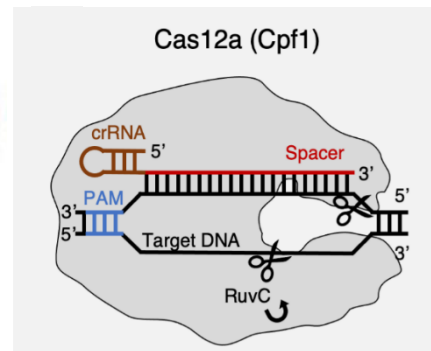


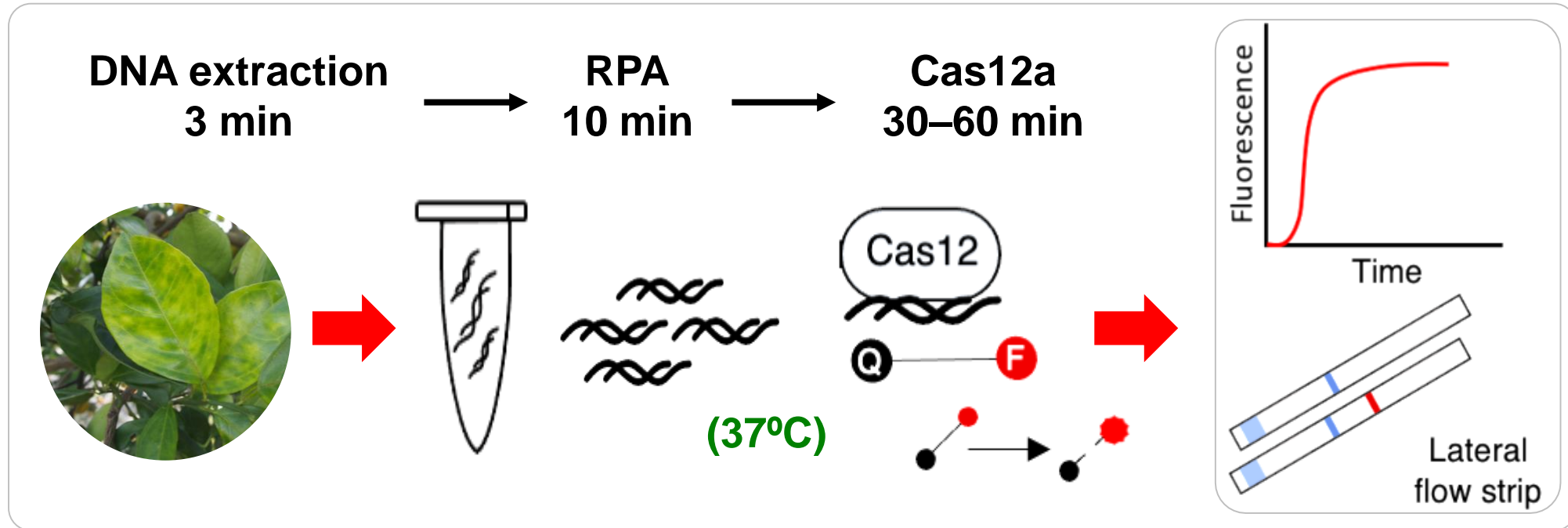
# Supersensitive and Specific Detection of Citrus Greening and Phytoplasmal Pathogens with RPA/Cas12a

Yinong Yang

Department of Plant Pathology and Environmental Microbiology,  
Huck Institutes of the Life Sciences, The Pennsylvania State University



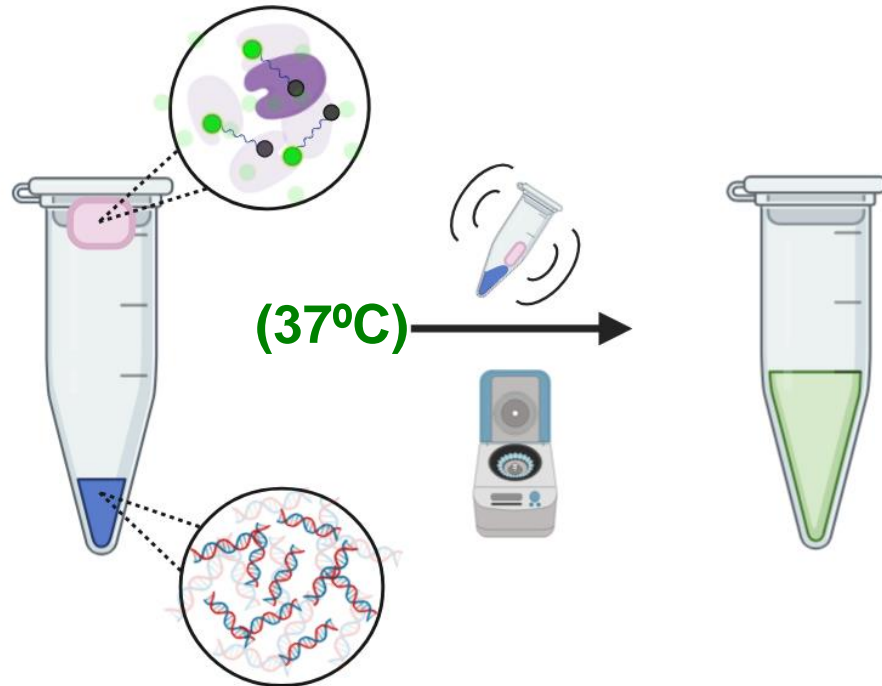
# Isothermal RPA/Cas12a Assay for Plant Disease Diagnostics



**RPA/Cas12a: recombinase polymerase amplification/Cas12a nuclease**  
**DETECTR: DNA endonuclease-targeted CRISPR trans reporter**

