Host Range of the Columbia Root-Knot Nematode

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ABSTRACT

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The Columbia root-knot nematode, Meloidogyne chitwoodi, a severe pest of potato in the Pacific Northwest, reproduced on 53 of 68 plant species tested under greenhouse conditions. Both monocotyledonous and dicotyledonous plant species were good hosts, indicating that M. chitwoodi has a wide host range. Principal crops used in rotation with potato in the Pacific Northwest include a poor host (alfalfa), but cereals such as barley, corn, and wheat were good hosts for this nematode.

An undescribed root-knot nematode was found attacking potato (Solanum tuberosum L.) in the Pacific Northwest in 1977-1978 (9). The new species was described as Meloidogyne chitwoodi Golden, O'Bannon, Santo, and Finley (4) and given the common name "Columbia root-knot nematode." It is presently known in Idaho, Oregon, Washington, northern California, and Nevada (6). Economically, potato is the major crop most severely damaged by M. chitwoodi, but it can reproduce and is pathogenic on oats (Avena sativa L.), barley (Hordeum vulgare L.), corn (Zea mays L.), and wheat (Triticum aestivum L.), crops that are used in rotation with potato (8,9).

The northern root-knot nematode, M. hapla, which also attacks potatoes, is limited to broad-leaf plants (2). Because of the need for effective rotation and weed control, a study was made to determine crop and weed hosts of M. chitwoodi.

MATERIALS AND METHODS

Several host range tests were conducted in a greenhouse for 2 yr. Four or five

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seeds, later thinned to one or two seedlings or a single seed piece, were either pregerminated or planted directly into 10-cm plastic pots containing loamy sand (82.3% sand, 14.8% silt, 2.9% clay) fumigated with methyl bromide. Pots were randomized in five replicates on greenhouse benches, and plants were inoculated after 2- or 3-wk growth with 3,000-4,000 M. chitwoodi eggs in a 25-ml water suspension added to the roots of plants in each pot. Inoculum came from infected tomatoes (Lycopersicon esculentum Mill. 'Rutgers') maintained in a greenhouse. Eggs were extracted from roots by the sodium hypochlorite method

Greenhouse temperatures were maintained at 20-25 C, which is favorable for nematode infection and reproduction (7). Plants were watered daily and fertilized with Hoagland's nutrient solution bimonthly. Tests were terminated after 50 days, and the roots were carefully removed and washed free of soil. To measure host susceptibility and nematode reproduction, roots were soaked (15 min) in a phloxine B solution (1) (150 mg/L) and examined with a dissecting microscope. After examination, nematode eggs were extracted (5) and brought to volume (100 ml), and a 1-ml aliquot sample was counted. Roots were ovendried and weighed, and the number of eggs per gram of dry root was calculated. Plants were rated as to relative degree of nematode reproduction based on egg masses and eggs per gram of oven-dry root. Ratings were 0 = no reproduction, nonhost; 0.1-1 = trace reproduction,

<100 eggs per gram of root, very poor host; 1.1-2 = light reproduction, 100-1,000 eggs, poor host; 2.1-3 =moderate reproduction, >1,000-10,000 eggs, moderate host; 3.1-4 = high reproduction, >10,000-100,000 eggs, good host; 4 + = very high reproduction, >100,000 eggs, very good host.

Because several plants observed in our studies are grown in a rotation program, we tested several varieties within a species to ascertain the possible existence of resistant germ plasm or nematodetolerant cultivars.

RESULTS AND DISCUSSION

M. chitwoodi was found to infect and reproduce on 53 of 68 plant species tested, indicating a wide host range (Table 1). Unlike M. hapla (2), many of the Gramineae as well as many dicotyledonous species were good host of M. chitwoodi. Because alfalfa is often used in the rotation, several Medicago sativa cultivars were tested and found to be either nonhosts or poor hosts. Most alfalfa varieties are susceptible to M. hapla, and several cereals susceptible to M. chitwoodi were also used in the rotation. Therefore, this information is of value to growers when practicing rotation with potatoes and alerts field and extension personnel to the necessity of species identification between M. chitwoodi and M. hapla.

In the field, galls and egg masses produced by M. chitwoodi are often difficult to see on roots of most host plants, even with the aid of a hand lens. To see egg masses better, roots should be stained with phloxine B, which stains the masses red for rapid identification (1). This can be done in the field and also serves as an excellent teaching example for the grower.

