# Parasitization of Pine Stem Rust Fungi by Monocillium nordinii

A. Tsuneda and Y. Hiratsuka

National Research Council of Canada visiting fellow and research scientist, respectively, Northern Forest Research Centre, Canadian Forestry Service, Environment Canada, 5320-122 Street, Edmonton, Alberta, Canada T6H 3S5.

Accepted for publication 17 May 1980.

#### ABSTRACT

TSUNEDA, A., and Y. HIRATSUKA. 1980. Parasitization of pine stem rust fungi by Monocillium nordinii. Phytopathology 70:1101-1103.

Monocillium nordinii that occurred in sori of Cronartium coleosporioides and Endocronartium harknessii in Alberta, Canada, was a destructive mycoparasite of these pine stem rust fungi. Detailed study of the parasitic process in the E. harknessii-M. nordinii interaction showed that hyphae of M. nordinii grew between the surface wart layers of rust spores and that swollen appressoriumlike bodies often were formed where contact was made. The host cytoplasm occasionally developed a papilla beneath the area of contact during the early stages of parasitism; however, the host cell

eventually lost most of its cytoplasm, except oil bodies, in most cases. Penetration of the rust spores by the *Monocillium* hyphae usually appeared to occur after the host cells were killed. *Monocillium* generally formed many conidiophores and conidia on the host spores. Eventually the surface morphology of the infected rust spores was greatly degraded. It is suggested that antifungal metabolites of *M. nordinii* are involved in this parasitic process.

Additional key word: mycoparasitism.

Western gall rust (caused by Endocronartium harknessii [J. P. Moore] Y. Hiratsuka) and stalactiform blister rust (caused by Cronartium coleosporioides Arth.) are major pine stem rusts in Western Canada that cause severe damage to hard pines (7,16). Outbreaks of these rusts have been severe in some years, but usually there is an interval of several years between major outbreaks. Field surveys in Alberta in 1977-1979 revealed that not only abiotic factors, but also some mycoparasites play important roles in suppressing the rust epidemics. In 1979, for example, Scytalidium uredinicola Kuhlman et al destroyed more than 80% of E. harknessii galls in some localities in Alberta (8,13). Cladosporium gallicola Sutton also occurred frequently on the rust and reduced its inoculum potential (12). Although Monocillium nordinii (Bourchier) W. Gams (= Cephalosporium nordinii Bourchier) (6) occurred less frequently than these mycoparasites, it also was found in sori of both E. harknessii and C. coleosporioides, where it appeared to produce strong antifungal metabolites. This fungus commonly has been isolated from the heartwood of lodgepole pine, Pinus contorta Dougl. var. latifolia Engelm. (4); however, this is the first report on its mycoparasitic activity.

### MATERIALS AND METHODS

Rust sori of C. coleosporioides and E. harknessii on lodgepole pine were collected at several locations in Alberta. All the collected samples of E. harknessii sori with natural infections of M. nordinii also were colonized by other mycoparasites, usually C. gallicola, but most samples of C. coleosporioides parasitized by M. nordinii were free from other mycoparasites. Sori of C. coleosporioides that were naturally infected by M. nordinii and those of E. harknessii that were artificially inoculated with M. nordinii were used for scanning electron microscopy (SEM). Artificial inoculation was performed by spraying young sporulating rust galls with about 1 ml of Monocillium spore suspension (about 10<sup>7</sup> spores per milliliter of distilled water) per gall. These galls were incubated at 22-25 C in sterile beakers lined with moist filter paper. Small pieces (about  $5 \times 5 \times 5$  mm) taken from samples of both rusts were fixed and dehydrated by either air drying or freeze-drying, as described previously (12,15). The dried materials were shadowed with gold, and at least six pieces of each rust were examined with a Cambridge Stereoscan S<sub>4</sub> scanning electron microscope.

For light microscopy, 0.5 ml of the spore suspension of M.