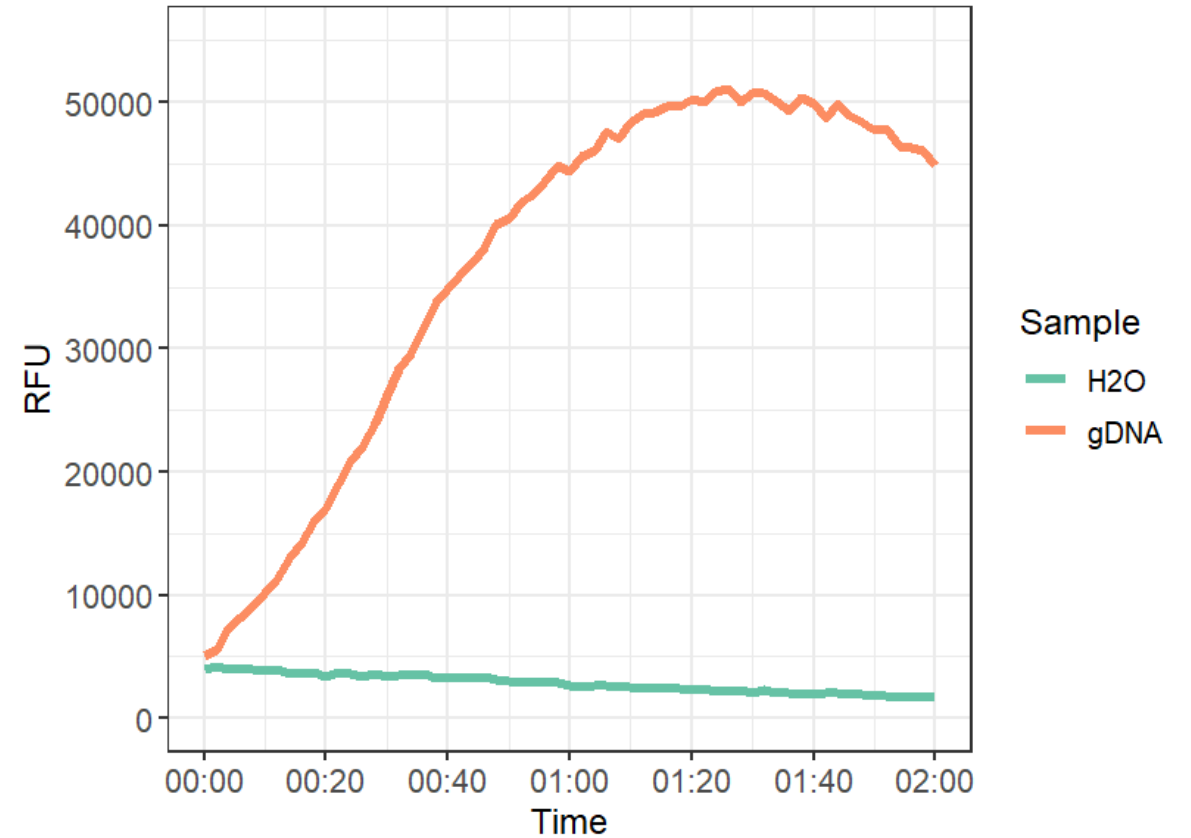
# One-pot RPA/Cas12a Detection Assay

10  $\mu$ l Cas12a reagent mix



10  $\mu$ l RPA reaction

HLB One Pot DETECTR



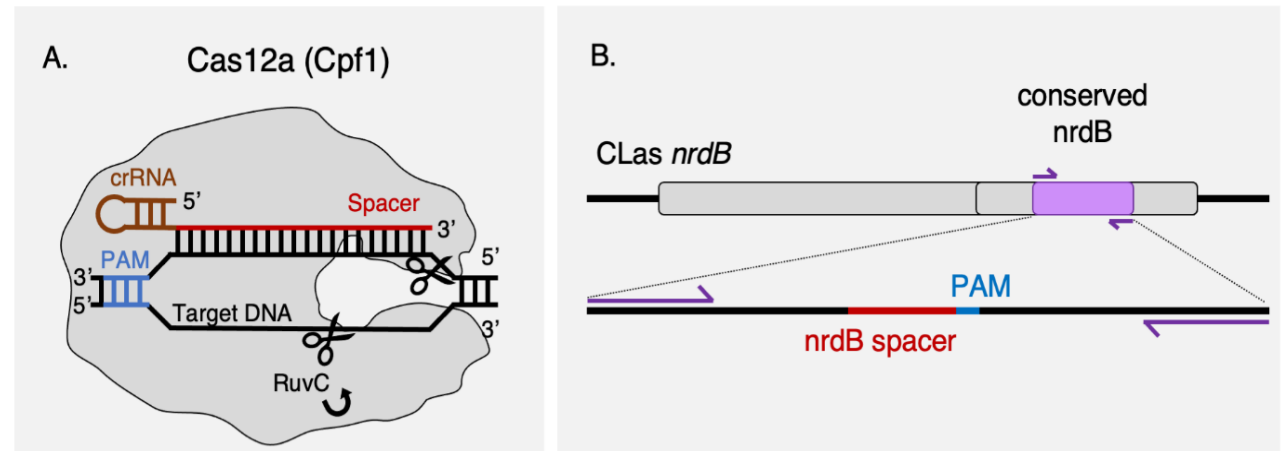
# RPA/Cas12a Detection of Citrus Greening Pathogen



## *Candidatus Liberibacter asiaticus* (CLas)

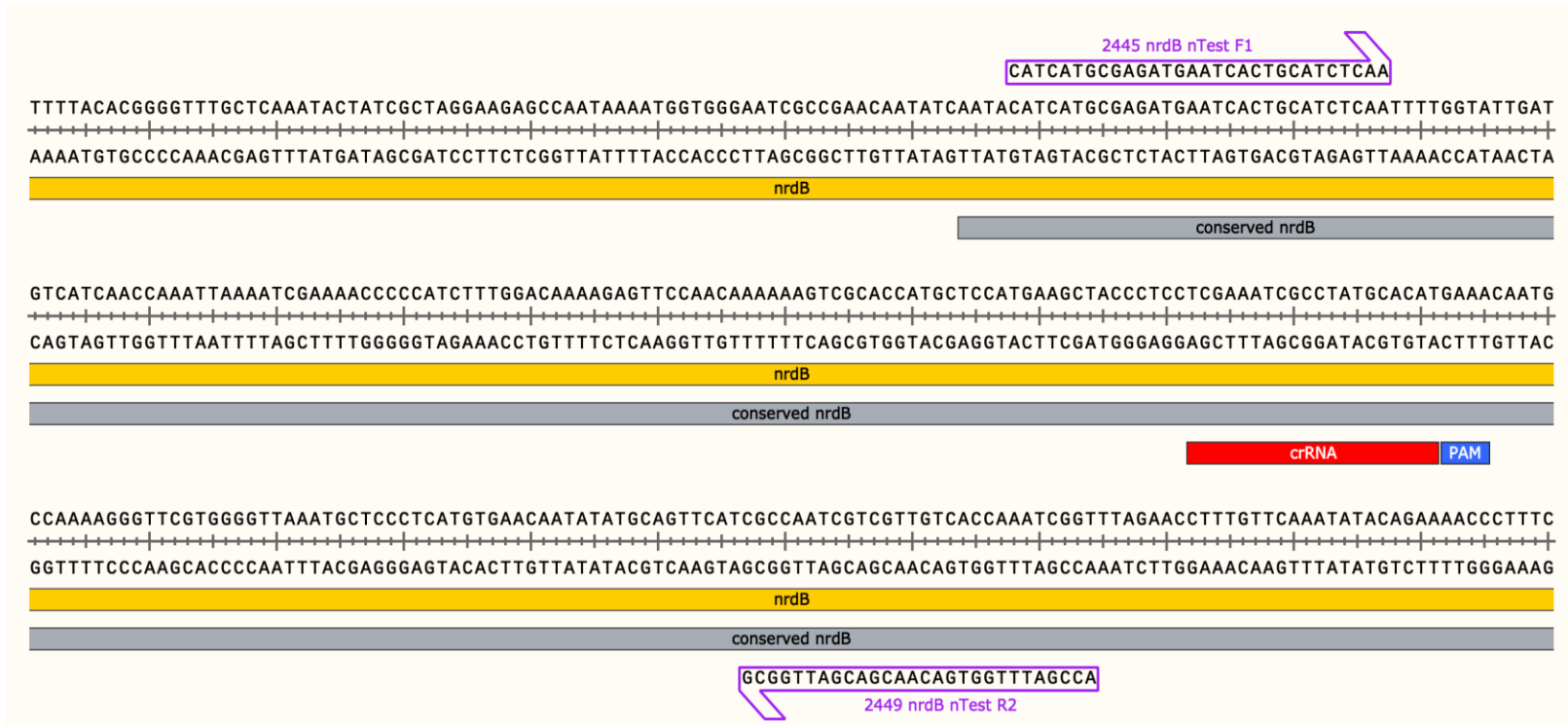
- ✓ *CLas* contains five copies of *nrdB* that encodes the  $\beta$ -subunit of conserved ribonucleotide reductase (RNR)
- ✓ *nrdB* loci enable robust detection of *CLas* using qPCR with 3 times more sensitivity than 16S rDNA

(Zheng et al 2016 Scientific Reports)

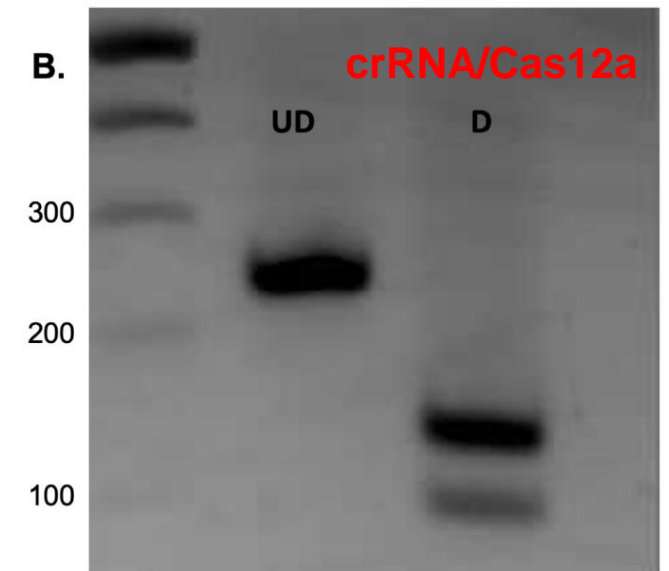
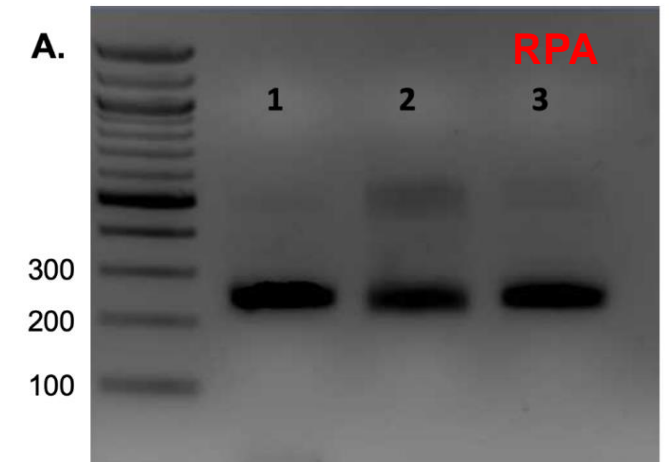


