

Disease Notes

Plant Disease "Disease Notes" are originally published on-line and are available several weeks before being reprinted here. A note's official date of publication is the date it was placed on-line. Information concerning on-line access appears at the front of this journal under *Editorial Policies*.

First Report of Powdery Mildew of Greenhouse Pepper Caused by *Leveillula taurica* in Canada. R. F. Cerkauskas and J. Brown, Agriculture & Agri-Food Canada; and G. Ferguson and S. Khosla, Ontario Ministry of Agriculture, Food & Rural Affairs, Greenhouse & Processing Crops Research Centre, Harrow, Ontario, Canada N0R 1G0. *Plant Dis.* 83:781, 1999; published on-line as D-1999-0524-01N, 1999. Accepted for publication 19 May 1999.

In 1999, powdery mildew on cvs. Oberon and Triple 4 of greenhouse pepper (*Capsicum annum* L.) and cv. FireFlame of chili pepper was reported simultaneously in two commercial greenhouses at two separate locations, Leamington and Vineland, geographically separated by 290 km, in southern Ontario. Losses of 10 to 15% each in the 2 and 3.1 ha greenhouse pepper operations were noted. The greenhouse pepper industry in Canada consists of 89.4 ha with sales of \$43.6 million (Canadian). Lesions appeared as a white, powdery coating on the abaxial leaf surface only, generally on the lower foliage of pepper plants, while diffuse chlorotic spots were present at corresponding locations on the adaxial surface. In chili pepper, this chlorosis was restricted to interveinal tissue, causing the leaves to have a somewhat netted appearance. In some cases, pale yellow spots appeared on the adaxial portion of the affected foliage during later stages of disease development. No cleistothecia were observed. Pyriform and cylindrical, hyaline, single-celled conidia were present in equal numbers. Both conidia had a network of crests and granules containing tiny, thornlike projections on the surface. These projections were also evident on the lower half of conidiophores. Pyriform conidia ranged in length from 61.6 to 84.0 μm (mean = 70.8, SE = 0.7) and in width from 14.0 to 25.8 μm (mean = 21.1, SE = 0.4, $n = 50$), with a mean length to width ratio of 3.4. Cylindrical conidia ranged in length from 54.9 to 80.1 μm (mean = 66.3, SE = 0.9) and in width from 15.7 to 24.1 μm (mean = 18.6, SE = 0.3, $n = 50$), with a mean length to width ratio of 3.1. Short conidial chains borne on conidiophores consisted of a pyriform conidium first followed by cylindrical conidium. Stomatal penetration and extensive endophytic mycelial growth in the mesophyll layer were evident in foliar tissue cleared with glacial acetic acid:EtOH (1:2) and stained with lactophenol cotton blue. To confirm pathogenicity, conidia from infected pepper leaves were dusted onto the water-misted abaxial leaf surface of 41-day-old greenhouse pepper cvs. Cubico and Edison, and onto the similarly treated adaxial leaf surface of 58-day-old field pepper cv. Renegade. Plants were enclosed in plastic bags for 24 h on a bench. Control plants were treated identically but not inoculated. Inoculated plants developed foliar powdery mildew symptoms, including sporulation similar to that of naturally infected plants. Occurrence of the fungus on the abaxial surface of pepper foliage, presence of endophytic mycelium, and the morphological characteristics of the imperfect state confirm the identity of the fungus. This is the first report of *Leveillula taurica* (Lév.) G. Arnaud on pepper in Canada.

Reference: J. Palti. *Bot. Rev.* 54:423, 1988.

Occurrence of a Subpopulation of *Xanthomonas oryzae* pv. *oryzae* with Virulence to Rice Cv. IRBB21 (Xa21) in Southern India. V. Brindha Priyadarisini and S. S. Gnanamanickam, Center for Advanced Studies in Botany, University of Madras, Chennai, India 600 025. E-mail: mmanick@hotmail.com. *Plant Dis.* 83:781, 1999; published on-line as D-1999-0524-02N, 1999. Accepted for publication 20 May 1999.

