Genetics

Inheritance of Bacterial Blight Resistance in Rice

J. O. Olufowote, Gurdev S. Khush and H. E. Kauffman

Former Research Scholar, Plant Breeder, and Plant Pathologist, respectively, International Rice Research Institute, Los Baños, The Philippines. Current address of the senior author is: Federal Department of Agricultural Research, Moor Plantation, P.M.B. 5042, Idaban, Nigeria.

Accepted for publication 29 November 1976.

ABSTRACT

OLUFOWOTE, J. O., G. S. KHUSH, and H. E. KAUFFMAN. 1977. Inheritance of bacterial blight resistance in rice. Phytopathology 67: 772-775.

We studied the inheritance and allelic relationships of the genes for resistance to bacterial blight in three rice breeding lines IR1330-3-2, IR944-102, IR1698-241, and in three cultivars, Pelita 1/1, Kele, and Chinsurah Boro II. The study of F_1 , F_2 , F_3 , and reciprocal backcross populations of crosses between resistant parents and the susceptible cultivar Taichung Native 1 (TN-1) revealed that resistance in these rices is under monogenic control. The resistance of IR1330-3-2 and Pelita I/1 is incompletely dominant, whereas that of

Kele and Chinsurah Boro II is recessive. The genes that govern resistance in IR944-102 and IR1698-241 show recessive gene action with the method of inoculation used in this study. Evidence from allele tests with known genes for resistance indicates that Xa4 governs resistance in IR1330-3-2 and Pelita I/1, and xa5 in Kele and Chinsurah Boro II. Rice breeding lines IR944-102 and IR1698-241 have the same gene for resistance which is closely linked to Xa4.

Additional key words: Oryza sativa, Xanthomonas oryzae, allelic relationships.

Bacterial blight of rice [which is caused by *Xanthomonas oryzae* (Uyeda and Ishiyama) Dowson] causes serious damage to that crop throughout Asia (16). The disease has been identified in Japan since the latter 19th century. In tropical Asia, however, it has been noted only during the last 25 yr. Bacterial blight incidence has increased recently owing to the use of intensive agronomic practices that favor the development of the disease, such as high rates of nitrogenous fertilizers, close spacing, and continuous cropping with susceptible cultivars. Chemical control has not been effective (17). The control of the disease through cultivar resistance is now emphasized in most areas (7).

Numerous rice cultivars that are resistant to Japanese isolates of the bacterium have been identified (1, 2, 4, 10, 11, 21). Three loci for resistance have been identified in several of these cultivars (3, 12, 15, 20, 22); Xal and Xa2 loci are closely linked, whereas Xa3, formerly known as Xaw, segregates independently.

Many cultivars that are resistant to Philippine isolates of the bacterium also have been identified (5, 18); these are used in the breeding program at the International Rice Research Institute (9) and elsewhere, and several improved resistant cultivars have been released. Two loci for resistance to Philippine isolates of the bacterium have been identified (6, 13, 14, 19). A dominant gene at the Xa4 locus conveys resistance in Sigadis, TKM 6, IR20, and IR22; a recessive gene at xa5 locus controls resistance in BJ 1 and DZ 192. Genes Xa4 and xa5 segregate independently of each other (19).

This study was undertaken to investigate the mode of inheritance and the allelic relationships of resistance genes in three breeding lines and three cultivars of rice.

MATERIALS AND METHODS

Table 1 lists the rice cultivars and selections used in this study. For inheritance studies, Taichung Native 1 (TN1), a semidwarf cultivar from Taiwan that is highly susceptible to Philippine isolates of the bacterium, was used as a susceptible parent in crosses with resistant cultivars or lines. Rice line IR20, with resistance derived from TKM 6, is homozygous for Xa4 and was used in crosses for allelism tests with Xa4. Line IR1545-339, which was derived from DZ 192, is homozygous for xa5 and has improved plant type. It was used in crosses for allelism tests with xa5. The remaining six cultivars and selections listed in Table 1 were subjected to genetic analysis.

