom-end rot in watermelon, which is in agreement with the tomato findings, could

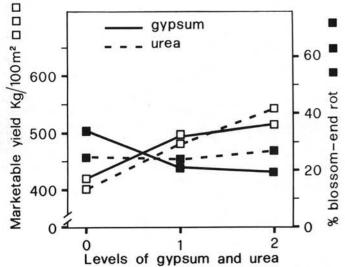


Fig. 3. Influence of field applications of calcium (as gypsum) and nitrogen (as urea) on the blossom-end rot and marketable yield of watermelon cultivar Crimson Sweet grown in loam soil, Policoro, Italy, in 1976. Legend: gypsum $(0 = 0 \text{ kg}/100 \text{ m}^2, 1 = 20 \text{ kg}/100 \text{ m}^2, \text{and } 2 = 40 \text{ kg}/100 \text{ m}^2)$; urea $(0 = 0 \text{ kg}/100 \text{ m}^2, 1 = 5 \text{ kg}/100 \text{ m}^2, \text{and } 2 = 10 \text{ kg}/100 \text{ m}^2)$.

be obtained by foliar spraying with calcium chloride during periods of high Ca requirement of the plant (10,12) and applying sprinkler irrigations during highly evaporative weather conditions (14).

It appears that in studies of such physiologic disorders, the effect of mineral elements may vary according to environmental conditions (low or high evaporative climate, open field or greenhouse) and to the substrata (field soil or nutrient solution media) in which plants are tested. In our field trials, blossom-end rot of watermelon was not increased by high levels of nitrogen (urea). This differs from what is generally observed with tomato (4,11,12,19,23). However, in some cases, applications of nitrogen either caused a reduction (2), or did not increase (25), the disease. The different responses to nitrogen applications in tomato may be due to differences in environment, substrata for growth of plants, and cultivars used. However, the effect that nitrogen has shown in our experiments also may be attributed to the metabolic differences between tomato and watermelon.

In conclusion, the lack of conclusive information on the numerous nutritional, water supply (6), varietal (6) and, (perhaps) hormonal factors (see Mullison and Mullison [21], Struckmeyer [24], and Burström [3]), and their interactions, makes it impossible to obtain complete control of the blossom-end rot of watermelon in commercial fields. Nonetheless, in order to significantly reduce the incidence of the disease, balanced fertilizations, applications of gypsum, frequent irrigation, and the use of spherical-fruited resistant cultivars are recommended.

LITERATURE CITED

- BIDDULPH, O., R. CORY, and S. BIDDULPH. 1959. Translocation of calcium in the bean plant. Plant Physiol. 34:512-519.
- BROOKS, C. 1914. Blossom-end rot of tomatoes. Phytopathology 4:345-374.
- BURSTRÖM, H. 1952. Studies on growth and metabolism of roots.
 VII. Calcium as a growth factor. Physiol. Plant. 5:391-402.
- CHAMBERLAIN, E. E. 1933. Blossom-end rot of tomatoes. N. Z. J. Agric. 46:293-296.
- CÎRULLI, M. 1974. II "marciume apicale" dell' Anguria. Phytopathol. Mediterr. 13:77-81.
- CIRULLI, M. 1974a. Primi risultati sulla resistenza varietale e sull' influenza della frequenza irrigua nel "marciume apicale" dell' Anguria. Phytopathol. Mediterr. 13:82-86.
- CONRAD, C. M. 1926. A biochemical study of the insoluble pectic substances in vegetables. Am. J. Bot. 13:531-547.
- DAVIDSON, F. M., and C. LONG. 1958. The structure of the naturally occurring phosphoglycerides. 4. Action of cabbage leaf phospholipase D on ovolecithin and related substances. Biochem. J. 69:458-466.
- DODDS, J. A. A., and R. J. ELLIS. 1966. Cation-stimulated ATPase activity in plant cell walls. Biochem. J. 101:31p.
- EVANS, H. J., and R. V. TROXLER. 1953. Relation of calcium nutrition to the incidence of blossom-end rot in tomatoes. Proc. Am. Soc. Hortic. Sci. 61:346-352.
- FOSTER, A. C. 1939. Effect of environment on metabolism of tomato plants as related to development of blossom-end rot of the fruit. (Abstr.) Phytopathology 29:7.
- GERALDSON, C. M. 1957. Control of blossom-end rot of tomatoes. Proc. Am. Soc. Hortic. Sci. 69:309-317.
- GERARD, C. J., and W. R. COWLEY. 1966. A study of blossom-end rot of pear shaped tomatoes. Tex. Agric. Exp. Stn. Mimeo. Pap. 814.
- GERARD, C. J., and B. W. HIPP. 1968. Blossom-end rot of "Chico" and "Chico grande" tomatoes. Proc. Am. Soc. Hortic. Sci. 93:521-531.
- HOOYMANS, J. J. M. 1964. The role of calcium in the absorption of anions and cations by excised barley roots. Acta Bot. Neerl. 13:507-540.
- HYDE, A. H. 1966. Nature of the calcium effect in phosphorus uptake by barley roots. Plant Soil 24:328-332.
- JACOBSON, L., R. J. HANNAPEL, D. R. MOORE, and M. SCHAEDLE. 1961. Influence of calcium on the selectivity of the ion absorption process. Plant Physiol. 36:58-61.
- JACOBSON, L., D. R. MOORE, and R. J. HANNAPEL. 1960. Role of calcium in absorption of monovalent cations. Plant Physiol. 36:352-358.
- LEOPOLD, A. C., and F. S. GUERNESEY. 1953. The effect of nitrogen upon fruit abnormalities in the tomatoes. Proc. Am. Soc. Hortic. Sci. 61:333-338.
- 20. MARINOS, N. G. 1962. Studies on submicroscopic aspect of mineral

- deficiences. I. Calcium deficiency in the shoot apex of barley. Am. J. Bot. 49:834-841.
- MULLISON, W. R., and E. MULLISON. 1948. Effects of several plant growth-regulators on fruit set, yield and blossom-end rot of six tomato varieties grown under high temperatures. Bot. Gaz. 109:501-506.
- NIGHTINGALE, G. T., R. M. ADDOMS, W. R. ROBBINS, and L. G. SCHERMERHORN. 1931. Effects of calcium deficiency on nitrate absorption and on metabolism in tomato. Plant Physiol. 6:605-631.
- RALEIGH, S. M., and J. A. CHUCKA. 1944. Effect of nutrient ratio and concentration on growth and composition of tomato plants and on the occurrence of blossom-end rot of the fruit. Plant Physiol.
- 19:671-678.
- STRUCKMEYER, B. E. 1949. Effect of alphanaphalene acetamide upon the anatomical structure of cocklebur grown in a nutrient medium deficient in calcium. Bot. Gaz. 111:130-139.
- STUCKEY, H. P., and J. C. TEMPLE. 1911. Tomatoes: Part II. Blossom-end rot. Bull. Georgia Agric. Exp. Stn. 96:69-91.
- TAYLOR, G. A., and C. B. SMITH. 1957. The use of plant analysis in the study of blossom-end rot of tomato. Proc. Am. Soc. Hortic. Sci. 70:341-349.
- Von GUTTENBERG, H., and A. BEYTHIEN. 1951. Uber den Einflüss von Wirkstoffen auf die Wasserpermeabilität des Protoplasmas. Planta 40:36-69.