(Wheatley and Yang 2021 Phytopathology)

# Evaluation of RPA Primers and crRNA/Cas12a DNA Cleavage Efficiency

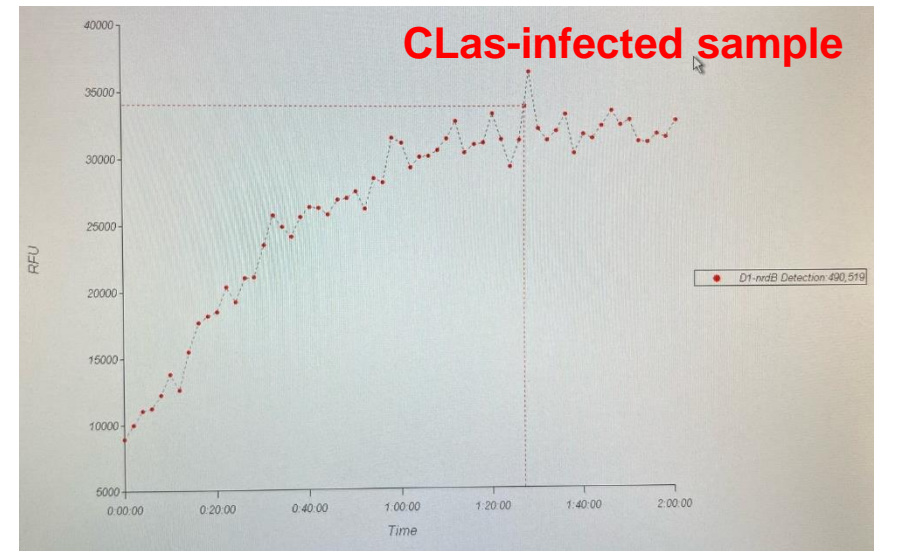
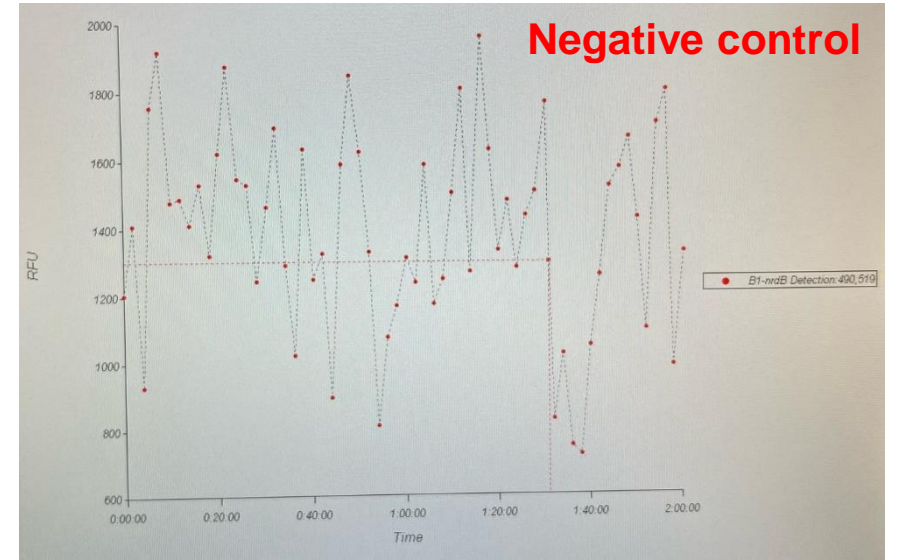
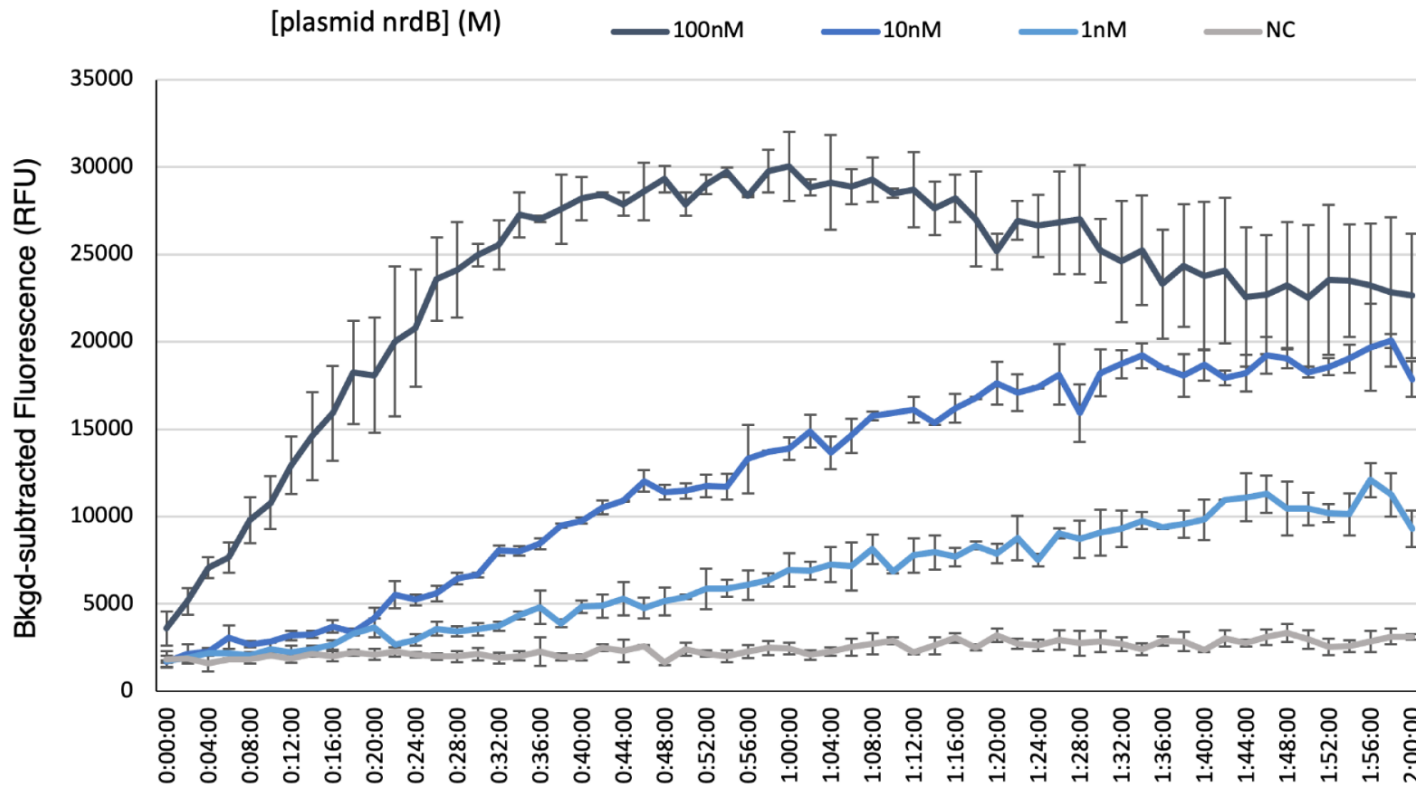