To aid interested persons in developing diagnostic evaluation for species separation of suspicious sites, specific hosts for M. chitwoodi or M. hapla should be used. Wheat cv. Nugaines, a host of M. chitwoodi, and pepper cv. California Wonder, a host of M. hapla,

Table 1. Response of several plant species to Meloidogyne chitwoodi

cientific name	Common name	Horticultural variety	Rating as host ^a
MARANTHACEAE	Total		
Amaranthus graecizons A. retroflexus	Tumbleweed Pigweed		0.0
SCLEPIADACEAE	. Ignocu		0.0
Asclepias syriaca	Common milkweed		0.1
ANNABACEAE	www.		1979
Humulus lupulus HENOPODIACEAE	Нор		0.0
Beta vulgaris	Fodder beet		1.6
	Sugarbeet	U&I Hybrid No. 9	2.1
Chenopodium album	Lambsquarter		0.1
Salsola kali OMPOSITAE	Russian thistle		1.1
Cirsium arvense	Canada thistle		1.0
C. vulgare	Bull thistle		3.5
Sonchus asper	Spiny sowthistle		3.0
Taraxacum officinale	Dandelion		1.1
RUCIFERAE Brassica hirta	Oil mustard	Kirby	2.3
B. juncea	Mustard weed		1.1
B. napus	Rape	Baraska	2.1
) Y	W. 1912 2 W	Bar-Bn IVa 814-5	1.4
B. rapa	Stubble turnip	Alander Endiviana	2.4 2.3
		Gelria	2.4
		Ponda	2.3
		Samson	2.4
		Vollenda Weseler	2.1 2.2
	Turnip	Barive	2.2
		Purpletop	2.5
B. rapa var. rapa	Stubble turnip × Chinese cabbage	Appin	1.6
B. rapa var. silvestris	Stubble turnip × Chinese cabbage	Daisy	2.2
Capsella bursa-pastoris Raphanus sativus	Shepherd purse Fodder radish	Nerus	1.3 0.0
CUCURBITACEAE	Foddel ladisii	Nerus	0.0
Citrullus vulgaris	Watermelon	Charleston Grey	0.1
RAMINEAE	\$5400 ASSASSASSASSASSASSASSASSASSASSASSASSASS		
Aegilops cylindrica	Jointed goatgrass		0.7
Agropyron repens Alopecurus pratensis	Quackgrass Meadow foxtail		1.0 0.0
Arrhenatherum elatius	Tall oatgrass		0.0
Avena sativa	Oats	Appaloosa	4+
		Cayuse	4 +
		Corbit -	4 + 4 +
		Markton Menominee	4+
		Otana	4+
		Park	3.3
		Random	4+
Bromus tectorum	Downy brome	Terra	4 + 1.1
Cenchrus incertus	Field sandburr		0.0
Cynodon dactylon	Bermuda grass		0.5
Dactylis glomerata	Orchardgrass		2.1
Echinochloa crus-galli Eragrostis orcuttiana	Barnyardgrass Lovegrass		1.2 1.8
Festuca arundinacea	Meadow fescue		2.1
	Tall fescue		0.1
(867 Pa 951)	Red fescue		1.8
Hordeum vulgare	Barley	Advance	3.9
	*	Boyer Hesk	3.2 3.1
		Kamiak	4+
Hordeum vulgare	The state of the s	Klages	3.3
Hordeum vulgare	Barley		
Hordeum vulgare	Barley	Mal	4+
Hordeum vulgare	Barley	Morex	4+
Hordeum vulgare	Barley	Morex Piroline	4 + 4 +
Hordeum vulgare	Barley	Morex	4+
Panicum capillare	Witchgrass	Morex Piroline Steptoe	4 + 4 + 3.4 4 + 2.1
Panicum capillare Poa annua	Witchgrass Annual bluegrass	Morex Piroline Steptoe	4 + 4 + 3.4 4 + 2.1 1.6
Panicum capillare Poa annua P. pratensis	Witchgrass Annual bluegrass Kentucky bluegrass	Morex Piroline Steptoe	4 + 4 + 3.4 4 + 2.1 1.6 0.5
Panicum capillare Poa annua	Witchgrass Annual bluegrass	Morex Piroline Steptoe	4 + 4 + 3.4 4 + 2.1 1.6
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard	4 + 4 + 3.4 4 + 2.1 1.6 0.5 1.2 2.1 0.1
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail	Morex Piroline Steptoe Vanguard	4 + 4 + 3.4 4 + 2.1 1.6 0.5 1.2 2.1 0.1 4 +
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin	4+ 4+ 3.4 4+ 2.1 1.6 0.5 1.2 2.1 0.1 4+ 4+
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin Fielder	4 + 4 + 3.4 4 + 2.1 1.6 0.5 1.2 2.1 0.1 4 + 4 + 4 +
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin	4+ 4+ 3.4 4+ 2.1 1.6 0.5 1.2 2.1 0.1 4+ 4+
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin Fielder Fieldwin Owens Sawtell	4 + 4 + 3.4 4 + 2.1 1.6 0.5 1.2 2.1 0.1 4 + 4 + 4 + 4 + 4 +
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin Fielder Fieldwin Owens Sawtell Sterling	4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 +
Panicum capillare Poa annua P. pratensis Setaria lutescens S. viridis Sorghum vulgare	Witchgrass Annual bluegrass Kentucky bluegrass Yellow foxtail Green foxtail Sudangrass	Morex Piroline Steptoe Vanguard Borah Dirkwin Fielder Fieldwin Owens Sawtell	4 + 4 + 3.4 4 + 2.1 1.6 0.5 1.2 2.1 0.1 4 + 4 + 4 + 4 + 4 +