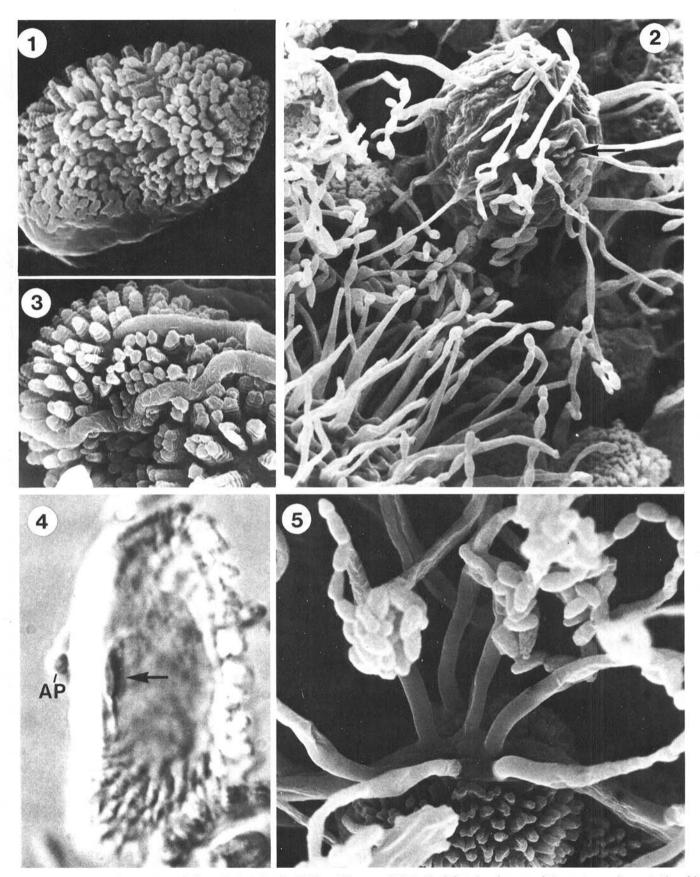
nordinii was spread evenly on a cellophane membrane disk (8-cm diameter) that was placed on 2% water agar in a petri dish. Mature spores of *E. harknessii* were dusted onto the membrane, and the petri dish was kept at 20 C in the dark. After incubation for 48 or 96 hr, the cellophane membrane was removed, cut into strips, mounted in water or in cotton blue-lactophenol on a microscope slide, and examined with a Leitz Orthoplan microscope with Nomarski interference-contrast optics.

## RESULTS AND DISCUSSION

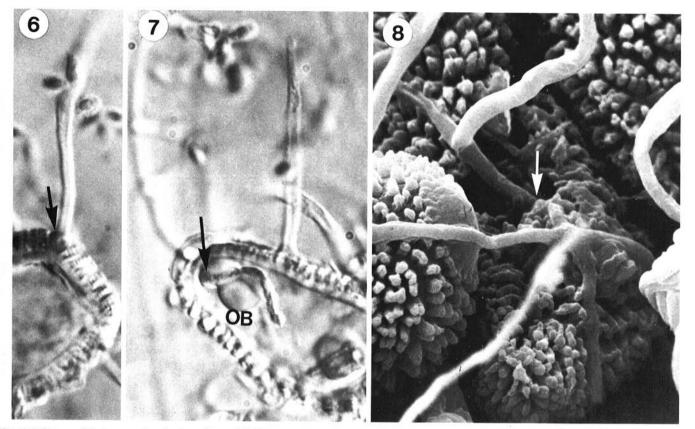
Sori of *C. coleosporioides* that were naturally infected by *M. nordinii* had a whitish, fluffy appearance because of the numerous conidiophores and conidia produced by the mycoparasite. Many annulated warts ornamented the surface of noninfected aeciospores (Fig. 1), but the warts of infected spores appeared degraded. In extreme cases, warts were not discernible, and hyphae of *M. nordinii* were covered with an amorphous material that appeared to be the degraded wart material (Fig. 2). Wart degradation may be either an autolytic process induced in the host by the parasitic attack, the result of activity of the cell wall hydrolyzing enzymes of the mycoparasite, or both. Similar wart degradation occurs commonly in *E. harknessii* spores parasitized by other mycoparasites (12,13).