247 bp amplicon

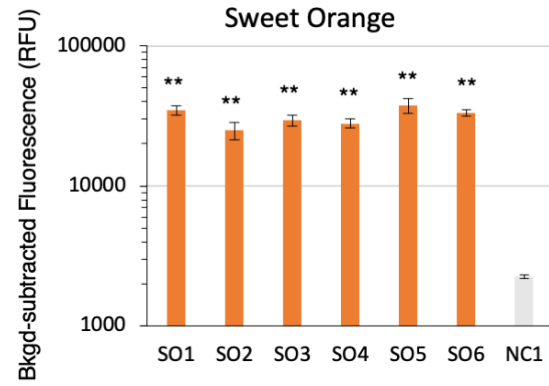




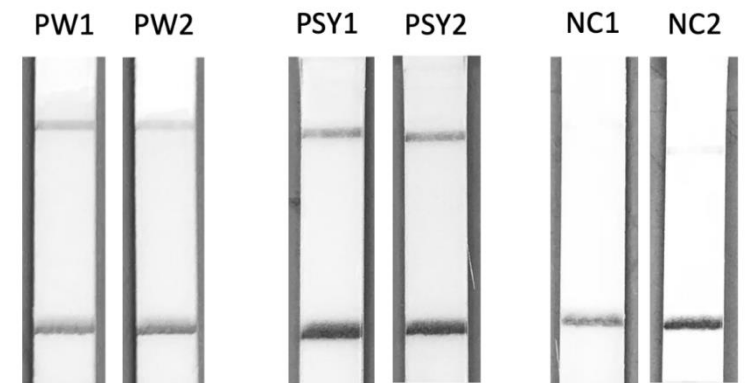
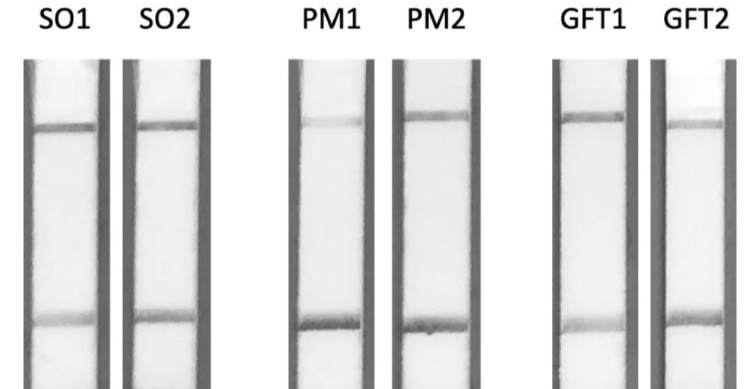
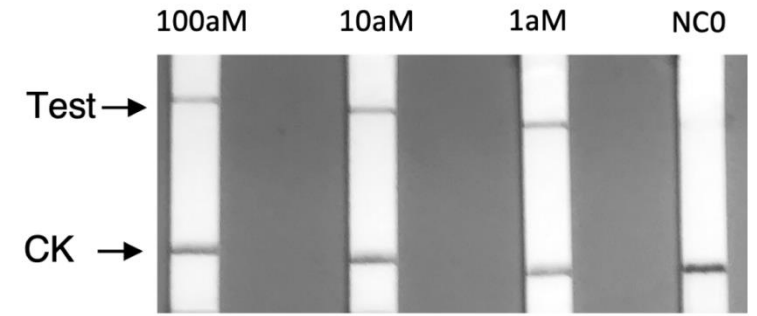
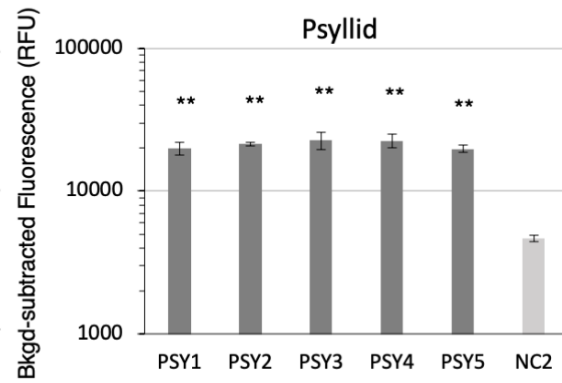
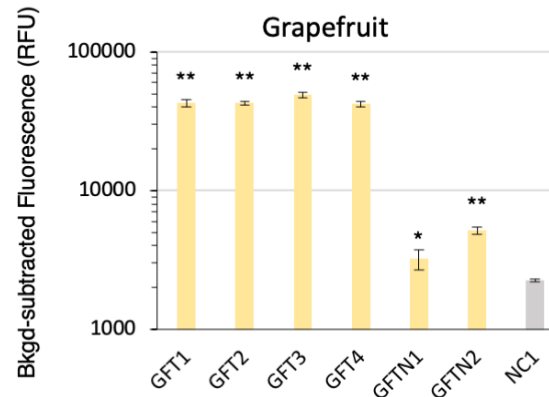
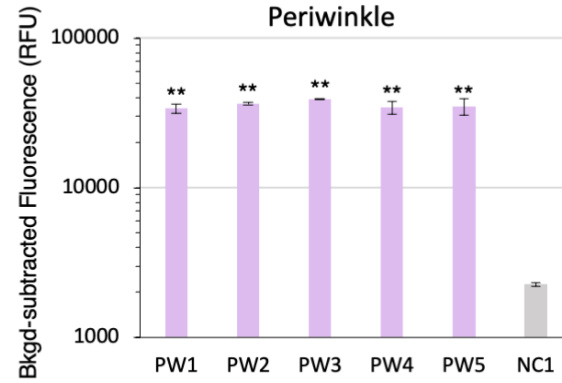
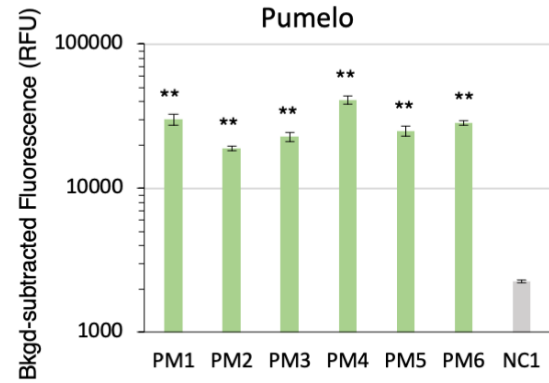
# Cas12a Detection of *nrdB* DNA



(Wheatley et al 2021 Phytopathology)