Scientific name	Common name	Horticultural variety	Rating as host ^a
		Hyslop	3.3
		Luke	3.3
		Moro	3.1
		Neeley	3.3
Total	W0	Nugaines	4+
Triticum aestivum	Wheat (winter)	Palouse	3.3
		Prodax	4 +
	323	Stephens	4+
		Tyee	3.4
-		Wanser	3.5
Zea mays	Corn	Cenex 2119	2.4
		Funks G-4444 MF	3.2
		Funks G4574	2.1
		Idahybrid 216	3.1
		Idahybrid 303	0.9
		JX 122A	2.2
		JX 162	1.5
		JX 1153	2.3
		Northrup King 497	0.6
		Northrup King 589	3.2
	2	Northrup King PX 46	2.3
. E. C.	Sweet corn	Jubilee	2.0
ABIATAE			
Mentha cardiaca	Spearmint	Scotch	0.4
M. piperita	Peppermint	Todd	0.3
Photograph, these visits		Black	0.2
M. spicata	Spearmint	Native	0.6
EGUMINOSAE			
Arachis hypogaea	Peanut	Florrunner	0.0
Medicago sativa	Alfalfa	African	0.1
		DuPuit	0.0
		Lahanton	0.1
		Mn PL 9HF (Syn 2)	1.3
		Moapa	0.0
		Nevada Syn XX	0.4
		Nevada Syn VV	0.1
		123	0.1
		Ranger	0.1
		Rhizoma	0.0
		Saranac	0.2
		Thor	0.1
		Vernal	0.1
		Vertus	0.1
		W9S ₂ R ₂ W ₁	0.1
		W12S ₂ R ₂ W ₁ Fu ₁	0.0
		Washoe	0.2
Phaseolus limensis	Lima bean	Henderson Baby Bush	
P. vulgaris	Snap bean	Apollo	0.0 2.5
Pisum sativum	Edible podded pea	ARS 244219-B	
1 isam samam	Garden pea	Alaska	3.1
	Garden pea	Dark Skin Perfection	2.1
		PH 14-119	2.1
		PI-189171	1.7
			2.4
Trifolium pratense	Red clover	PI-257593	2.4
T. repens			2.8
	White clover	Caller to Division at the	2.8
Vigna sinensis	Cowpea	California Black Eye No. 5	0.2
LIACEAE	0-:	D. J.	
Allium cepa	Onion	Rocket	0.3
Annuary - 10-1 11	W & County County Comment	Walla Walla Sweet	1.1
Asparagus officinalis	Asparagus	Mary Washington	0.0
ALVACEAE	6	1	5250,975
Gossypium hirsutum	Cotton	Delta Pine 16	0.0
Hibiscus esculentus	Okra	Dwarf Green	1.1
OSACEAE	Table 1 (1) (1) (1) (1) (1) (1)		AC 94
Fragaria chiloensis	Strawberry		0.1
Malus sylvestris	Apple	Golden Delicious	0.0
DLANACEAE		943 844 Carlo - Data Carlo Car	
Capsicum annuum	Pepper	California Wonder	0.0
Lycopersicon esculentum	Tomato	Ace	3.5
		Beefsteak	2.7
		Big Boy	2.7
		Bonny Best	2.5
		Columbian	3.9
		Patriot	2.9
		Roza	4+
		Rutgers	3.7
		Saladmaster	3.4
		Sunray	3.1
		Tiny Tim	2.9
		Yellow Pear	3.2
	Wild tomato	renow real	
neruvianum	wild tolliato	PI 270435	2.3 1.1
L. peruvianum			1.1
	Tahaaas	11270433	
Nicotiana tabacum	Tobacco Nightshada	11270433	0.0
L. peruvianum Nicotiana tabacum Solanum nigrum S. melongena	Tobacco Nightshade Eggplant	11270433	