Selection IR1330-3-2 has an improved plant type and is used extensively in IRRI's breeding program. It derives its resistance to bacterial blight from W1263. The tall cultivar from India, W1263, derives its resistance from MTU 15. It is derived from the cross MTU 15 \times Eswarakorra. Pelita I/1 is a popular, high-yielding cultivar from Indonesia that derives its resistance from Syntha. An improved breeding line, IR944-102, derives its resistance from the Philippine cultivar Malagkit Sungsong. Selection IR1698-241 is an improved type that derives its resistance from Zenith. Selections IR944-102 and IR1698-241, like their resistant parents Malagkit Sungsong and Zenith, respectively, show susceptible reactions at seedling and maximum tillering stages, but they are resistant when inoculated at the booting stage or at 70 or more days of age. Kele and Chinsurah Boro II are tall, traditional cultivars from India that are highly resistant to bacterial blight.

The six cultivars and selections were crossed with TN 1; we investigated segregation for resistance in the F_1 , F_2 , and F_3 populations, and reciprocal backcross

Copyright © 1977 The American Phytopathological Society, 3340 Pilot Knob Road, St. Paul, MN 55121. All rights reserved.

populations from most of the crosses. The cultivars that showed dominant gene action for bacterial blight resistance were crossed with IR20, and those that showed recessive gene action were crossed with IR1545-339. We studied F_1 and F_2 populations from these crosses to determine the allelic relationships of the resistance genes.

The hybrid populations were inoculated with the PXo 25 strain of the bacterial blight pathogen from The Philippines, most at maximum tillering stage (45-50 days old). The hybrid populations from the crosses involving IR944-102 and IR 1698-241 however, were inoculated at the booting stage or 70-75 days after seeding. Cultivar TN1 and the resistant parents were grown along with hybrid populations as checks. Plants were inoculated by the clipping technique (8), which consists of clipping off the leaf tips with scissors, or special inoculation clippers, dipped in a suspension of the bacterial blight organism containing 10^9 cells/ml. The bacterial pathogen was cultured on the Wakimoto medium.

Disease scores were taken 2 wk after inoculation following the standard system of Kauffman et al. (8). In the F_1 , F_2 , and backcross populations the disease score was taken on an individual plant basis. Individual plants of the F_3 lines also were scored and each family was classified as homozygous resistant, segregating, or homozygous susceptible.

RESULTS

Inheritance studies.—The F1 hybrids of the cross IR1330-3-2 \times TN1 and of Pelita I/1 \times TN1 were moderately resistant, indicating that resistance in IR1330-3-2 and Pelita I/1 is incompletely dominant. The F1 hybrids of crosses of TN1 with Kele, Chinsurah Boro II, IR944-102, and IR1698-241 were susceptible, showing that these rice lines have recessive resistance. The backcross populations of (IR1330-3-2) × TN1 and (Pelita $I/1 \times TN1 \times TN1$ segregated in a resistant:susceptible ratio of 1:1 (Table 2). Their reciprocal backcross progenies were resistant. These results show that resistance in IR1330-3-2 and Pelita I/1 is controlled by a single dominant gene. The backcross progenies of (Kele \times TN1 × TN1, (IR944-102 × TN1) × TN1, and (IR1698- $241 \times TN1 \times TN1$ were susceptible. But the progenies of their reciprocal backcrosses segregated in a resistant:susceptible ratio of 1:1 (Table 2), thereby showing that resistance is controlled by a single recessive gene in Kele, IR944-102, and IR1698-241.

The reactions of F_2 populations from the crosses of resistant parents with TN1 were determined (Table 3). The F_2 populations of IR1330-3-2×TN1 and Pelita I/1×TN1 segregated in a ratio of 3 resistant to 1 susceptible, thus confirming that resistance in IR1330-3-2 and Pelita

TABLE 1. Rice cultivars and selections used in this study and their reactions to bacterial blight, which is caused by Xanthomonas oryzae

Cultivar/Selection	Origin	Parents	Reaction
Taichung Native 1	Taiwan	$DGWG^a \times Ts$ 'ai-yuan-chung	Susceptible
IR20	IRRI	$(\text{Peta}/3 \times \text{TN } 1^{\text{b}}) \times \text{TKM6}$	Resistant
IR1545-339	IRRI	$\mathbf{R}24 \times \mathbf{D}Z$ 192	Resistant
IR1330-3-2	IRRI	$(IR8 \times Leuang Tawng)$	
		× W1263	Resistant
Pelita I/1	Indonesia	$IR5 \times Syntha$	Resistant
IR944-102	IRRI	$(TN1 \times Malagkit Sungsong)$	
		× IR8	Resistant ^c
IR 1698-241	IRRI	$IR8/4 \times Zenith$	Resistant ^c
Kele	India		Resistant
Chinsurah Boro II	India		Resistant

^aCoding DGWG stands for Dee-Geo-Woo-Gen.