About 200 isolates of the bacterial blight pathogen *Xanthomonas oryzae* pv. *oryzae* were obtained from infected rice (*Oryza sativa* L.) leaf samples from the Palghat District of Kerala State in southern India during January and February, 1998, when there was a severe epidemic of bacterial blight in the farmers' fields. One hundred and forty of these isolates were assayed for pathogenicity on rice cv. IR24 (no *R* gene), IRBB21 (Xa21), and NH56 (Xa4+xa5+xa13+Xa21) at their maximum tillering stage by the standard clip-inoculation technique (1) at the Regional Agricultural Research Station (Kerala Agricultural University), Pattambi. In two separate experiments, all the isolates were found to be pathogenic to IR24, producing lesions greater than 10 cm in length. None of the isolates induced susceptible symptoms on cv. NH56 and the average lesion length was less than 2 cm. Twenty of the 140 isolates, however, showed

virulence to IRBB21 (seeds supplied by S. R. McCouch, Cornell University, and by P. Ronald, University of California-Davis) and induced lesions of 10.0 to 18.0 cm. We report here the presence of Indian races of *X. oryzae* pv. *oryzae* that are virulent to IRBB21 (Xa21), contradicting an earlier report of avirulence (2). These results suggest that the deployment of Xa21 alone will not be a sound strategy for the management of bacterial blight in southern India.

References: (1) S. S. Gnanamanickam et al. *Plant Dis.* 78:173, 1994. (2) G.-L. Wang et al. *Mol. Plant-Microbe Interact.* 9:850, 1996.

Symptomatology of Barley Yellow Dwarf Virus-RMV Infection in Sweet Corn. R. L. C. Itnyre, Cleora J. D'Arcy, J. K. Pataky, and W. L. Pedersen. Department of Crop Sciences, University of Illinois, Urbana 61801. *Plant Dis.* 83:781, 1999; published on-line as D-1999-0525-01N, 1999. Accepted for publication 21 May 1999.

Infection of sweet corn (*Zea mays* L.) by barley yellow dwarf viruses (BYDVs) caused different symptoms on hybrids with *shrunken-2* (*sh-2*) when compared with hybrids with *sugary-1* (*su-1*) endosperm mutations. Sweet corn hybrids inoculated with BYDV-RMV in Urbana, IL, developed either yellow or red-purple leaf symptoms similar to those caused by phosphorus and/or potash deficiency (1). In a field trial near Rochelle, IL, 329 of 339 sweet corn hybrids naturally infected by BYDV exhibited one or the other of these symptoms. Eighteen of the hybrids with severe symptoms were tested for two strains of BYDVs in enzyme-linked immunosorbent assays (1). All of the hybrids tested were positive for BYDV-RMV; none were positive for BYDV-PAV. Symptoms on *su-1* hybrids were typically characterized by dark red to purple coloration of leaf margins from the tip down to the mid-leaf. Leaves of *sh-2* hybrids exhibited chlorosis in a similar pattern. A single *sh-2* hybrid exhibited red-purple leaf symptoms, probably as a result of a crossover between the anthocyanin production (*a1*) and *sh-2* genes, which are linked about 0.25 map units apart on chromosome 3 (2). Infection of *sh-2* sweet corn hybrids by BYDV-RMV is most likely to induce chlorosis, whereas infection of *su-1* sweet corn hybrids can result in either chlorosis or red-purple discoloration of leaves. Sweet corn hybrids with these symptoms should be tested for BYDVs, particularly the RMV strain.

References: (1) R. L. C. Itnyre et al. *Plant Dis.* 83:566, 1999. (2) E. B. Mains. *J. Hered.* 40:21, 1948.

Fairy Rings in Turf Associated with the Bird's Nest Fungus *Cyathus stercoreus*. J. Mercier, Department of Plant Pathology, 495 Borlaug Hall, University of Minnesota, St. Paul 55108; and T. D. Carson and D. B. White, Department of Horticulture, Alderman Hall, University of Minnesota, St. Paul 55108. *Plant Dis.* 83:781, 1999; published on-line as D-1999-0526-01N, 1999. Accepted for publication 25 May 1999.

During the summer of 1998, several fairy rings with the fruiting bodies of a bird's nest fungus were observed on two greens at the University of Minnesota turf plots in St. Paul. Both greens, a 3-year-old annual bluegrass green and a 2-year-old creeping bentgrass green, were constructed of a 90% sand: 10% reed sedge peat soil mix. The fungus was identified as *Cyathus stercoreus* (Schwein.) de Toni (Nidulariaceae) (1). The fruiting bodies were approximately 15 mm tall and 8 mm wide. The fairy rings were irregular in shape, with a diameter of up to 50 cm, and contained up to 120 fruiting bodies per ring. On annual bluegrass, chlorotic and necrotic grass was observed at the edge of the rings in July but those damaged areas recovered in August. No symptoms were associated with the rings in creeping bentgrass. Rings were first discovered in June and early July and expanded by about 10 cm in 30 days. New fruiting bodies appeared during July and August as the rings expanded. The application of the fungicide chlorothalnil did not affect the rings. This is the first report of a bird's nest fungus associated with fairy ring in turf.