Scientific name	Common name	Horticultural variety	Rating as host
S. tuberosum	Potato	Bel-rus	2.5
5. Inverosum		Lemhi	3.1
		Russet Burbank	3.5
		White Rose	2.5
UMBELLIFERAE		Gold Pak	2.2
Daucus carota	Carrot	Half Long	1.1
		Imperator	0.1
VITACEAE		2000 C 20	0.0
Vitis labrusca	Grape	Concord	0.2
V. vinifera	Wine grape	Semillon	0.4

^{*0 =} no reproduction, nonhost; 0.1-1 = trace reproduction, <100 eggs per gram of root (oven-dry), very poor host; 1.1-2 = light reproduction, 100-1,000 eggs, poor host; 2.1-3 = moderate reproduction, >1,000-10,000 eggs, moderate host; 3.1-4 = high reproduction, >10,000-100,000 eggs, good host; 4+= very high reproduction, >100,000 eggs, very good host.

are good indicator hosts for the respective species. If soil, root, or tuber samples infested with root-knot nematodes are available, L₂ juveniles and eggs can be extracted and inoculated onto individual seedlings. After a given period of growth (55 days), plants can be harvested and the roots immersed in phloxine B. In conjunction with the indicator host test, female perineal patterns and tail shape of L₂ juveniles are of greater value to technical laboratories.

Even though certain plant species tested were poor hosts when used in rotation with potato, low populations of *M. chitwoodi* remaining from the previous crop can increase rapidly on potato and diminish the tuber quality, which is the major concern with potato

(3,8). This problem is also evident with several weed species examined that act as infection reservoirs for *M. chitwoodi*, thus necessitating a good weed control program.

LITERATURE CITED

- Dickson, D. W., and Struble, F. B. 1965. A sieving-staining technique for extraction of egg masses of *Meloidogyne incognita* from soil. (Abstr.) Phytopathology 55:497.
- Faulkner, L. R., and McElroy, F. D. 1964. Host range of northern root-knot nematode on irrigated crop plants and weeds in Washington. Plant Dis. Rep. 48:190-193.
- Finley, A. M. 1981. Histopathology of Meloidogyne chitwoodi (Golden et al) on Russet Burbank potato. J. Nematol. 13:486-491.
- Golden, A. M., O'Bannon, J. H., Santo, G. S., and Finley, A. M. 1980. Description and SEM observations of *Meloidogyne chitwoodi* n. sp. (Meloidogynidae), a root-knot nematode of

- potato in the Pacific Northwest, J. Nematol. 12:319-327.
- Hussey, R. S., and Barker, K. R. 1973. A comparison of methods of collecting inocula of Meloidogyne spp. including a new technique. Plant Dis. Rep. 57:1025-1028.
 Nyczepir, A. P., O'Bannon, J. H., and Santo, G. S.
- Nyczepir, A. P., O'Bannon, J. H., and Santo, G. S. 1981. Distribution of *Meloidogyne chitwoodi* on potato in the Pacific Northwest. (Abstr.) J. Nematol. 13:453.
- Santo, G. S., and O'Bannon, J. H. 1981. Effect of soil temperature on the pathogenicity and reproduction of *Meloidogyne chitwoodi* and *M. hapla* on Russet Burbank potato. J. Nematol. 13:483-486.
- Santo, G. S., and O'Bannon, J. H. 1981. Pathogenicity of the Columbia root-knot nematode (Meloidogyne chitwoodi) on wheat, corn, oat, and barley. J. Nematol. 13:548-550.
- Santo, G. S., O'Bannon, J. H., Finley, A. M., and Golden, A. M. 1980. Occurrence and host range of a new root-knot nematode (*Meloidogyne* chitwoodi) in the Pacific Northwest. Plant Dis. 64:951-952.