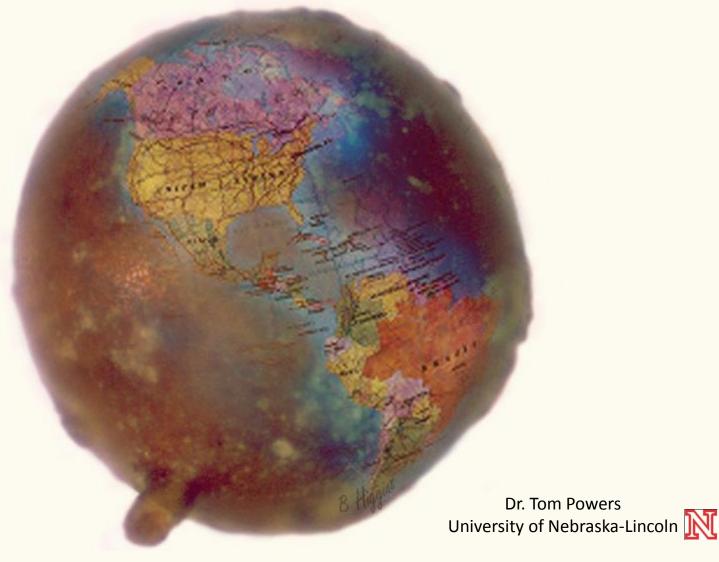
Recovery Plan for Root-knot and Cyst Nematodes

Parasites of Agronomic and Horticultural Plants throughout North America





Number of Species Worldwide :

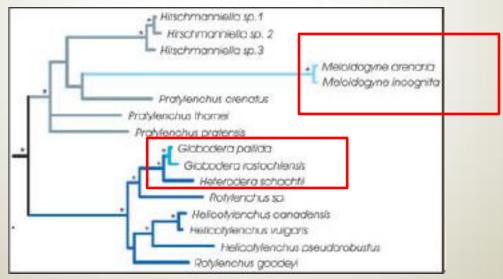
Root-knot Nematodes: ~100

Meloidogyne- 44 species in GenBank

Cyst Nematodes: ~100

Heterodera- 48 species in GenBank Globodera- 10 species ""

Evolutionary divergence- from early Cenozoic?



Holterman et al., 2006



Similarities and Differences between Root-knot Nematodes and Cyst Nematodes



Large females

Cannot migrate very far under their own power

Must establish feeding sites within the plant root

Subject to many antagonistic and predatory organisms in the soil pore space ("suppressive soil")

Include both males and females (although many *Meloidogyne* species are parthenogenetic)



Eggs **RKN-** deposited in the soil Cyst- encased in the cyst

Hatch

RKN- readily hatch in the soil Cyst- require presence of hatch-inducing chemicals

Survival

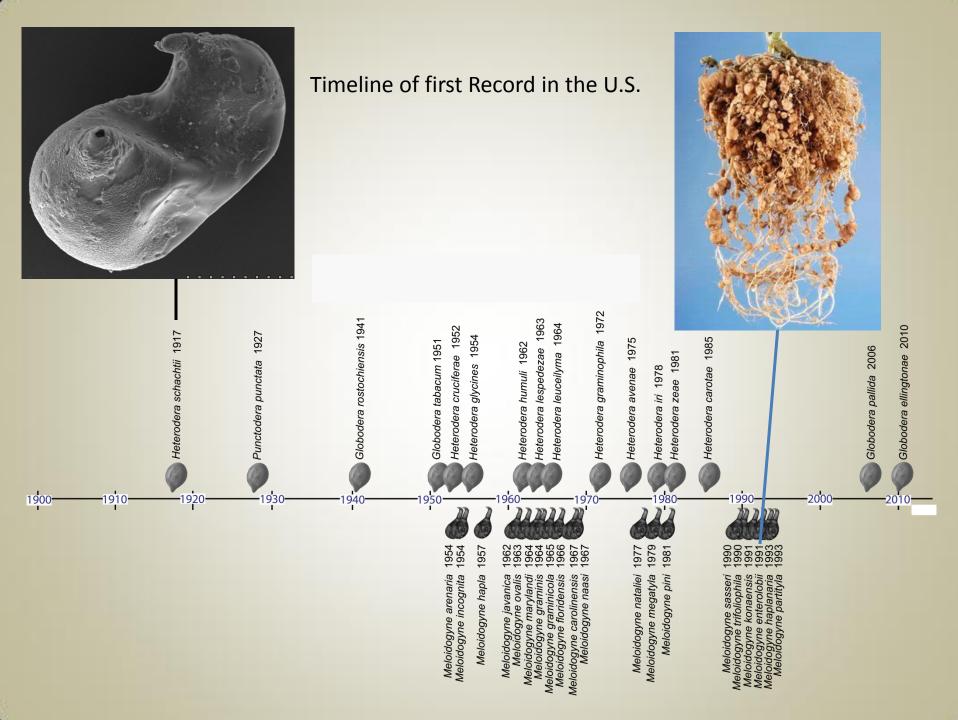
RKN- no specialized structures Cyst-highly resistant casing