Hyphae of M. nordinii grew profusely in sori of E. harknessii that were sprayed with M. nordinii spore suspensions. The mycoparasite hyphae grew between the surface wart layers of the rust spores and were appressed to the host cell wall (Fig. 3). Swollen appressoriumlike bodies often were formed at these contact points (Fig. 4). Host cytoplasm occasionally formed a papilla beneath the appressoriumlike body (Fig. 4), but only where contact was made during the very early stages of infection. Papillae are not induced in the cytoplasm of E. harknessii by C. gallicola or S. uredinicola (12,13). We speculate, therefore, that E. harknessii develops the structure in response to a specific metabolite released by M. nordinii. Similar structures have been reported for fungal (9-11, 14, 15) and vascular plant cells (1,5) infected by fungal parasites. These structures are thought either to have a protective role in delaying or preventing penetration by the parasites (5,15) or to represent a sign of necrosis without any positive role in host resistance (11). Induction of a papilla in E. harknessii by M. nordinii occurs only while the host cells are fully viable, suggesting that the structure is not simply a product of necrosis.

Formation of conidiophores and conidia by M. nordinii occurred frequently on the host spores (Fig. 5). The conidiophores



Figs. 1-5. Host-parasite interactions of pine stem rust fungi and Monocillium nordinii. 1, Noninfected aeciospore of Cronartium coleosporioides with normal annulated warts on the surface (×3,000). 2, Portion of a C. coleosporioides sorus showing an advanced stage of natural infection by M. nordinii. Note a rust spore showing a severely degenerated surface with a few warts (arrow) of nearly original shape (×1,800). 3, M. nordinii hyphae growing between the surface wart layers of a peridermioid teliospore of Endocronartium harknessii (×5,000). 4, Papilla (arrow) in cytoplasm of E. harknessii formed in response to contact by an appressoriumlike body (AP) of M. nordinii (Nomarski light optics ×3,500). 5, Formation of conidiophores and conidia by M. nordinii on a spore of E. harknessii during a late stage of infection (×3,600).



Figs. 6-8. Spores of Endocronartium harknessii parasitized by Monocillium nordinii. 6, Conidiophore of the parasite swollen at the base (arrow) in contact with the host (Nomarski light optics ×2,800). 7, E. harknessii spore penetrated by an M. nordinii hypha (arrow). Oil bodies (OB) remain within the host hypha (Nomarski light optics ×2,800). 8, Severely degenerated E. harknessii spore penetrated by an M. nordinii hypha (arrow) (×4,000).

usually became swollen at the base and anchored themselves between the warts of the host spore (Fig. 6). Although penetration was common (Figs. 7 and 8), it rarely occurred from appressoriumlike bodies and was not observed at the points where the papillae were present. Penetration appeared to occur usually after the host cells were killed. The host cytoplasm eventually lost most of its contents, except oil bodies (Fig. 7).

Similar destructive mycoparasitism (3) also has been reported in the interactions between *E. harknessii* and its two other mycoparasites, *C. gallicola* (12) and *S. uredinicola* (13). The detailed mode of parasitism, especially the importance of penetration, however, varies in these interactions: *Cladosporium gallicola* often penetrates and kills living host spores, *M. nordinii* appears to penetrate only after the host is killed, and no penetration is involved in the parasitism by *S. uredinicola*. *Cladosporium gallicola* forms appressoria and often develops penetration pegs from them, while penetration seldom occurs from the appressoriumlike bodies of *M. nordinii*.