^bCoding TN 1 stands for susceptible cultivar Taichung Native 1.

Susceptible at seedling stage but resistant at booting and postflowering stages.

TABLE 2. Classification of rice plants from reciprocal backcrosses for resistance to bacterial blight, which is caused by Xanthomonas oryzae

		Plants (no.)		γ^2	P-value
Cross	Resistant	Susceptible	Total	(1:1)	(1:1)
$(IR1330-3-2 \times TN 1) \times TN 1^{a}$	44	48	92	0.17	0.50-0.75
(Pelita $I/1 \times TN 1$) × TN 1	39	42	81	0.11	0.50-0.75
$(Kele \times TN1) \times TN-1$		84	84		
$(1R944-102 \times TN 1) \times TN 1$		79	79		
$(1R1698-241 \times TN1) \times TN1$		56	56		
$(IR1330-3-2 \times TN-1) \times IR1330-3-2$	70		70		
(Pelita I/1 \times TN 1) \times Pelita I/1	68		68		
(Kele \times TN 1) X \times Kele	46	47	93	0.01	0.90-0.95
$(IR944-102 \times TN 1) \times IR944-102$	41	36	77	0.32	0.50-0.75
$(IR1698-241 \times TN 1) \times IR1698-241$	45	42	87	0.10	0.50-0.75

^aCoding TN 1 stands for susceptible cultivar Taichung Native 1.

PHYTOPATHOLOGY

I/1 is controlled by a single dominant gene. On the other hand, the F₂ populations of Kele \times TN1, IR944-102 \times TN1, and IR1698-241 \times TN1 crosses segregated in a ratio of 1 resistant to 3 susceptible, thus confirming that single recessive genes govern the resistance in Kele, IR944-102, and IR1698-241. Analysis of the F₃ confirmed the hypothesis regarding monogenic control of resistance in IR1330-3-2, Pelita I/1, IR944-102, and IR1698-241. The proportion of resistant, segregating, and susceptible F₃ families in each cross closely approximates the 1:2:1 ratio expected for monogenic control of resistance (Table 4).

Allele tests.—The F_1 progenies of the crosses IR1330-3-2 × IR20 and Pelita I/1 X IR20 were resistant. All the plants of the F_2 populations of these two crosses also were resistant (Table 5) which showed that IR1330-3-2 and Pelita I/1 possess Xa4 for resistance. The F_1 hybrids of Kele × IR1545-339 and Kele × Chinsurah Boro II were also resistant, and the F_2 populations did not segregate for susceptibility (Table 5). Therefore, it appears that Kele and Chinsurah Boro II possess the same recessive gene, xa5, for resistance. The F_1 progenies of IR1698-241 × IR1545-339 were susceptible, indicating that these two selections have non-allelic genes. The F_1 progenies of IR1698-241 × IR20 were resistant and 1.3 percent of the F_2 plants of this cross were susceptible (Table 5), which showed that the recessive gene for resistance in IR1698-241 is closely linked with Xa4. The F_1 progenies of IR1698-241 × IR944-102 were resistant and all the F_2 plants of this cross also were resistant (Table 5), thereby showing that these two selections possess the same recessive gene for resistance.

In the F_2 population of Kele × IR22, 19.8% susceptible plants were observed (Table 5). This proportion agrees well with the 18.7 percent expected for two independently segregating genes. These results confirm the conclusions of Petpisit et al. (19) regarding independence of Xa4 and xa5

DISCUSSION

The results of this study suggest that IR1330-3-2 and Pelita I/1, which derive their resistance from MTU 15 and Syntha, respectively, have gene Xa4 for resistance. The same gene confers resistance in TKM6, Sigadis, and IR22 (19). The resistance in Kele and Chinsurah Boro II is governed by the recessive gene xa5, which also confers resistance in DZ192 and BJ1 (19). The four cultivars with

TABLE 3. Classification of F_2 plants for bacterial blight resistance from crosses of resistant parents with the susceptible cultivar Taichung Native 1^a