Host range RKN- wide (400+) host range

Cyst-limited host range (ie: H. carotae exists only on carrot)







Above ground: wilt, nutrient deficiency, stunting, uneven growth

M. chitwoodi juvenile

Symptoms Root-knot Nematode

1º Com





Below ground: galls, egg masses





Tomato roots: RKN galls (left) healthy (right)

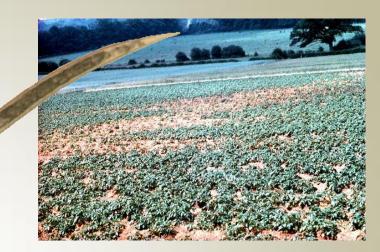


Above ground: wilt, nutrient deficiency, stunting, uneven growth



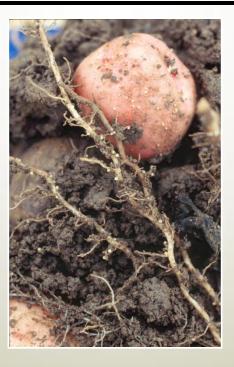
Symptoms Cyst Nematode

H. graminophila juvenile



Below ground: cysts







Historical Case Studies

Golden Potato Cyst Nematode G. rostochiensis (New York)

- Post WWI on military equipment
- NY 1934 observation
- State/Federal quarantines
- Quarantine success story

Mitigation point #1. There generally exists a significant lag-time between nematode introduction and nematode detection.

Soybean Cyst Nematode H. glycines (Eastern and Central U.S.)

- NC 1954 observation
- Turn of century importing of Asian soil
- East of Rockies distribution

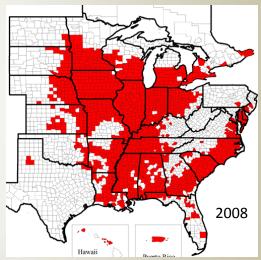
Mitigation point #2. Once established, it is extremely difficult to prevent further spread of plant-parasitic nematodes.

Columbia Root-knot Nematode M. chitwoodi (Western U.S.)

- 1981 Washington state
- Asymptomatic infected seed pieces

Mitigation point #3. Endoparasitism and asymptomatic infections by root-knot nematodes emphasize the need for soil surveys to detect infective juvenile stages.







Newly Emergent Case Studies

Cyst nematodes on potato *G. pallida* and *G. ellingtonae* (Idaho and Oregon)

- 2006- Immediately closed markets in Canada, Mexico, Korea, Japan
- To date 17 infected fields, 2015 acres
- Discovered as part of a CAPS survey, in a non-standard tare sample
- *G. ellingtonae* status?

Mitigation point #4. Eradication, if possible, will require an expensive, highly regulated, large-scale operation that will include multiyear applications of general biocides.

Meloidogyne enterolobii (syn. M. mayaguensis) (Atlantic coast)

- Discovered in 1991 through reproduction on Mi gene
- Cryptic species, highly aggressive
- Reproduces on nematode resistant germplasm

Mitigation point #5. The next major nematode pest may emerge from populations already resident in the U.S.





M. enterolobii on gardenia

Poll Results of Professional Nematologists with field experience (43 Respondents of 56 contacted)

Question #1. Do you think it is likely that within 5-10 years, novel species or races of nematodes will be encountered in the U.S. that are capable of causing economic damage to our agricultural or horticultural crops?

- 100% of respondents felt it was likely or highly likely or inevitable that U.S. agriculture will be confronted with new economically damaging nematodes.
- Over 50% of respondents expressed the opinion that the development of new races or pathotypes, or the redistribution of species currently in the U.S. was of equal concern to exotic introduction.