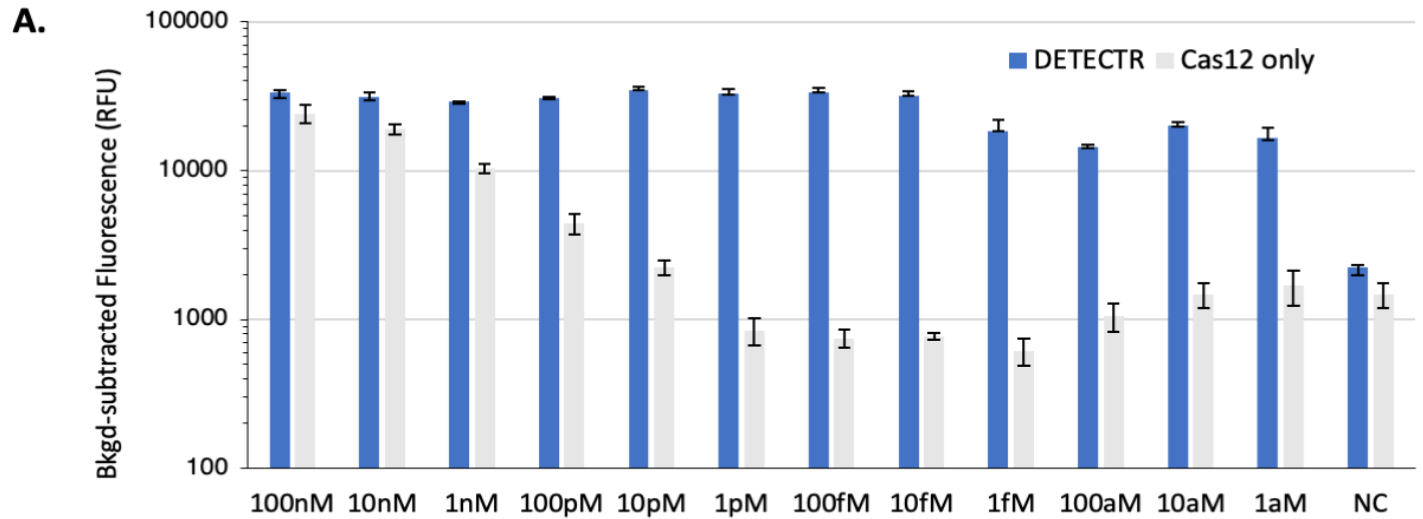
\* = p < 0.05  
 \*\* = p < 0.001



**Fluorescence Microplate Reader**

**Lateral Flow Assay**

# Detection Sensitivity of RPA/Cas12a vs SYBR Green qPCR



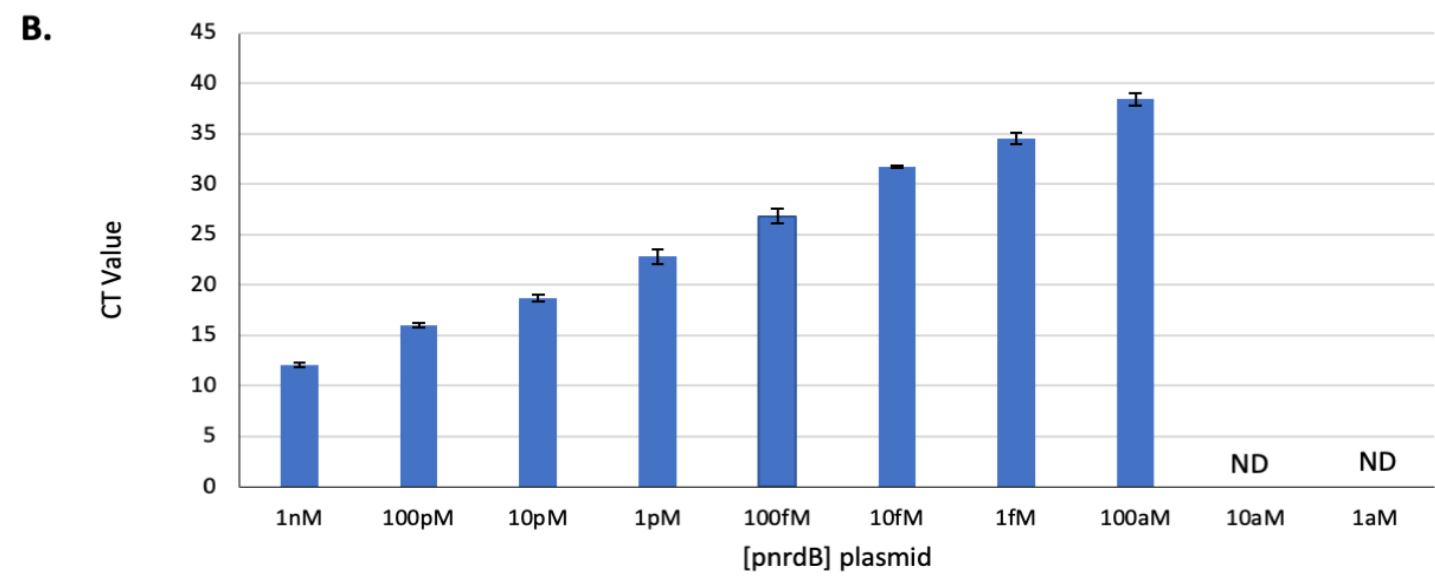
**DETECTR: DNA endonuclease-targeted CRISPR trans reporter (i.e., RPA/Cas12a)**

**NC: negative control**

**ND: not detected**

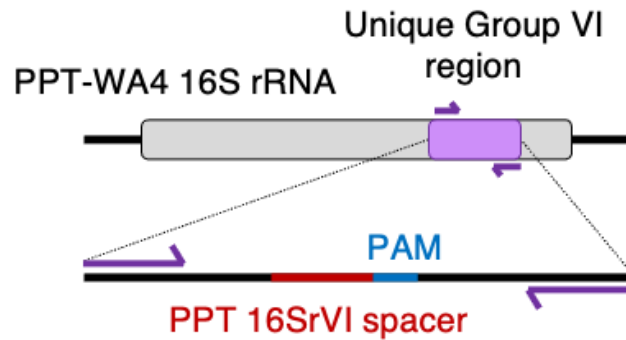
**Error bars represent mean  $\pm$  s.d. when  $n = 3$  replicates**

**(Wheatley et al 2021 Phytopathology)**





# Potato Purple Top Disease



PPT 16S-23S ITS Amplicon – 156 bp

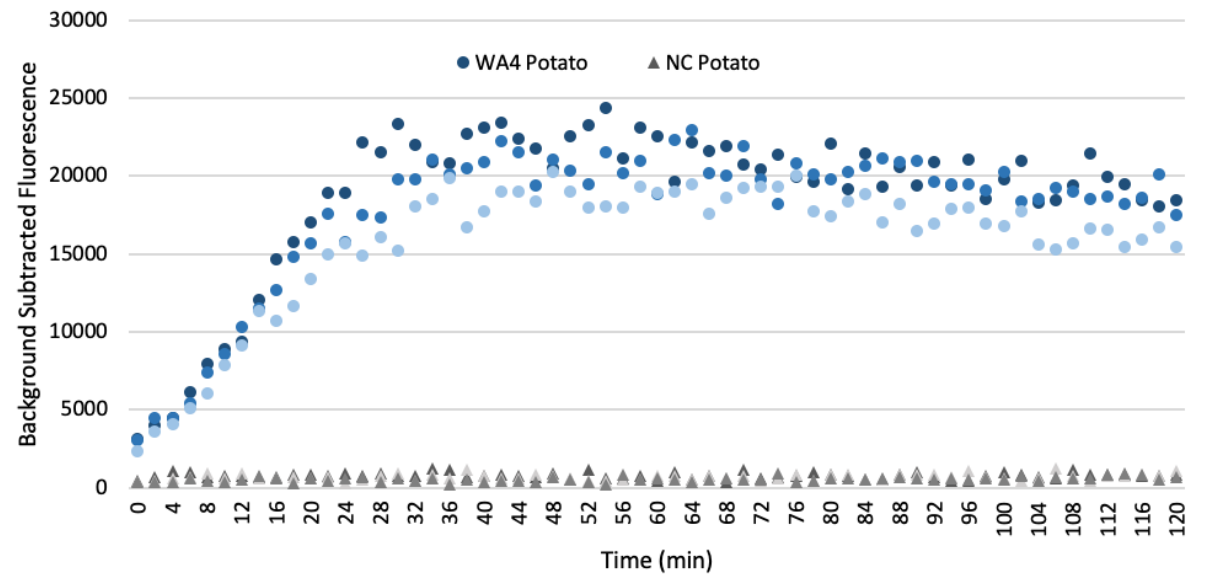
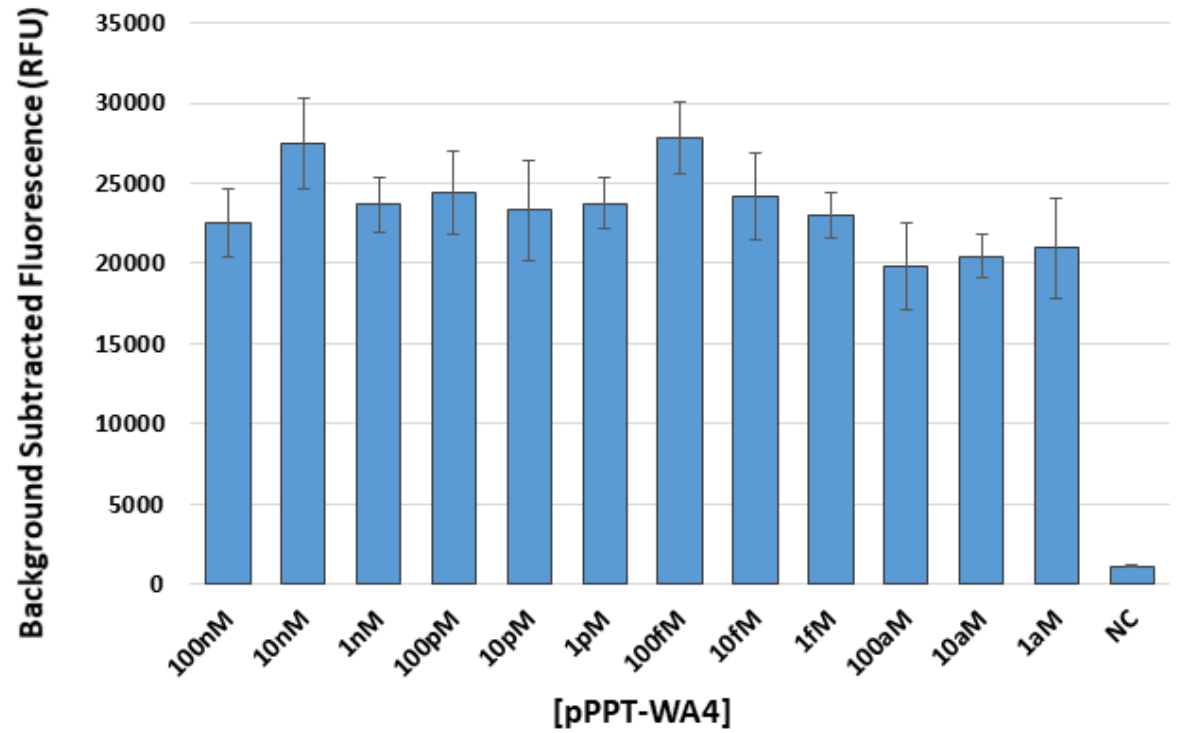
**TGATAAGCGTGAGGTCGGTGGTTCAAGT**  
**CCATTTAGGCCACCAAAAAAGGTCCT**  
**GCTTAAGAAGTTCTTTGAAAAGTAGGTA**  
**AACATGCTTTAAAATTTTCATAAAGTTG**  
**AAGGAAGTAAGGCATATAGTGGATGCC**  
**TTGGACTAAGA**