Reference: (1) H. J. Brodie. 1975. *The Bird's Nest Fungi*. University of Toronto Press, Toronto.

(Disease Notes continued on next page)

Disease Notes (continued)

First Report of the Green Rosette Variant of Groundnut Rosette Disease in Kenya. A. W. Wangai, National Plant Breeding Research Center, Njoro, Kenya; S. S. Pappu and H. R. Pappu, Department of Plant Pathology; University of Georgia, Coastal Plain Experiment Station, Tifton 31793; N. Okoko, Kisii Regional Research Center, P.O. Box 523, Kisii, Kenya; and C. M. Deom and R. A. Naidu, Department of Plant Pathology; University of Georgia, Athens 30602. E-mail: <hrp@tifton.cpes.peachnet.edu> Plant Dis. 83:782, 1999; published on-line as D-1999-0525-02N, 1999. Accepted for publication 19 May 1999.

Groundnut (*Arachis hypogaea* L.) is an important food crop in sub-Saharan Africa. One of the major production constraints is groundnut rosette disease, which is caused by a complex of two viruses, groundnut rosette assistant luteovirus (GRAV) and groundnut rosette umbravirus (GRV) together with the associated satellite RNA (satRNA) (1). Two main forms of the disease have been described: chlorotic and the green rosette. Variants of the satRNA have been shown to be largely responsible for the different forms of the disease (1). Chlorotic rosette has been the predominant form in all of sub-Saharan Africa while green rosette has been reported in the western and southern regions of Africa (2). During the 1997–1998 crop season, disease surveys conducted in Kenya showed the incidence of the rosette disease in farmers' fields to be 24 to 40% in a total of 23 fields surveyed in the western regions of the country (Homabay, Kendubay, Kisumu) and 30% in 8 fields sampled in the Rift Valley (Cheplamus, Marigat) regions. Representative peanut plants showing rosette symptoms were analyzed for the presence of GRV by reverse transcription polymerase chain reaction (RT-PCR). With primers specific to a portion of ORF4 of GRV RNA (3), RT-PCR gave a product of expected size (approximately 300 bp). The PCR product was cloned in pGEM-T vector and sequenced. The sequenced region showed 89% nucleotide sequence identity with published GRV sequences. Green rosette was observed on groundnut cultivars Nyaela Red and Homabay Local in the Kendu Bay region. The incidence of the green rosette was 5.3% of the plants with rosette symptoms.

References: (1) A. F. Murant and I. K. Kumar. Ann. Appl. Biol. 117:85, 1990. (2) R. A. Naidu et al. Ann. Appl. Biol. 132:525, 1998. (3) M. E. Taliensky et al. J. Gen. Virol. 77:2335, 1996.

First Report of Powdery Mildew on *Cornus florida* in Connecticut Caused by *Microsphaera pulchra*. V. L. Smith, Department of Plant Pathology and Ecology, Connecticut Agricultural Experiment Station, New Haven 06504. Plant Dis. 83:782, 1999; published on-line as D-1999-0525-03N, 1999. Accepted for publication 25 May 1999.

Powdery mildew of dogwood (*Cornus florida* L.) has been observed on landscape trees since 1992, and has been increasing in severity recently. In 1998, mildewed leaves ($n = 100$) were collected from each of four locations in Connecticut. White mildew colonies first appeared in the early summer, 4 to 6 weeks after leaf emergence, and the initial colonies rapidly coalesced to cover the entire adaxial leaf surface. Incidence on observed trees reached nearly 100%. Premature autumn coloration and defoliation occurred on those trees where incidence was highest; tree mortality due solely to powdery mildew was not observed. Cleistothecia were apparent on both adaxial and abaxial leaf surfaces within 4 to 6 weeks of initial infection. Mycelium was white in color, and conidiophores were three-celled with a slightly bent foot cell. Fibrosin bodies were not present in conidia ($n = 400$), which measured $14.1 \text{ m} \pm 0.80 \times 28.1 \text{ m} \pm 1.60$ (mean + standard error of the mean). Cleistothecia measured $114.7 \text{ m} \pm 1.77$ in diameter, and had 4 to 10 dichotomously branched appendages, which measured $140.7 \text{ m} \pm 3.95$ in length ($1.23 \times$ cleistothecial diameter). The fungus was identified as *Microsphaera pulchra* based on mycelial, conidial, and cleistothecial characteristics (1). In addition, cleistothecia were found on twigs and on fallen leaves of dogwood; these overwintering cleistothecia contained mature asci and mature ascospores in March 1999. *Microsphaera penicillata* recently was redescribed to include only that species causing powdery mildew on *Alnus* spp., and having appendages $1.5 \times$ cleistothecial diameter (2). The fungus causing powdery mildew in Connecticut previously had been reported to be *M. penicillata*, which was not found in the present study.