Question #2. If a new potentially damaging species is introduced, do you think we have the knowledge, infrastructure, and resources to limit its damage?

- Several respondents identified the rapid and comprehensive actions be APHIS following the 2006 discovery of *G. pallida* in Idaho, as a model for addressing a potentially destructive nematode species.
- States that lack a trained field nematologist will impair early detection and management efforts.
- Lack of training opportunities

Question #3. In managing the existing pest nematode species in the U.S., would you say we are winning the battle, staying even, or losing the battle?

- 18 Respondents thought we are losing the battle
- 19 Respondents said we are staying even
- General frustration with post-nematicide era management options

Recovery Plan Realities

- 1. Recent history clearly indicates that new species will be introduced into the U.S.
- 2. It is highly likely that new species or genotypes already exist in the U.S. presently undetected.
- 3. There will be a significant lag time between nematode introduction and detection.
- 4. Early detection of established infestations is critical for mitigation success.
- 5. Many states to not have personnel trained to act as nematode infestation "first responders."
- 6. Opportunities for training in nematology are decreasing.
- 7. USDA APHIS/PPQ has the incidence command structure to rapidly respond to new nematode detections. States do not.
- 8. Once a nematode species is widely established it is practically impossible to eradicate.
- 9. The management tools of 2013 are essentially the same as they were 50 years ago, minus the variety of chemical nematicides.
- 10. The current management tools have greater precision but require an increased understanding of the site-specific nematode problem.
- 11. Genetic resistance is available for some crops and some regions and effective against some nematode genotypes.
- 12. No one knows how climate change will affect future nematode management.



Detection, surveys and identification - Recommendations

- 1. Expand the nematode survey component of the CAPS program.
- 2. Build reference databases to facilitate rapid identification and geographic location of species.
- 3. Increase taxonomic resolution of ongoing surveys to accurately record endemic species, regional diversity, host-races, and resistance-breaking genotypes.
- 4. Encourage the development of more SOPs for nematode identification.
- 5. Increase Nematology training of diagnostic "first responders."
- 6. Increase resources for pest risk assessment models and establish linkages between modelers and nematologists through Multistate projects.
- 7. Support the generation of biological, developmental, physiological, and environmental parameters for model development and improvement of management tactics .



Management tools- Recommendations

- 1. Provide incentives for participants in Multistate/Regional Nematology projects to work jointly on specific integrated management approaches.
- 2. Just as nematode "first-responders" require training, field-savvy nematologists need to educate the next generation of nematologists to facilitate the implementation of integrated management.
- 3. Establish a nationwide program of field nematology internships.
- 4. Support broad-based approaches nematode management, if only as a backup for potential failure of "silver-bullet" solutions.
- 5. Use the Society of Nematologists as organizing body to facilitate recommendations.



Selected Nematology poll respondents and sources for Nematology information

George Abawi, Cornell University Byron Adams, Brigham Young University Ole Becker, University of California George Bird, Michigan State University Janete Brito, Division of Plant Industry, Florida Lynn Carta, USDA-ARS Beltsville, Florida William Crow, University of Florida Eric Davis, North Carolina State University Donald Dickson, University of Florida Patricia Donald, USDA-ARS Jackson, Tennessee Jonathan Eisenbeck, Virginia Tech Axel Elling, Washington State University Robin Giblin-Davis, University of Florida Howard Ferris, University of California-Davis John Halbrendt, Penn State University Russell Ingham, Oregon State University Tamra Jackson-Ziems, University of Nebraska James Kotcon, West Virginia University Gary Lawrence, Mississippi State University Kathy Lawrence, Auburn University

Mike McClure, University of Arizona Michael McKenry, University of California-Riverside Robert McSorley, University of Florida Haddish Melakeberhan, Michigan State University John Mueller, Clemson University Deborah Neher, University of Vermont James Noe, University of Georgia Joe Noling, University of Florida Andrew Nyczepir, USDA-ARS Byron, Georgia Charles Overstreet, Louisiana State University Antoon Ploeg, University of California-Riverside Robert Robbins, University of Arkansas Brent Sipes, University of Hawaii James Starr, Texas A&M University Steve Thomas, New Mexico State University Patricia Timper, USDA-ARS Tifton, Georgia Tim Todd, Kansas State University Greg Tylka, Iowa State University Inga Zasada, USDA-ARS Corvallis, Oregon