*Candidatus Phytoplasma trifolii*  
related strains (16SrVI)



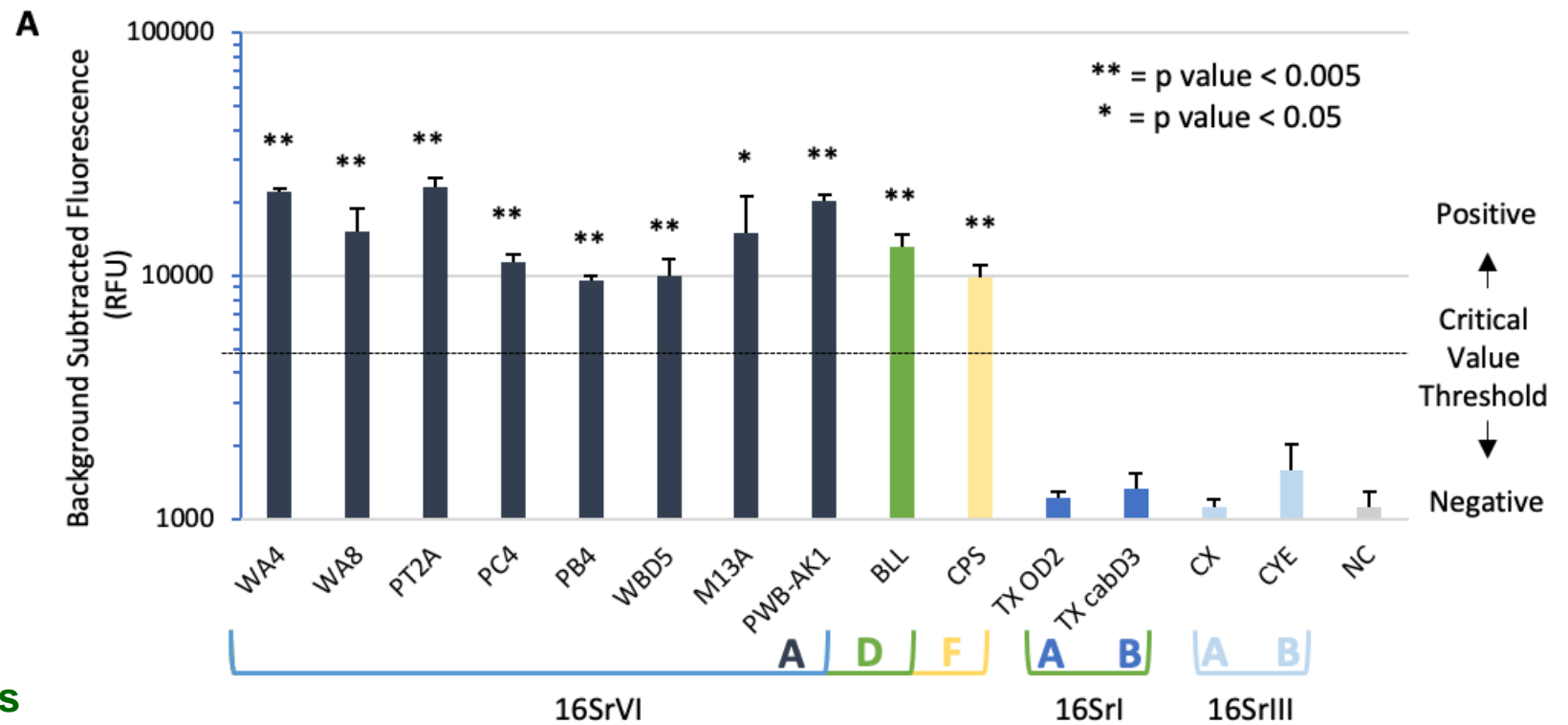
# Supersensitive and Specific Detection of 16S-23S ITS Target from PPT-WA4

(Wheatley et al 2022 Plant Disease)

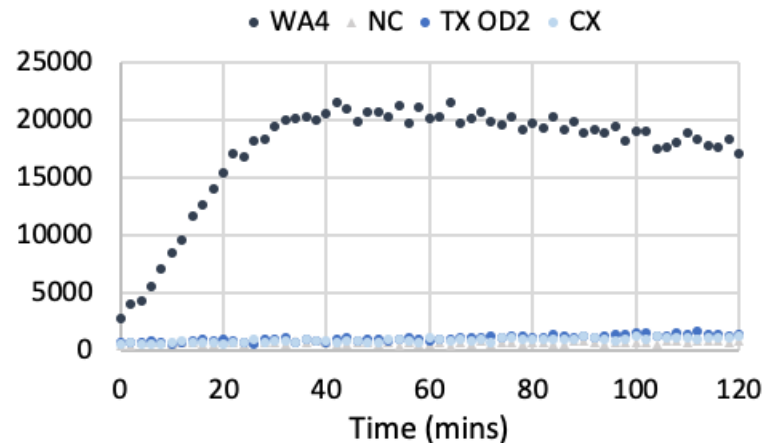


# Specific Detection of Group 16SrVI Phytoplasma by RPA/Cas12a

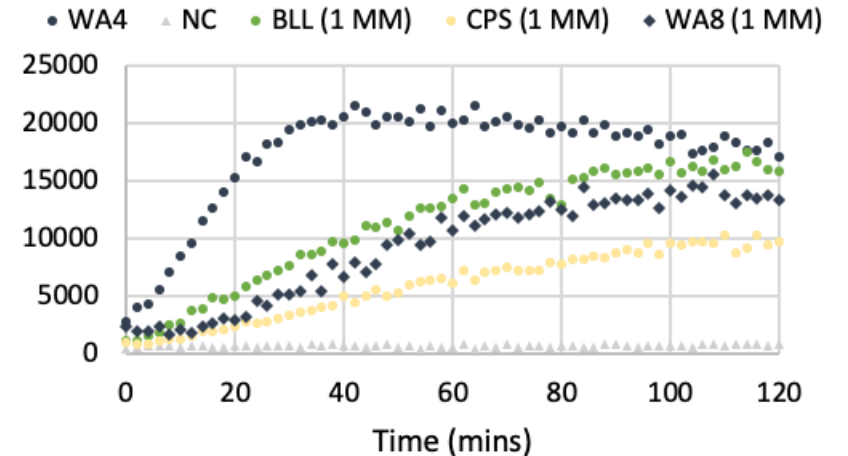
- Cas12a can still cleave protospacers with a single mismatch (MM)
- Turnover is lower, resulting in delayed signal intensity
- More than a single MM abolishes cleavage activity



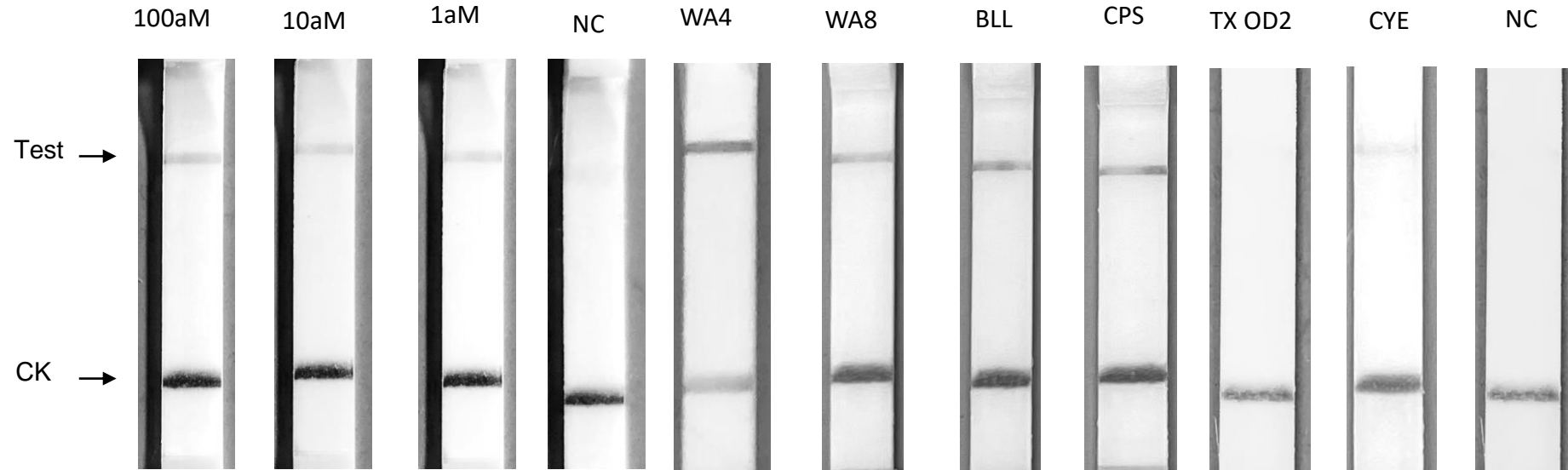
**B** Group 16Sr VI vs Group 16Sr I and 16Sr III