References: (1) U. Braun. Nova Hedwigia 89:1, 1987. (2) U. Braun. The Powdery Mildews (Erysiphales) of Europe. Gustav Fischer, New York, 1995.

First Report of Onion Yellow Dwarf Virus in Ontario. L. W. Stobbs and L. Van Driel, Agriculture and AgriFood Canada, Southern Crop Protection and Food Research Center, Vineland Station, Box 6000, Ontario LOR 2E0. Plant Dis. 83:782, 1999; published on-line as D-1999-0603-01N, 1999. Accepted for publication 1 June 1999.

Onion yellow dwarf virus (OYDV) (1) was identified in a commercial planting of garlic (*Allium sativum* L.) near Delhi, Ontario, in 1998. Infected plants exhibited mild mosaic symptoms that became less noticeable by mid-July. Many plants were co-infected with garlic latent virus (GLV) (4), which was mechanically transmitted to leek (*A. porrum* L.). Since leek is not susceptible to OYDV (3), it is an appropriate host for the differentiation of GLV and OYDV. OYDV was transmitted non-persistently by *Myzus persicae* (Schultz) from garlic to garlic and onion (*A. cepa* L.). Infected onion exhibited yellow striping, leaf curling, and pronounced stunting. Necrotic lesions were not present on inoculated leaves of *Chenopodium amaranticolor* Coste & Reyn. and *C. quinoa* Willd., which are often associated with co-infection with leek yellow dwarf virus (2). The isolate reacted strongly in enzyme-linked immunosorbent assay with OYDV antisera (from M. Fukami, Chiba Prefectural Agric. Exp. Stn., Chiba 266, Japan; D. Z. Maat, DLO Research Inst., Plant Prot., Wageningen, The Netherlands). Electron microscopic observations of negatively stained preparations of infected leaf tissues revealed virus particles averaging 765 ± 45 nm with typical pinwheel inclusions.

References: (1) L. Bos. CMI/AAB Descrip. Plant Viruses no. 158, 1976. (2) K. Graichen. Nachrichtenbl. Pflanzenschutz DDR 32: 245, 1978. (3) K. Graichen and H. U. Leistner. Arch. Phytopathol. Pflanzenschutz 23:165, 1987. (4) L. W. Stobbs et al. Plant Dis. 80:343, 1996.

First Report of *Verticillium dahliae* Causing Artichoke Wilt in California. R. G. Bhat and K. V. Subbarao, Department of Plant Pathology, University of California, Davis, 1636 E. Alisal St., Salinas, CA 93905; and M. A. Bari, Artichoke Research Association, 1636 E. Alisal St., Salinas, CA 93905. Plant Dis. 83:782, 1999; published on-line as D-1999-0616-01N, 1999. Accepted for publication 11 June 1999.

In mid-August 1998, artichoke (*Cynara scolymus* L.) plants of cultivar Imperial Star in a field in the Salinas area of the central coast of California developed wilt symptoms. The plants were stunted with chlorotic, drooping, and dried leaves near the bottom and middle of the plants as previously described in Italy (1). Diseased plants produced smaller edible buds and, in severe cases, buds were discolored with dried outer bracts. Roots exhibited the characteristic vascular discoloration of *Verticillium* infection. In one part of the infested field, artichoke was near harvest with 85% of plants showing wilt symptoms with vascular discoloration whereas the other part had a 60-day-old crop with 98% of plants infected. Yield in the field was reduced by as much as 50%. *Verticillium dahliae* was isolated from infected plant samples on NP-10 medium (2), and isolates were single spored before storing on potato dextrose agar at 4°C. Identity of the pathogen was confirmed based on colony morphology and formation of microsclerotia. In root-dip inoculation tests in the greenhouse, two *V. dahliae* isolates from artichoke infected 1-month-old artichoke seedlings that wilted within 6 weeks of inoculation. *V. dahliae* was reisolated from plants showing vascular discoloration. Cross-inoculation studies revealed that artichoke isolates caused a moderate level of disease in lettuce, but only a trace of vascular discoloration in cauliflower. Lettuce isolates caused a severe wilt in artichoke. Cauliflower isolates did not cause wilt in lettuce and caused only slight vascular discoloration in artichoke. This is the first report of a *Verticillium* wilt of artichoke in California.