Cross		Plants (no.)			P-value	
	Resistant	Susceptible	Total	(3:1/1:3)	(3:1/1:3)	
IR1330-3-2 × TN 1	911	310	1,221	0.04	0.75-0.90	
Pelita $I/1 \times TN 1$	589	223	812	2.62	0.10-0.25	
$IR944-102 \times TN 1$	344	914	1,258	3.68	0.05-0.10	
IR1698-241 \times TN 1	202	558	760	1.01	0.25-0.50	
Kele \times TN 1	553	1,772	2,325	1.83	0.10-0.25	

^aDesignated TN 1 in the indicated crosses.

TABLE 4. Classification of F_3 families for bacterial blight resistance from crosses of resistant parents with the susceptible cultivar Taichung Native 1^a

		Families (no.)			
Cross	Homozygous- resistant	Segregat- ing	Homozygous- susceptible	χ^2 (1:2:1)	<i>P</i> -value (1:2:1)
IR1330-3-2 × TN 1	31	67	28	0.65	0.50-0.75
Pelita I/1 \times TN 1	35	63	29	0.57	0.50-0.75
$IR944-102 \times TN 1$	29	58	31	0.10	0.95-0.97
$IR1698-241 \times TN 1$	28	66	29	0.67	0.50-0.75

^aDesignated TN 1 in the indicated crosses.

TABLE 5. Classification of F2 rice plants for bacterial blight resistance from crosses between resistant parents

Cross	a			
	Resistant	Susceptible	Total	- Susceptibl (%)
IR1330-3-2 × IR20	1,431	0	1,431	
Pelita I/1 \times IR20	1,475	0	1,475	
Kele \times IR1545-339	750	0	750	
Kele \times Chinsurah Boro II	1,492	0	1,492	
IR1698-241 × IR20	735	10	745	1.3
IR1698-241 × IR944-102	500	0	500	
Kele \times IR22	897	222	1,119	19.84

recessive genes for resistance are from Bangladesh and the adjoining Indian state of West Bengal.

Selections IR944-102 and IR1698-241 probably have the same gene for resistance, which is non-allelic to Xa4 and xa5. However, it appears closely linked to Xa4. Similar observations were made by Murty et al. (14) who found only a few susceptible plants in the F₂ population of Zenith \times Sigadis. Because IR1698-241 derives its resistance from Zenith, and because Sigadis was shown to possess Xa4 for resistance (19), the third locus for resistance to the Philippine isolates of the bacterium is firmly established. This source of resistance is being used in our breeding program.

The gene for resistance in IR944-102 and IR1698-241 appears to be recessive when the segregating populations are inoculated at the booting stage or at about 70-75 days after sowing, but there is some evidence that the pattern of segregation is monogenic dominant when plants are inoculated after flowering or at about 90-95 days after sowing. Plants that are heterozygous for the resistance gene seem to show a susceptible reaction at the booting stage, but they have a resistant reaction at post-flowering stages. Thus, an F_2 population that is inoculated at booting stage would segregate 3 susceptible: I resistant, as in this study, but the same F_2 population would segregate I susceptible: 3 resistant if inoculated after flowering. The gene symbol to this locus will be assigned after this problem is thoroughly investigated.

LITERATURE CITED

- 1. EZUKA, A., and O. HORINO. 1974. Classification of rice varieties and Xanthomonas oryzae strains on the basis of their differential interactions. Bull. Tokai-Kinki Nat. Agric. Exp. Stn. 27:1-19.
- EZUKA, A., O. HORINO, K. TORIYAMA, H. SHINODA, and T. MORINAKA. 1975. Inheritance of resistance of rice variety Wase Aikoku 3 to Xanthomonas oryzae. Bull. Tokai-Kinki Nat. Agric. Exp. Stn. 28:124-130.
- EZUKA, A., Y. WATANABE, and O. HORINO. 1970. Differences in resistance expression to Xanthomonas oryzae between seedlings and adults of Wase Aikoku group rice varieties. Bull. Tokai-Kinki Nat. Agric. Exp. Stn. 27:20-25.
- 4. FUJII, K., and M. OKADA. 1967. Progress in breeding of rice varieties for resistance to bacterial leaf blight in Japan. Pages 51-61 in Proc. Symp. on Rice Diseases and Their Control by Growing Resistant Varieties and Other Measures. Agric. For. Fish., Res. Council, Min. Agric. For., Tokyo, Japan. 250 p.
- 5. GOTO, M. 1965. Resistance of rice varieties and species of wild rice to bacterial leaf blight and bacterial leaf streak disease. Philipp. Agric. 48:329-338.
- 6. HEU, M., T. T. CHANG, and H. M. BEACHELL. 1968. The inheritance of culm length, panicle length, duration of heading and bacterial leaf blight resistance in a rice

cross Sigadis \times Taichung (Native) 1. Jpn. J. Breed. 18:7-12.