**C** Group 16Sr VI-A vs Group 16Sr VI-D and Group 16Sr VI-F



# RPA/Cas12a Detection with Lateral Flow Assay

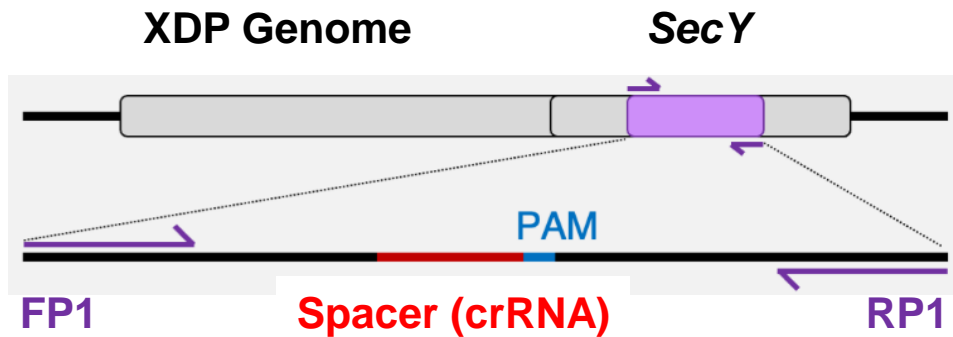


(Wheatley et al 2022 Plant Disease)

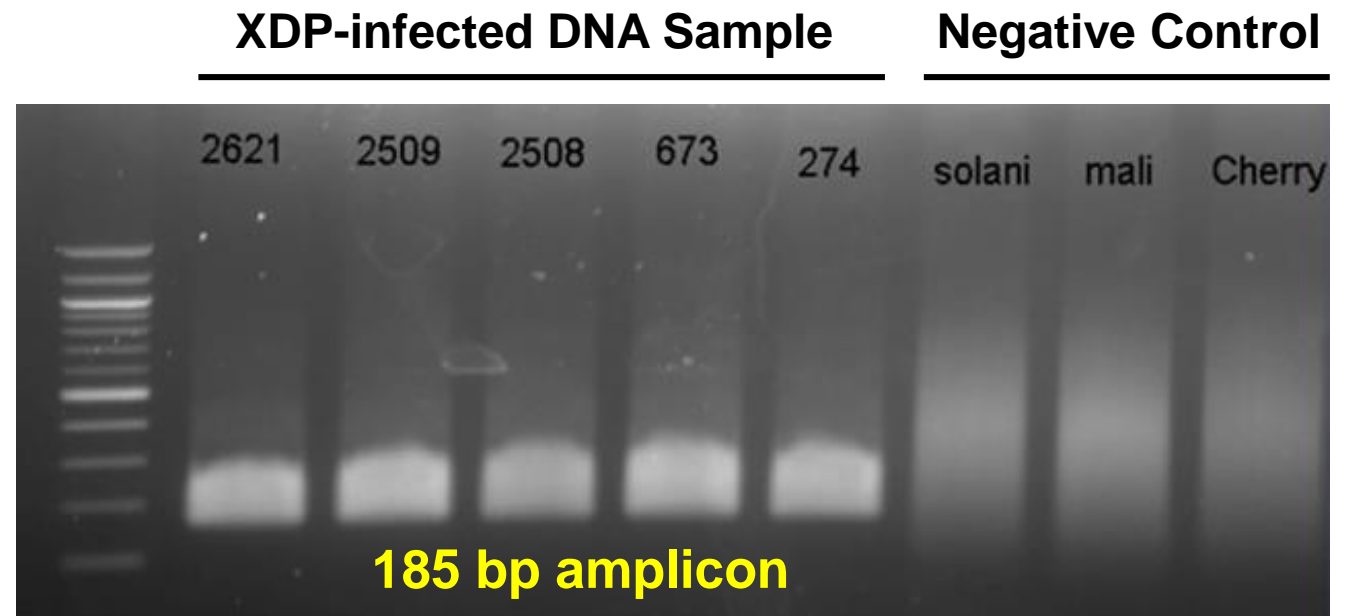
# RPA/Cas12a Detection of Cherry X Disease Phytoplasma



*Candidatus Phytoplasma pruni* (16SrIII)

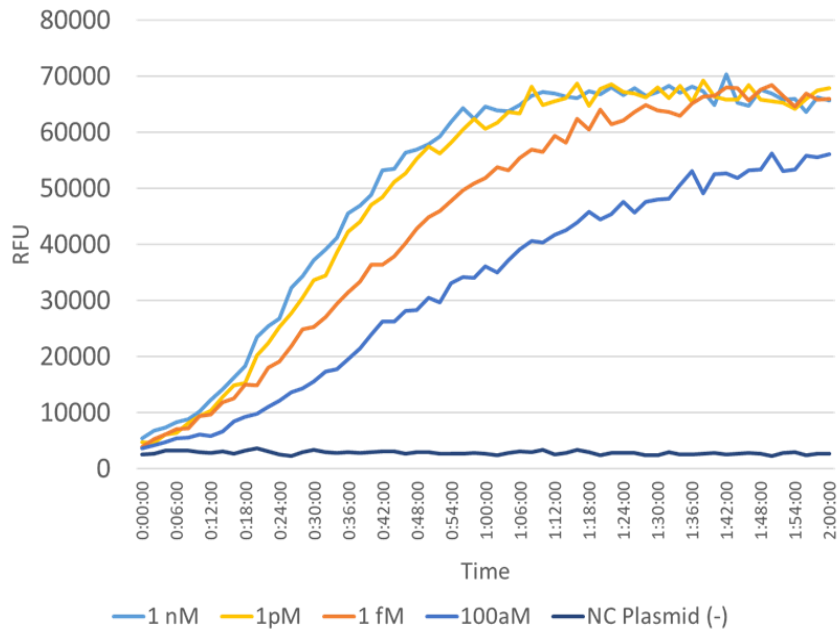


- ✓ Alignment of *SecY* sequences across the 16SrIII subgroups of phytoplasmas
- ✓ Identification and evaluation of highly specific RPA primer pairs and crRNAs