References: (1) M. Cirulli et al. Plant Dis. 78:680, 1994. (2) L. H. Sorensen et al. Phytopathology 81:1347, 1991.

(Disease Notes continued on next page)

First Report of Crown Gall, Caused by *Agrobacterium tumefaciens* on Geraldton Wax (*Chamelaucium uncinatum*) in South Africa. E. Carstens and Z. Dawood, Directorate Plant Production, Health and Quality, Private Bag X5015, Stellenbosch, 7599, South Africa; E. L. Mansvelt, ARC-Fruit, Vine and Wine Research Institute, Private Bag X5013, Stellenbosch, 7599, South Africa; S. Serfontein, Private Bag X134, Pretoria, 0001, South Africa; and D. G. Malan, Fynflor, 4 Kleineweide, Stellenbosch, 7600, South Africa. *Plant Dis.* 83:783, 1999; published on-line as D-1999-0610-01N, 1999. Accepted for publication 9 June 1999.

Geraldton wax (*Chamelaucium uncinatum*, family Myrtaceae) plants are grown as cutflowers for the export market in South Africa. In July 1998, gall-like structures were observed on collars and roots of Geraldton wax plants in commercial fields in Wellington. The galls were observed after plants exhibited poor growth. The galls varied in size and in texture from soft and spongy to hard. Secondary symptoms involved poor root development and browning of stem tissues near galls. Isolations from the galls yielded nearly pure cultures of a Gram negative, rod-shaped bacterium on Roy Sauer medium (2), typical of an *Agrobacterium* sp. Carbon source utilization testing with the Biolog GN Bacterial Identification System (version 3.50) confirmed the bacterium as a biovar of *A. tumefaciens* with a similarity of 0.88. Pathogenicity was confirmed by injecting 4- to 6-week old tomato and tobacco plants and 1-year-old Geraldton wax plants with approximately 5 µl of the bacterial suspension (10⁸ CFU/ml) in sterile, distilled water. Inoculated plants were then transferred to a greenhouse at 25°C. Galls developed 1 month after inoculation. The bacterium was readily reisolated from the inoculated plants. *A. tumefaciens* is endemic to South Africa and has a very wide host range that includes several ornamentals (1). This is the first report of *A. tumefaciens* on Geraldton wax plants in South Africa.

References: (1) J. F. Bradbury. 1986. Guide to Plant Pathogenic Bacteria. CAB Int., Slough, U.K. (2) N. W. Schaad. 1988. Laboratory Guide for Identification of Plant Pathogenic Bacteria. The American Phytopathological Society, St. Paul, MN.

A Serious Outbreak of Blast Disease (Gray Leaf Spot) of Perennial Ryegrass in Golf Course Fairways in Pennsylvania. W. Uddin, M. D. Soika, F. E. Moorman, and G. Viji, Department of Plant Pathology, The Pennsylvania State University, University Park 16802. *Plant Dis.* 83:783, 1999; published on-line as D-1999-0614-02N, 1999. Accepted for publication 8 June 1999.

Blast disease (gray leaf spot) of perennial ryegrass (*Lolium perenne* L.), caused by *Pyricularia grisea* (Cooke) Sacc., in golf course fairways in Pennsylvania, was first reported in 1991 (1). The disease was primarily confined to the southeastern region of the state. In August and September 1998, severe outbreaks of blast disease occurred in perennial ryegrass fairways in numerous golf courses in the same region in Pennsylvania. Additionally, in 1998, the disease was diagnosed in ryegrass fairways for the first time in the western region of the state, where serious disease outbreaks occurred. The disease develops on leaf blades as small, water-soaked lesions that become necrotic spots. The spots expand rapidly, and develop into gray, grayish-brown, or light brown, circular spots with purple to dark brown borders often surrounded by a yellow halo on the leaf blades. As the disease progresses, the circular necrotic spots coalesce, become irregular in shape, and cause partial (tip blight) or complete blighting of the leaf blades. The blighted blades may often appear twisted. Turfgrass loss due to the disease was over 90% in several golf courses. During the 1998 epidemics, a survey was conducted in the affected regions, which included three golf courses in western Pennsylvania and 15 golf courses in southeastern Pennsylvania. *P. grisea* was con-