Antifungal metabolites of M. nordinii apparently are involved in its parasitic process on pine stem rusts. Our preliminary experiments indicated that the ether extract of an M. nordinii culture medium had a strong inhibitory effect on the rust fungi as well as other fungi such as Ceratocystis ulmi (Buism.) C. Moreau, the pathogen of Dutch elm disease. Subsequently six compounds, mainly consisting of monorden ( $C_{18}H_{17}O_6Cl$ ) and a new antibiotic, monocillin I ( $C_{18}H_{18}O_6$ ), were isolated from liquid culture of M. nordinii (2).

# LITERATURE CITED

- AIST, J. R. 1976. Papillae and related wound plugs of plant cells. Annu. Rev. Phytopathol. 14:145-163.
- AYER, W. A., S. P. LEE, A. TSUNEDA, and Y. HIRATSUKA. 1980. The isolation, identification, and bioassay of the antifungal metabolites produced by *Monocillium nordinii*. Can. J. Microbiol. 26:766-773.

- 3. BARNETT, H. L., and F. L. BINDER. 1973. The fungal host-parasite relationship. Annu. Rev. Phytopathol. 11:273-292.
- BOURCHIER, R. J. 1961. Two new imperfect fungi from the heartwood of living lodgepole pine. Can. J. Bot. 39:1781-1784.
- BRACKER, C. E., and L. J. LITTLEFIELD. 1973. Structural concepts of host-pathogen interfaces. Pages 159-317 in: R. J. W. Byrde and C. W. Cutting, eds. Fungal Pathogenicity and the Plant's Response. Academic Press, New York. 499 pp.
- GAMS, W. 1971. Cephalosporium-artige Schimmelpilze (Hyphomycetes). Gustav Fischer Verlag, Stuttgart, Germany. 262 pp.
- HIRATSUKA, Y., and J. M. POWELL. 1976. Pine stem rusts of Canada. Can. Dep. Environ., Can. For. Serv., For. Tech. Rep. 4. 103 pp.
- HIRATSUKA, Y., A. TSUNEDA, and L. SIGLER. 1979. Occurrence of Scytalidium uredinicola on Endocronartium harknessii in Alberta, Canada. Plant Dis. Rep. 63:512-513.
- HOCH, H. C., and M. S. FULLER. 1977. Mycoparasitic relationships. I. Mycological features of interactions between Pythium acanthicum and several fungal hosts. Arch. Microbiol. 111:207-224.
- SWART, H. J. 1975. Callosities in fungi. Trans. Br. Mycol. Soc. 64:511-515.
- TRAQUAIR, J. A., and W. E. McKEEN. 1978. Necrotrophic mycoparasitism of Ceratocystis fimbriata by Hirschioporus pargamenus (Polyporaceae). Can. J. Microbiol. 24:869-874.
- TSUNEDA, A., and Y. HIRATSUKA. 1979. Mode of parasitism of a mycoparasite, Cladosporium gallicola, on western gall rust, Endocronartium harknessii. Can. J. Plant Pathol. 1:31-36.
- TSUNEDA, A., Y. HIRATSUKA, and P. J. MARUYAMA. 1980. Hyperparasitism of Scytalidium uredinicola on western gall rust, Endocronartium harknessii. Can. J. Bot. 58:1154-1159.
- TSUNEDA, A., and W. P. SKOROPAD. 1980. Interactions between Nectria inventa, a destructive mycoparasite, and fourteen fungi associated with rapeseed. Trans. Br. Mycol. Soc. 74:501-507.
- TSUNEDA, A., W. P. SKOROPAD, and J. P. TEWARI. 1976. Mode of parasitism of Alternaria brassicae by Nectria inventa. Phytopathology 66:1056-1064.
- ZILLER, W. G. 1974. The tree rusts of Western Canada. Can. Dep. Environ., Can. For. Serv. Publ. 1329. 272 pp.