- JAYARAJ, D., D. V. SESHU, and S. V. S. SHASTRY. 1972. Genetics of resistance to bacterial leaf blight in rice. Indian J. Genet. Plant Breed. 32:77-89
- KAUFFMAN, H. E., A. P. K. REDDY, S. P. Y. HSIEH, and S. D. MERCA. 1973. An improved technique for evaluating resistance of rice varieties to Xanthomonas oryzae. Plant Dis. Rep. 56:537-541.
- KHUSH, G. S., and H. M. BEACHELL. 1972. Breeding for disease and insect resistance at IRRI. Pages 309-322 in Rice breeding. International Rice Research Institute, Los Baños, The Philippines. 738 p.
- KONDO, G., and M. FUKUNAGA. 1953. On the varietal difference to the resistance of paddy rice to bacterial leaf blight. Jpn. J. Breed. 3(2):29-32.
- 11. MIZUKAMI, T. 1966. Resistance of rice plant to bacterial leaf blight and strains of causal bacterium. Jpn. Agric. Res. Quart. 1:6-11.
- 12. MURATA, N. 1967. Genetic aspects on resistance to bacterial leaf blight in rice and variation of its causal bacterium. Pages 39-49 *in* Proc. Symp. on Rice Diseases and Their Control by Growing Resistant Varieties and Other Measures. Agric. For. Fish. Res. Council, Min. Agric. For., Tokyo, Japan. 250 p.
- MURTY, V. V. S., and G. S. KHUSH. 1972. Studies on the inheritance of resistance to bacterial leaf blight in rice varieties. Pages 301-305 in Rice breeding. International Rice Research Institute, Los Baños, Philippines. 738 p.
- MURTY, V. V. S., G. S. KHUSH, and N. F. JENSEN. 1973. Inheritance of resistance to bacterial leaf blight, Xanthomonas oryzae (Uyeda and Ishiyama) Dowson in rice. I. Allelic relationships of resistance genes in donor varieties. Jpn. J. Breed. 23:325-328.
- NISHIMURA, Y., and S. SAKAGUCHI. 1959. Inheritance of resistance in rice to bacterial leaf blight (Bacterium oryzae Uyeda and Ishiyama) Nakata. Jpn. J. Breed. 9:58.
- OU, S. H. 1972. Rice diseases. Commonwealth Mycological Institute, Kew, Surrey, England. 368 p.
- OU, S. H. 1973. A handbook of rice diseases in the tropics. The International Rice Research Institute, Los Baños, Philippines. 58 p.
- OU, S. H., F. L. NUQUE, and J. P. SILVA. 1971. Varietal resistance to bacterial blight of rice. Plant Dis. Rep. 55:17-21.
- PETPISIT, V., G. S. KHUSH, and H. E. KAUFFMAN. 1976. Inheritance of resistance to bacterial blight in rice. Crop Sci. 17:(In press).
- SAKAGUCHI, S. 1967. Linkage studies on the resistance to bacterial leaf blight, Xanthomonas oryzae (Uyeda and Ishiyama) Dowson, in rice. Bull. Nat. Inst. Agric. Sci. Japn (Ser. D)16:1-18.
- SAKAGUCHI, S., T. SUWA, and N. MURATA. 1967. Studies on the resistance to bacterial leaf blight, Xanthomonas oryzae (Uyeda and Ishiyama) Dowson in rice. Bull. Nat. Inst. Agric. Sci. Japn (Ser. D)18:1-29.
- 22. TORIYAMA, K. 1972. Breeding for resistance to major rice diseases in Japan. Pages 253-281 in Rice breeding. The International Rice Research Institute, Los Baños, The Philippines. 738 p.