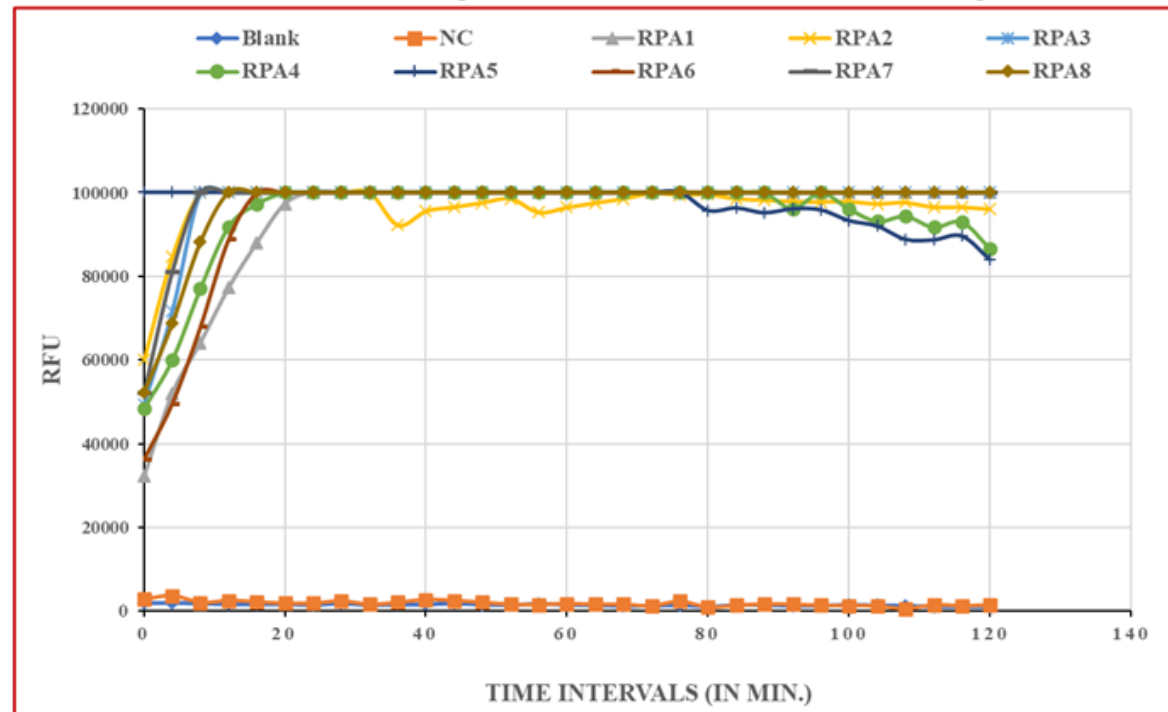


Recombinase Polymerase Amplification of XDP DNA

# Development of Specific RPA/Cas12a Assay for Sensitive Detection of XDP DNA



Pure DNA sample with estimated DNA copies



Sample	Copy no.
RPA1	1
RPA2	10
RPA3	100
RPA4	1000
RPA5	10000
RPA6	100000
RPA7	1000000
RPA8	10000000

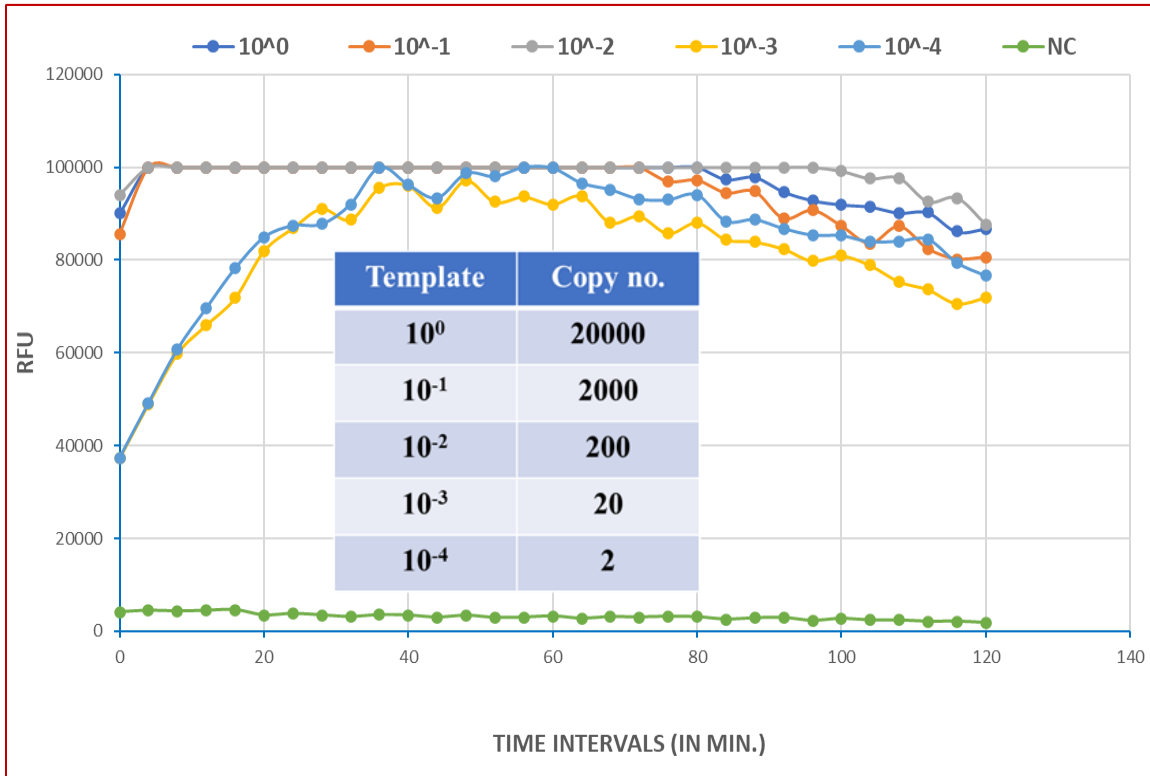
NC: Negative control (non-infected sample); Blank: Water.

The max fluorescence reading is 100000 RFU.



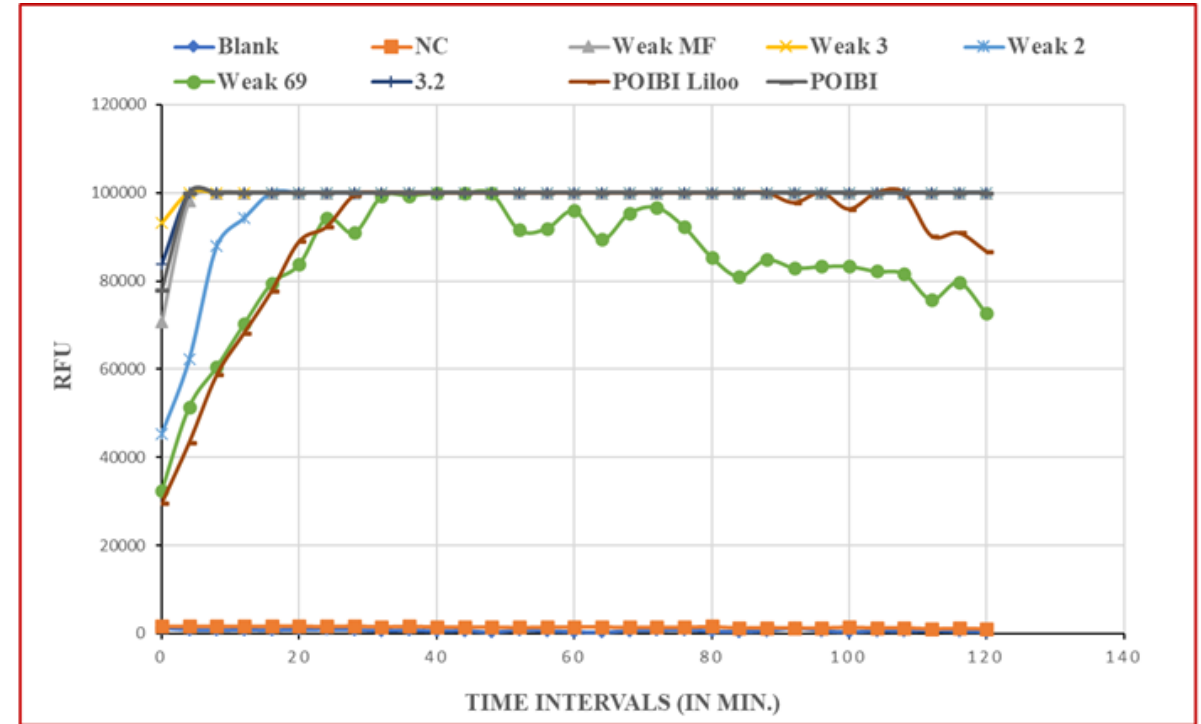
# Supersensitive and Rapid Detection of XDP from Infected Cherry Tree Samples Using RPA/Cas12a Method

XDP infected Cherry DNA sample



NC: Negative control (non-infected sample)

The max fluorescence reading is 100000 RFU.



NC: Negative control (non-infected sample); Blank: Water.

The max fluorescence reading is 100000 RFU.



# Conclusions and Perspectives

RPA/Cas12a assay is particularly appealing for early detection of low titer plant pathogens such as *Candidatus Liberibacter asiaticus* and phytoplasmas in asymptomatic tissues with the following advantages.

- ✓ **Supersensitivity** at the attomolar level ( $10^{-18}$  or 0.6 copy DNA per  $\mu\text{l}$ )
- ✓ **High specificity** at the single nucleotide level
- ✓ **Isothermal** RPA and Cas12a reactions at 37- 42 °C
- ✓ **Rapid** assays within 1 or 2 hr
- ✓ **High throughput** with a 384 fluorescence microplate reader or real-time PCR system
- ✓ **Field deployable** with a portable Genie III, lateral flow immunostrip or colorimetric assay
- **One-pot assay** to simplify the detection protocol and reduce potential contamination
- **Amplification-free methods**
- **Microfluidics, electrochemical sensors and mobile phone platforms**

# Acknowledgments

**Penn State University:** Matthew Wheatley, Sydney Ostlund, Qin Wang

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**Washington State University:** Youfu Zhao, Scott Harper