sistently isolated from the symptomatic ryegrass leaf blades of turf samples (12 to 28 samples per location) collected from the blighted fairways. Of the 122 isolates of *P. grisea* collected from the 18 golf courses, seven isolates (two from western Pennsylvania and five from southeastern Pennsylvania) were selected for pathogenicity assays. Five Pennfline perennial ryegrass plants were grown in each Cone-Tainer (4 mm in diameter) that was filled to 1 cm below the rim with granular calcine clay medium (Turface MVP; Allied Industrial Material, Buffalo Grove, IL). Three weeks after seeding, grasses were fertilized with water-soluble 20-20-20 fertilizer (1.3 g per liter of water) once a week to field capacity of the growing medium. Treatments (the isolates) were arranged as a randomized complete block with four replications (a Cone-Tainer per replication). Six-week-old ryegrass plants were atomized with an aqueous suspension of *P. grisea* conidia (approximately 8 × 10⁴ conidia per ml of sterilized, distilled water) until the leaves were completely wet. Individual Cone-Tainers were placed in clear polyethylene bags, enclosed, and were placed in an incubator that was maintained at continuous 28°C and 12-h-day fluorescent light (72 µE s⁻¹ m⁻²). Three days after inoculation, water-soaked lesions (<2 mm in diameter) developed on leaves of the ryegrass inoculated with each isolate of *P. grisea*. No lesions developed on leaves of the control plants. Seven days after inoculation, the polyethylene bags were removed, and disease incidence (percent infected leaves) was assessed. *P. grisea* was isolated from the necrotic lesions or the blighted leaf blades of every plant inoculated with the fungus. This is the first report of blast disease outbreak in golf course fairways in western Pennsylvania. The impact of blast disease epidemics on golf courses in the northeastern United States in 1998 was significant, and caused serious concern to turf managers. The survey indicates that blast disease of perennial ryegrass may be emerging as a new problem for the turfgrass industry in the northeastern United States.

Reference: (1) P. J. Landschoot and B. F. Hoyland. *Plant Dis.* 76:1280, 1992.

First Report of *Cercospora traversiana* on *Trigonella coerulea* in Bulgaria. S. G. Bobev, Higher Agricultural Institute, 4000 Plovdiv, Bulgaria; A. F. Margina, Research Institute for Roses, Essential and Medicinal Plants, Kazanlak, Bulgaria; and J. de Gruyter, Plant Protection Service, The Netherlands. *Plant Dis.* 83:783, 1999; published on-line as D-1999-0614-01N, 1999. Accepted for publication 11 June 1999.

In the spring of 1996, diseased plants of *Trigonella coerulea* were noted in an experimental area at Kazanlak, Bulgaria. The primary symptoms were leaf spots 8 to 2 mm in diameter, light brown then becoming gray and slightly zonate, and surrounded by a diffuse chlorotic margin. When single lesions occur the disease normally develops as a typical leaf spot. However, the development of more than one spot is followed rapidly by yellowing and withering of entire leaflets. Stem lesions usually begin at the point of leaf attachment, extend both directions about 15 to 20 mm, and often encircle the stems. Isolations on potato dextrose agar (PDA) yielded a slow-growing, gray fungus. Conidia from the isolated fungus when inoculated at 3.10⁴ spores per ml on potted *T. coerulea* seedlings and kept in a moisture chamber for 48 h caused foliar spots in 5 to 8 days and sporulating structures similar to those seen in field observations. Simultaneous inoculation of *T. foenum-graecum* (fenugreek) plants produced similar symptoms, but the percentage of successfully inoculated leaves (3.6%) was lower than in *T. coerulea* (27.6%). Conidiophores of the fungus are dark, arising in clusters, unbranched, septate (1 to 3), and have small conidial scars. Conidia are hyaline, straight, multicelled (1 to 17), tapering at the base, and measured 30 to 212 × 4 to 6 µm. The pathogen was identified as *Cercospora traversiana* Sacc. and this is the first report of its occurrence in Bulgaria.